
BioMax Environmental

Environmental Consulting and Industrial Hygiene Services

November 18th, 2008

Mr. Doug Button
Deputy Director
Real Estate Services Division
707 Third Street - 8th Floor
West Sacramento, CA 95605

**Microbial Assessment and
Mitigation Procedures for 5th and 6th Floor Storage and Electrical Room Areas
Department of General Services Board of Equalization Building
450 N. Street
Sacramento, California**

Dear Mr. Button,

BioMax Environmental, LLC (BioMax) is pleased to provide the Department of General Services (DGS) with this letter summary report detailing BioMax's findings and recommendations pertaining to our inspection and microbial sampling assessment services provided within the 5th and 6th Floor Storage and Electrical Room of your 450 N Street Building (subject building) located in Sacramento, California. BioMax understands that these microbial inspection and sampling assessment services were contracted with BioMax in an effort to evaluate the recently discovered visible moisture damage and potential microbial growth identified within the noted areas. According to information provided to BioMax from DGS personnel, such areas were identified by LaCroix Davis consultants (LCD) during the performance of their floor-by-floor forensic inspection within the BOE building. Such information was provided as a "list" of areas which DGS directed BioMax to, simply, investigate further.

Hence, these microbial inspection and assessment services are intended to provide a visual assessment as well as obtain analytical sampling and physical data pertaining to the current environmental conditions present within the affected interior areas identified. Site access was provided on October 29th and November 5th, 2008 by site representatives. On these days, Mr. Michael A. Polkabra, CIH, REA of BioMax performed a site inspection and sampling assessment within and adjacent to the areas of concern identified by LCD and DGS representatives. Based on current information provided and our visual observations gathered at this time, BioMax collected a series of airborne and surface microbial samples within and surrounding the areas of concern and representative affected materials so as to evaluate and assess the current environmental microbial conditions within and surrounding the impacted areas at this time.

SITE OBSERVATIONS

On-site inspection and sampling assessment activities were performed by Mr. Michael A. Polkabla, CIH, REA, of BioMax in accordance with currently recognized microbial assessment and sampling guideline procedures. Mr. Polkabla has been certified in the Comprehensive Practice of Industrial Hygiene by the American Board of Industrial Hygiene and holds the right to the designation "Certified Industrial Hygienist" (CIH) under certification number CP 7104. Mr. Polkabla is also certified by the California Environmental Protection Agency (Cal/EPA) as a Class I Registered Environmental Assessor (REA) under Cal/EPA certification number 05011. A summary of significant notations and observations gathered during BioMax's site inspection and assessment of the subject areas are compiled as follows:

1. At the time of our preliminary site inspection performed on October 29th, 2008 interior areas environmental conditions within the subject area consisted of a temperature range between 60 degrees F to 75 degrees F with a relative humidity range from 27 to 36%. Ambient outdoor conditions both prior to and following our interior assessment consisted of mild sunny conditions with predominant winds noted at approximately 0-5 knots from the northwesterly direction. Outdoor temperatures ranged between 72 to 75 degrees and relative humidity range of 27 to 29 %, respectively.
2. At the time of our follow-up site inspection performed on November 5th, 2008 interior area environmental conditions within the subject area consisted of an indicated temperature within the storage room of 72 degrees F with a relative humidity range from 28 to 30%. Ambient outdoor conditions both prior to and following our interior assessment consisted of cool and mild conditions with predominant winds noted at approximately 0-5 knots from the northwesterly direction. Outdoor temperatures ranged between 60 to 61 degrees and relative humidity range of 38 to 40%, respectively.
3. Site observations noted within the subject areas are as follows:

Storage Room 6C – At the time of our 10/29/08 assessment, storage room 6C was observed with excessive quantities of stored cubicle dividers and furniture which significantly limited physical access into the room. Upon inspection of accessible areas, however, BioMax verified the presence of dark colored "smudges" present on the upper wall to the left of the door entry point which had been indicated through interviews with LCD personnel. Although such "smudges" contained evidence of what appeared to be that of physical contact abrasion, BioMax elected to collect a surface sample and airborne SporeTrap sample for confirmatory microbial analysis as a precautionary measure. Utilization of a hand-held moisture meter also indicated normal moisture content present within the accessible structural areas at the time of our assessment.

Storage Room 6B - At the time of our 10/29/08 visual assessment of storage room 6C, BioMax found no visual evidence of mold-like staining present on any of the interior

accessible surfaces at this time. An airborne sample was collected, however, as reported below. Utilization of a hand-held moisture meter also indicated normal moisture content present within the accessible structural areas at the time of our assessment.

Storage Room 5C – At the time of our 10/29/08 assessment, BioMax inspected the MAP Scanning Room with a shared wall located to the west and adjacent to the noted break room. No visual evidence of mold-like staining was observed present on the shared wall. Utilization of a hand-held moisture meter also indicated normal moisture content present within the wallboard material at the nearest approximate location to the impacted area at the time of our assessment. A single air sample was similarly collected from this accessible area for comparative purposes.

Storage Room 5B – At the time of our follow-up assessment performed on 11/05/08, BioMax identified the presence of mold-like surface staining near the floor level present behind a metal storage rack with shelved computer equipment and supplies. BioMax also noted the presence of what appeared to be “stuffed” paint on the wall to ceiling interface immediately above the mold like staining. Such areas, however, did not contain the presence of staining on sheetrock or paint underlayment surfaces inspected at the time of this assessment. Utilization of a hand-held moisture meter also indicated normal moisture content present within the accessible structural areas at the time of our assessment. A SporeTrap airborne sample was similarly collected from this tenant accessible area for comparative purposes.

5th Floor Electrical and Telephone Equipment Room – Upon entrance into the northern portion of the electrical and Telephone Equipment Room located on the eastern side of the 5th floor, BioMax noted the significant reduction in interior temperature as compared to the adjacent occupied hallway area. Information provided to BioMax from DGS has indicated that such temperature conditions were mechanically maintained within this localized area due to the sensitive nature requirements of the internal electrical and telephone equipment therein. Inspection of the southern wall within the noted area revealed the presence of “fuzzy” black surface deposits present on the unpainted and exposed wallboard materials covering over 25 square feet of surface area at the time of our assessment. BioMax collected a surface sample of these noted “fuzzy” deposits as well as an interior SporeTrap sample for microbial analysis as indicated below. Utilization of a hand-held moisture meter also indicated normal moisture content present within the accessible structural areas at the time of our assessment.

4. As noted above, BioMax collected a series of airborne and surface samples from representative impacted building materials surfaces and areas located within and surrounding the visibly affected interior materials noted above. A series of digital images were also collected during BioMax’s inspection and sampling assessment activities. Images are attached to this summary report for further reference, as necessary. A detailed site map sketch indicating the extent of visibly affected areas noted at the time of this assessment and relative surface sampling locations may also be provided for further reference upon request.

5. In an effort to further evaluate the potential for exposure to airborne spores and/or spore deposition resultant from airborne transmission from the noted areas, BioMax also collected an airborne SporeTrap samples within the adjacent immediate work area as noted within Table 1 below. Such supplemental samples were collected from accessible surfaces during normal daily building ventilation conditions and routine employee operations and activities so as to model representative conditions within the working environment.

SAMPLING PROCEDURES

On-site inspection and sampling assessment activities were conducted by Mr. Michael A. Polkabl, CIH, REA, of BioMax Environmental. All sampling equipment, supplies, calibration materials, and collection media were provided by BioMax as part of the performance of this scope of work. Sample collection procedures and methods were performed using aseptic sampling methods following techniques prescribed by the contracted analytical laboratory.

SporeTrap Airborne Sampling:

The collection of airborne Spore Trap microbial samples was achieved using Zefon Air-O-Cell sampling cassette collection devices placed in each of the areas identified in Table 1. Airborne Spore Trap samples were collected within and outside the areas of concern at a height of approximately four feet above ground level using a tripod mounted Quick Take 15 air sampling pump manufactured by SKC International. Samples were collected at a calibrated flow rate of 15 liters per minute for a total of five minutes per sample. Resultant total sample volumes, therefore, corresponded to 75 liters collected for each sample. Field calibration of the SKC air sampling pump was conducted and recorded prior to and following sampling activities using a Bios Drycal primary standard flow meter and field rotometer. All spore trap air sampling and analytical procedures were performed in accordance with prescribed manufacturer guidelines as well as applicable professional certified industrial hygiene indoor air quality microbial investigation procedures and certified industrial hygiene practices.

Additional exterior samples were also similarly collected and analyzed during the collection of interior samples in an effort to evaluate and quantify typical background microbial taxa (types), rank order, and corresponding airborne spore levels present during the time of this assessment. Efforts were made in the collection of airborne samples to capture such samples during representative building occupancy conditions and activities so as to closely approximate normal air handling system ventilation conditions within each of the subject areas located within and surrounding the evaluated areas of concern. Sampling collection activities performed during this assessment included the collection of identifiable airborne microbial contaminants within the representative areas noted in Table 1 below:

Table 1. Airborne Spore Trap Sampling Locations:

Air Sample Number	Sampling Date	Spore Trap Air Sampling Location
14226819	10-29-08	Ambient Garage Rooftop
14226831	10-29-08	Storage room 6C
14226764	10-29-08	Storage Room 6B
13856254	10-29-08	Storage Room 5C
14226797	10-29-08	5 th Floor Electrical and Telephone Equipment Room
14226741	10-29-08	Ambient Floor Level Main Entry
14226828	11-05-08	Ambient Garage Rooftop
14226844	11-05-08	Storage Room 5B
14226634	11-05-08	Hallway near Storage Room 5B
14354900	11-05-08	Ambient Garage Rooftop

At the conclusion of sampling activities, preparation and shipping of the collected airborne samples were accomplished in accordance with standard industrial hygiene chain of custody (COC) documentation procedures and quality assurance/quality control practices. Once collected, labeled, and recorded, all samples were double sealed within airtight plastic Ziploc shipping containers and transported via Federal Express Priority Mail to Environmental Microbial Laboratories (EMLabs) in San Bruno, California. EMLabs holds current applicable analytical accreditation and specializes in microbial analytical procedures. Sampling and chain of custody records are provided as an attachment to this letter report for further reference.

BioTape Surface Sampling:

During our site inspection and sampling assessment activities, representative surface material samples were collected from interior areas and materials of concern noted within in Table 2 below. All surface samples were collected using "same-lot" BioTape collection media prepared and supplied by SKC International in accordance with manufacturers sampling guidelines as well as applicable professional certified industrial hygiene microbial sampling practices. Disposable gloves utilized during sample collection and changed between each sample.

Written sampling procedural guidance material prepared by the analytical laboratory and/or sample media manufacturer may also be provided upon request. A summary of surface material sampling locations are provided in Table 2. Specific sample locations may also be referenced within the digital image attachment, as necessary.

Table 2. Bulk Material and BioTape Surface Sample Locations:

Sample Number	Sampling Date	Material Sampling Location
S01	10-29-08	Storage Room 6C black "smudge" staining on wall to left of doorway entry
S02	10-29-08	Storage Room 5C black mold-like staining present behind metal framed storage shelves and noted computer equipment.
S03	10-29-08	5 th Floor Electrical and Telephone Equipment Room – Fuzzy black deposits on exposed unfinished/painted wallboard surface
S01	11-05-08	Storage Room 5B black mold-like staining present behind metal framed storage shelves at 6 inches afl.
S02	11-05-08	Duplicate sample within Storage Room 5B black mold-like staining present behind metal framed storage shelves.

Following sample collection, bulk material and surface samples were subsequently labeled and placed within individual plastic Ziploc storage bags for transportation via Federal Express Priority Mail to the analytical laboratory noted below. Preparation and shipping of the collected samples were accomplished in accordance with standard industrial hygiene chain of custody (COC) documentation procedures and quality assurance/quality control QA/QC practices. Once collected, labeled, and recorded, the samples were double sealed within airtight plastic Ziploc bag containers and similarly transported via Federal Express Priority Mail to Environmental Microbial Laboratories (EMLabs) of San Bruno, California. Sampling and chain of custody records are provided as an attachment to this letter report for further reference.

ANALYTICAL FINDINGS AND CONCLUSIONS

Airborne Spore Trap Sample Findings:

Laboratory analytical methods for the identification and enumeration of microbial (mold) taxa contaminants were conducted in accordance with prescribed analytical procedures and quality control/assurance measures. Original laboratory results including the enumeration of recognizable microbial spore types are also attached to this letter report for further detail. Analytical comments provided by the microbial laboratory regarding relative background debris and particulate levels are noted as a semi-quantitative assessment based on analyst interpretation and historical regional data. A summary of airborne Spore Trap microbial (mold) and particulate findings pertaining to each of the subject areas are presented in Table 3 below:

Table 3. Summary of Airborne Microbial Findings

Location Desc.	Total Mold Spores (Cts/m ³)	Background Debris (scale of 1-4)	Skin Cell Fragments (scale of 1-4)	Hyphal Fragments (units/M ³)
Ambient Garage Rooftop (10-29-08)	6,900	4+	1+	40
Storage Room 6C (10-29-08)	67	2+	1+	<13
Storage Room 6B (10-29-08)	250	3+	3+	<13
Storage Room 5C (10-29-08)	120	2+	2+	<13
5 th Floor Electrical and Telephone Equipment Room (10-29-08)	53	2+	1+	<13
Ambient Floor Level Main Entry (10-29-08)	4,800	4+	<1+	170
Ambient Garage Rooftop (11-05-08)	8,700	2+	<1+	40
Storage Room 5B (11-05-08)	160	2+	2+	<13
Hallway near Storage Room 5B (11-05-08)	270	2+	1+	13
Ambient Garage Rooftop (11-05-08)	5,300	1+	<1+	<13

The analytical findings presented in Table 3 indicate the presence of significantly lower concentrations of total microbial (mold) spores measured within each of the interior samples collected both within and surrounding the subject storage room and electrical room areas when compared to the levels currently measured within the samples collected from the corresponding ambient outside environment. Analytical findings also indicate similar fungal taxa distribution (mold types) and rank order (predominant taxa) of molds identified within the noted interior areas as well as the adjacent hallway areas sampled (area noted as "Hallway") when compared to the corresponding levels found within the ambient outside environmental samples. Data also

indicated the absence of elevated levels of "moisture indicator" mold taxa such as *Stachybotrys*, *Penicillium/Aspergillus*, *Chaetomium*, etc. within any of the noted interior areas. Analysis of fungal hyphal fragments (vegetative fungal growth structures) indicated fewer structures within the interior areas and adjacent interior spaces when compared to the corresponding levels found within the ambient outside environmental samples.

Although there are currently no regulatory standards or limits pertaining to allowable airborne fungal concentrations (for any mold taxa) present in indoor environments, there is a general consensus among indoor air quality experts that microbial contamination found within "typical healthy" working and living spaces are generally similar in kind and present at levels which are below those found in the corresponding native outside environment. BioMax believes that the absence of elevated levels of total airborne mold, absence of elevated hyphal structures, and typical taxa and rank order distribution are consistent with these generally acceptable conditions. BioMax, therefore, believes that these findings provide reasonable evidence indicating that current airborne microbial conditions present within the noted storage rooms and electrical and telephone equipment room areas are consistent with normal representative levels.

Therefore, BioMax believes that verification of such current airborne microbial conditions under the conditions sampled may be considered acceptable for normal BOE use of the storage room areas with precluded physical access of the visibly stained material surfaces at this time. Such limited access from contact with the visibly stained surfaces should be maintained (through plastic barrier and signage) until appropriate mitigative measures have been performed and successfully completed thorough verified post mitigation "clearance" sampling as recommended below.

Airborne Particulate Debris Findings:

Analytical findings of relatively low levels of airborne particulates debris identified within the interior collected air samples provide reasonable evidence indicating that current interior environmental conditions may be deemed unremarkable at this time.

Surface Sample Findings:

Laboratory analytical methods for the identification and enumeration of microbial taxa were conducted in accordance with prescribed analytical procedures and quality control/assurance measures. Laboratory analytical methods for the identification and enumeration of microbial fungal contaminants within the collected surface material samples were achieved through direct microscopic analysis using bright field microscopy.

Original laboratory results including the identification of recognizable microbial taxa are provided as an attachment to this letter report for further reference. Sampling and chain of custody records are provided as an attachment to this report for further reference. A summary of analytical findings pertaining to the collected surface samples are presented in Table 4 below:

Table 4. Summary of Surface Findings:

Sample Number	Sample Material and Location	Mold Genera Identified Present
S01 10-29-08	Storage Room 6C black "smudge" staining on wall to left of doorway entry	No Mold Spores Detected
S02 10-29-08	Storage Room 5C black mold-like staining present behind metal framed storage shelves and noted computer equipment.	Penicillium/Aspergillus-types (2,210 counts)
S03 10-29-08	5 th Floor Electrical and Telephone Equipment Room – Fuzzy black deposits on exposed unfinished/painted wallboard surface	Cladosporium (106 Counts) Other colorless (1 count)
S01 11-05-08	Storage Room 5B black mold-like staining present behind metal framed storage shelves at 6 inches afl.	Penicillium/Aspergillus-types (48 Counts) Ulocladium (264 Counts)
S02 11-05-08	Duplicate sample within Storage Room 5B black mold-like staining present behind metal framed storage shelves.	Penicillium/Aspergillus-types (430 Counts) Ulocladium (270 Counts)

Noted relative levels should be used for comparative purposes only and are not intended to establish "acceptable" indoor levels/conditions.

Analytical findings as presented in Table 4 above clearly indicated the presence of unique microbial fragments (spores) present in the majority of the materials sampled where mold-like staining was noted. The identified hydrophilic (moisture loving) mold taxa, such as Penicillium/Aspergillus and Ulocladium identified within the visibly "stained" surfaces sampled, represent what BioMax believes to be likely indicative of mold damage and growth likely resultant from historic incidents.

Although there are currently no regulatory standards or limits pertaining to allowable surface fungal concentrations (for any mold taxa) present on interior working environment surfaces, there is a general consensus among indoor air quality and microbial experts that significant visible microbial contamination found within occupied space building materials should be treated, removed, and/or otherwise minimized wherever practicable. Hence, BioMax believes that the findings detailed in this report warrant the implementation of the recommended precautions, continued area controls, and the performance of mitigative measures pertaining to the areas of identified visible microbial contamination as noted below.

RECOMMENDATIONS

Based on our preliminary observations within the subject areas and review of current analytical findings available at this time, BioMax recommends that the following corrective measures and mitigative actions be considered as follows:

Electrical and Telephone Equipment Room:

BioMax believes that the identification of relatively minimal levels of the ubiquitous mold taxa, *Cladosporium*, found present within the "fuzzy" surface debris is likely the result of particle deposition and accumulation resultant from electrostatic precipitation onto the noted wall surfaces within the electrical and telephone equipment room. BioMax believes that such accumulation is likely resultant from the observed cold (air conditioned) air currents traveling over the interior area's wallboard materials (of relative warmer temperature) which results in static charge and isometric dew point differential within the localized surfaces in the noted electrical and telephone equipment room. Such conditions, therefore, commonly result in the accumulation of particulate debris (including some mold spores) on the affected surfaces similar to those noted. Hence, it is BioMax's professional opinion that the physical removal of such loose material deposits present on the noted interior wall surfaces may be performed by the site mitigation contractor, JLS, utilizing HEPA filtered vacuum equipment as an additional precautionary measure. Due to the "unremarkable" verified microbial nature of the surface deposits and absence of elevated airborne mold spores as indicated by the airborne analytical findings, such activities can be achieved without the need for supplemental negative pressure containment barriers utilizing standard debris removal practices and procedures.

Storage Rooms 5B and 5C:

1. Due to the confirmed findings of elevated microbial contamination present within the sampled building material surfaces within the noted storage rooms, BioMax recommends that additional deconstructive inspection and appropriate mitigation the affected interior structures, walls, and wall cavities within the subject areas be performed as noted below. The purpose of these activities should be to adequately assess and evaluate the full extent of all moisture intrusion and microbial damages within each of the noted areas under appropriate microbial mitigative protective containment systems.
2. AS previously indicated, the airborne sampling data associated with storage Rooms 5B and 5C has indicated the presence of "unremarkable" mold spore levels which are consistent with normal interior levels and types. However, as an interim precautionary measure, BioMax recommends that the noted storage room surfaces containing the verified presence of mold growth be sealed/covered behind plastic barriers and posted with appropriate warnings to preclude physical contact and/or disturbance. In performing such mitigative measures, BioMax recommends that a qualified and experienced microbial abatement contractor be selected to construct and maintain such barrier protective systems.

3. Upon appropriate DGS authorization, BioMax recommends that BOE be provided with access to the storage areas to remove all remaining critical supplies and materials from the unaffected storage room area prior to the initiation of destructive inspection and forthcoming mitigative activities. Such supplemental destructive inspection and mitigative activities should only be performed within the noted storage rooms wherein critical containment barriers are established and maintained within each affected storage room area. The selected contractor must be specifically trained in the field of microbial abatement techniques and methods as well as maintain demonstrated proficiency in the establishment and use of appropriate barriers, personal protective equipment, abatement techniques and methods in the removal and decontamination of microbial affected and impacted materials.
4. Due to the current occupancy and client use within the building floor areas adjacent to the affected storage rooms, as a precautionary measure, BioMax recommends that the tenant be precluded from access into the break room once mitigative activities begin until the areas have been appropriately mitigated and verified as acceptable for reuse. The mitigation contractor should be directed to install a fully enclosed negative pressure environmental containment barrier encompassing the storage room area and affected materials during removal, inspection, and forthcoming treatment. These containment systems shall be designed for the purposes of containing and controlling possible fugitive emissions of airborne fungal spore contaminants during all forthcoming deconstruction, inspection, and mitigative activities. All critical containment systems shall be constructed of plastic and/or otherwise airtight materials so as to create a negative pressure system within the noted areas of concern. Due to physical constraints, all negative air pressure shall be maintained within the critical areas with the use of a High Efficiency Particulate Aerosol (HEPA) filtered "negative air machine" vented to the outside workspace environment. An adequate supply of filtered intake air shall also be established to allow an adequate supply of "clean" filtered make-up air into the critical containment. Wherever possible, clear translucent plastic observation windows shall be placed on the critical containment barrier within direct sight of the affected areas for the purposes of inspection during the performance of prescribed mitigative measures. BioMax is prepared to provide your selected contractor with additional and ongoing detail pertaining to the establishment maintenance, and specific locations of critical containment barriers, as necessary. Once, containment parameters have been established, the site contractor shall maintain an "as built" record of exact containment locations and materials for further review and reference.
5. As an additional precautionary measure, mitigative work shall be scheduled during non tenant-occupied hours wherein HEPA filtered air scrubbing equipment units will be operated in the hallway outside the containment area for the duration of such mitigative activities.
6. A series of similar plastic and/or otherwise impermeable zippered entry chambers shall be erected at the entrance of the containment systems for the purpose of establishing worker entrance/exit and clean personal protective equipment donning and decontamination area. HEPA filtered vacuum equipment capable of the effective removal of particulate contaminants from tools and personal protective equipment shall be placed within each of the zippered chambers closest to the working area. During such measures, appropriate signage

and warnings must be posted on the exterior of containment entrances to preclude uninformed access from unauthorized personnel and/or BOE staff. Data logging monitoring equipment employed to record pressure differentials on a 24-hour basis shall be used for the duration of functional barrier use.

7. Upon establishment of critical containment barriers, BioMax recommends that the selected microbial abatement contractor also places and maintains appropriate HEPA filtered air-scrubbing units within the affected areas. All Heating Ventilation and Air Conditioning (HVAC) supply vents and ceiling or wall mounted recessed lighting/ fan penetrations within the containment systems shall be deactivated and covered within similar plastic barrier systems. All appropriate wall and ceiling penetrations present within the containment systems shall also be sealed and/or otherwise rendered airtight and inoperable so as to minimize unfiltered particulate intrusion into and out of the established containment systems. It is specifically recommended that the ceiling tile level materials be critically sealed from the working areas within each of the noted containment rooms so as to preclude fugitive emissions from exiting the noted containments. Any smoke detectors and/or fire suppression systems shall NOT be covered nor rendered inoperable within the subject building unless authorized to do so under the direction and supervision of personnel.
8. Workers engaged in mold remediation/mitigation activities must be adequately trained and equipped with properly selected personal protective equipment (PPE) including, at minimum, hooded Tyvek coveralls, air purifying full face respirators with N100 minimum HEPA filter rating or similar PAPR systems, nitrile or latex gloves, chemical resistant boots or boot covers, with taped joints. Site control zones shall be established with exclusion, contaminant reduction (decontamination), and support zones in accordance with published Environmental Protection Agency (EPA) and California Department of Occupational Safety and Health (Cal/OSHA) guidelines. BioMax would be happy in providing the selected contractor with further site-specific detail regarding PPE regimen and appropriate site control zones, as necessary.
9. BioMax recommends that all interior items and/or furnishings remaining within the storage room areas not removed by BOE prior to the establishment of containment, be physically cleaned and sealed in plastic bags by the mitigation contractor and segregated into a non-impacted storage location. All remaining hard surface shelving and furnishings present within the storage room containment systems shall receive a thorough inspection, cleaning, mildicide wet-wiping, and HEPA vacuuming as part of these recommended procedures prior to subsequent clearance testing and reuse.
10. Following metal shelving removal, BioMax specifically recommends that all visibly damaged sheetrock materials and impacted sheetrock be removed and wallboard underlayment materials inspected for moisture related damages. As verified through visual inspection, any stained and/or moisture/mold affected interior sheetrock and building materials should be removed, wherever feasible, to the extent of visible staining, at a minimum. Any affected baseboard and floor tile materials shall also be removed and disposed under containment controls for appropriate inspection of subflooring at the direction of the project CIH.

Removal of moisture impacted and mold damaged materials may also employ the use of appropriate item-specific containment methods and systems (such as sealed plastic glove-bag containment systems, or equivalent) applicable to the materials being removed at the direction of the Project CIH. BioMax currently anticipates that all visually affected wallboard material exhibiting staining and "sluffing" paint, shall be removed for disposal, and physical inspection of wall cavities and underlayment, as necessary. Any underlayment materials exhibiting visible signs of moisture staining shall also be removed and/or decontaminated, at the direction of the Project CIH, as deemed necessary.

11. Other potentially affected areas and building materials encountered during these deconstructive and investigative stages, such as adjacent wall materials, floor materials, ceiling materials, and underlayment building material framing, etc., must be thoroughly inspected during these deconstructive stages to identify the extent of any additional microbial related materials and water damage indicators. In general, all microbial impacted materials shall be removed to the extent of visible staining and at least 2 feet beyond such identified perimeters, wherever possible.
12. All remaining moisture/mold affected porous and non-porous building materials deemed infeasible for removal and/or disposal (due to structural integrity concerns) shall be inspected and receive a series of decontamination treatment measures designed to minimize and control the presence of microbial related contaminants. Decontamination methods employed shall, at a minimum, include treatment of all identified surfaces with a series of thorough chlorine based mildicide (minimum 10 parts water to 1 part chlorine soln.) applications followed by a series of thorough HEPA filtered vacuuming procedures using power sanding and/or brush agitation. The duration and frequency of mildicide and HEPA sanding/brushing applications employed may vary depending on local material contamination but shall be sufficient in removing and decontaminating all visible surface staining to levels deemed by BioMax to be consistent with representative background levels. Reasonable additional mitigative measures and controls may be required, as necessary, upon discovery of additional contaminated materials as well as BioMax's site inspection findings and observations performed during this scope of work. BioMax will be available to provide ongoing consultation with the contractor pertaining to these measures and site/material specific decontamination measures upon request.
13. Upon completion of mitigation efforts performed by the selected mitigation contractor, BioMax recommends the performance of a visual inspection conducted by the Project CIH to verify that all significant mold related staining and moisture indicators have been removed and/or treated and that all prescribed mitigative efforts and measures have been appropriately achieved. Once established, the Project CIH will collect a series of microbial "clearance" air samples to verify that all affected interior areas have been appropriately decontaminated to acceptable background airborne levels and that the affected areas within the subject building are verified as "cleared" for reconstruction, forthcoming reoccupancy, and reuse. Such Post Mitigative "clearance" evaluation criteria have been developed in BioMax's February 15th, 2008 letter report titled Post Mitigation Clearance Assessment Protocols and previously approved by HygieneTech, Inc. (HTI) in their approval letter dated February 22nd, 2008.

Additional "punch-list" action items may be provided to the contractor following the performance of this site clearance inspection prior to receipt of analytical results, as deemed necessary.

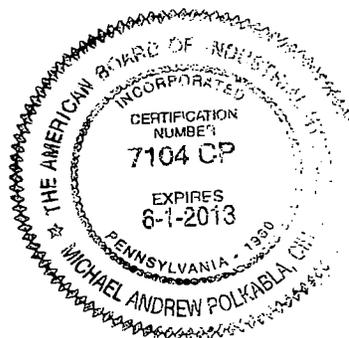
14. Upon review of analytical sampling results by the Project CIH and achievement of acceptable post mitigative clearance criteria, BioMax recommends that DGS directs the mitigation/reconstruction contractor apply a mildicide-based sealant onto all remaining organic-based building materials and previously treated surfaces. Use of a recognized commercially available sealant product with microbial growth inhibitors in accordance with manufacturer's application and use instructions is believed to be currently acceptable for these purposes. The provision of appropriate access shall be provided to BOE and its consultants for inspection of affected areas and materials prior to final encapsulation and reconstruction upon request.
15. Following the performance of these mitigative measures, the designated site reconstruction contractor is strongly encouraged to verify that repairs to any faulty and/or deficient building penetration, drainage, plumbing and/or building envelop sealing systems have been appropriately inspected, replaced/repared, and function tested prior to the reconstruction of the affected interior structures and cavities. Certainly, the repair/replacement and/or establishment of any such additional engineering controls (as recommended through additional professional consultation) must be performed and implemented in accordance with applicable standards, building codes, and ordinances, as necessary.
16. Upon completion, reconstruction of interior structural materials should be undertaken utilizing visibly clean (hand selected) construction grade materials in accordance with applicable building codes and requirements. The reconstruction contractor shall be required to only select materials which are obtained from reputable commercial sources and which are believed and visually verified to be free from elevated microbial contamination and/or elevated moisture content. New building materials, which are notably moist and/or visibly stained, shall NOT be used during the reconstruction of the subject structure. BioMax specifically recommends that reconstruction materials selected for use in the storage areas be specifically selected based on their moisture deterrent and anti-microbial properties wherever feasible.
17. Reasonable additional assessment and mitigative measures may also be required upon the identification of new or previously undiscovered materials and/or information related to moisture/microbial impacts, as necessary. Any reoccurrence of moisture intrusion following reconstruction should certainly be reviewed and addressed through further professional consultation, as necessary. BioMax would be happy to provide additional microbial consultative services pertaining to the mitigation of such structures so as to minimize potential adverse impacts to the interior working environment during the performance of any such activities upon request..

Once again, it has been a pleasure working with DGS on these important matters. If you have any additional questions, comments, or require further assistance, please do not hesitate to contact me directly at (510) 724-3100.

Sincerely,



Michael A. Polkabila, CIH, REA
Vice President, Principal



LIMITATIONS

Please note that the professional opinions presented in this review are intended for the sole use of DGS and their designated beneficiaries. No other party should rely on the information contained herein without the prior written consent of BioMax Environmental and DGS. The professional opinions provided herein are based on BioMax's review and understanding of current site information and observed site conditions present within the areas inspected at the time these services were performed. Professional recommendations provided as part of this limited scope of work are intended for client consideration only and are not intended as a professional or regulatory mandate. Implementation of any of the above measures or recommendations does not, in any way, warrant the day-to-day health and/or safety of building occupants, residents, site workers, nor regulatory or building code compliance status during normal and changing environmental conditions. As microbial contamination, by nature, may change over time due to additional moisture intrusion, favorable growth conditions, and changing environments, the findings of this report are subject to change in the event that such conditions and/or environments arise. Also, the professional opinions expressed here are subject to revision in the event that new or previously undiscovered information is obtained or uncovered.

It is expressly noted that all risk management and safety related decisions are criteria dependent and situation specific requiring extensive knowledge and value assessments to be properly determined and implemented by competent professionals. Hence, the recommendations provided and contained in this and any other applicable report communication is intended for consideration purposes only. It is not intended, nor should it be construed as providing legal advice or warranting any level of safety or regulatory compliance. The sole purpose of such information is to assist with the identification, evaluation and control of observed or measured levels of contamination resultant from physical, chemical, and/or biological hazards. Any action taken based on this information, including but not limited to opinions, suggestions and recommendations, whether implied or expressed, is the sole responsibility of the individual taking the action.

These services were performed by BioMax in accordance with generally accepted professional industrial hygiene principals, practices, and standards of care. Under the existing Industrial Hygiene Definition and Registration Act, all reports, opinions or official documents prepared by a Certified Industrial Hygienist (CIH) constitutes an expression of professional opinion regarding those facts or findings which are subject of a certification and does not constitute a warranty or guarantee, either expressed or implied.

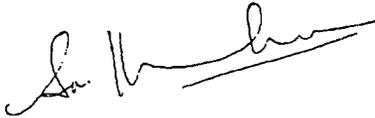
EMLab P&K

Report for:

Mr. Michael Polkabila
Biomax Environmental
775 San Pablo Ave.
Pinole, CA 94564

Regarding: Project: 102908-01; DGS Building, 2014 BR, 6th Floor, 5th Floor
EML ID: 483560

Approved by:



Lab Manager
Dr. Kamashwaran Ramanathan

Dates of Analysis:
Spore trap analysis: 10-31-2008

Project SOPs: Spore trap analysis (I100000)

This coversheet is included with your report in order to comply with AIHA and ISO accreditation requirements.

For clarity, we report the number of significant digits as calculated; but, due to the nature of this type of biological data, the number of significant digits that is used for interpretation should generally be one or two. All samples were received in acceptable condition unless noted in the Report Comments portion in the body of the report. Due to the nature of the analyses performed, field blank corrections of results is not a standard practice. The results relate only to the items tested.

EMLab P&K ("the Company") shall have no liability to the client or the client's customer with respect to decisions or recommendations made, actions taken or courses of conduct implemented by either the client or the client's customer as a result of or based upon the Test Results. In no event shall the Company be liable to the client with respect to the Test Results except for the Company's own willful misconduct or gross negligence nor shall the Company be liable for incidental or consequential damages or lost profits or revenues to the fullest extent such liability may be disclaimed by law, even if the Company has been advised of the possibility of such damages, lost profits or lost revenues. In no event shall the Company's liability with respect to the Test Results exceed the amount paid to the Company by the client therefor.

Document Number: 200091 - Revision Number: 5

EMLab P&K

1150 Bayhill Drive, Suite 100, San Bruno, CA 94066
(650) 829-5800 Fax (650) 829-5852 www.emlab.com

Client: Biomax Environmental
C/O: Mr. Michael Polkabla
Re: 102908-01; DGS Building, 2014 BR, 6th Floor,
5th Floor

Date of Sampling: 10-29-2008
Date of Receipt: 10-30-2008
Date of Report: 10-31-2008

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	14226819: Ambient, garage roof top		13856112: Occupied work area near 2014		14226836: Break room 2014, behind critical		14226831: Storage rm 6C	
Comments (see below)	None		None		None		None	
Lab ID-Version†:	2135133-1		2135134-1		2135135-1		2135136-1	
	raw ct.	spores/m3	raw ct.	spores/m3	raw ct.	spores/m3	raw ct.	spores/m3
Alternaria	1	13						
Arthrinium								
Ascospores*	4	440			1	53		
Aureobasidium								
Basidiospores*	21	2,300	1	53				
Bipolaris/Drechslera group	1	13			1	13		
Botrytis								
Chaetomium	3	40						
Cladosporium	18	2,000	1	53	3	160	1	53
Curvularia								
Epicoccum			1	13				
Fusarium								
Nigrospora	2	27						
Other brown	2	27			1	13	1	13
Penicillium/Aspergillus types†	17	1,900	2	110	2	110		
Pithomyces								
Rusts*								
Smuts*, Periconia, Myxomycetes*	6	80						
Stachybotrys								
Stemphylium								
Torula								
Ulocladium								
Background debris (1-4+)††	4+		3+		2+		2+	
Hyphal fragments/m3	40		< 13		< 13		< 13	
Pollen/m3	< 13		< 13		13		< 13	
Skin cells (1-4+)	< 1+		2+		2+		1+	
Sample volume (liters)	75		75		75		75	
§ TOTAL SPORE/m3		6,900		230		350		67

Comments:

* Most of these spore types are not seen with culturable methods (Andersen sampling), although some may appear as non-sporulating fungi.

Most of the basidiospores are "mushroom" spores while the rusts and smuts are plant pathogens.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

†† Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.

The Limit of Detection is the product of a raw count of 1 and 100 divided by the percent read. The analytical sensitivity (counts/m3) is the product of the Limit of Detection and 1000 divided by the sample volume.

‡ A "Version" greater than 1 indicates amended data.

§ Total Spores/m3 has been rounded to two significant figures to reflect analytical precision.

TestAmerica Environmental Microbiology Laboratory, Inc.

EMLab ID: 483560, Page 1 of 2

EMLab P&K

1150 Bayhill Drive, Suite 100, San Bruno, CA 94066
(650) 829-5800 Fax (650) 829-5852 www.emlab.com

Client: Biomax Environmental

C/O: Mr. Michael Polkabla

Re: 102908-01; DGS Building, 2014 BR, 6th Floor,
5th Floor

Date of Sampling: 10-29-2008

Date of Receipt: 10-30-2008

Date of Report: 10-31-2008

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	14226764: Storage rm 6B		13856254: Storage rm 5C		14226797: Elec/comp room East side		14226741: Ambient main entry	
Comments (see below)	None		None		None		None	
Lab ID-Version†:	2135137-1		2135138-1		2135139-1		2135140-1	
	raw ct.	spores/m3	raw ct.	spores/m3	raw ct.	spores/m3	raw ct.	spores/m3
Alternaria	1	13					2	27
Arthrinium								
Ascospores*							2	230
Aureobasidium								
Basidiospores*							7	770
Bipolaris/Drechslera group	1	13						
Botrytis								
Chaetomium								
Cladosporium	2	110	2	110	1	53	27	3,000
Curvularia								
Epicoccum								
Fusarium								
Myrothecium								
Nigrospora							1	13
Other brown								
Penicillium/Aspergillus types†	2	110					6	670
Pithomyces								
Rusts*	1	13	1	13				
Smuts*, Periconia, Myxomycetes*							4	53
Stachybotrys								
Stemphylium								
Torula								
Ulocladium								
Background debris (1-4+)††	3+		2+		2+		4+	
Hyphal fragments/m3	< 13		< 13		< 13		170	
Pollen/m3	< 13		< 13		< 13		13	
Skin cells (1-4+)	3+		2+		1+		< 1+	
Sample volume (liters)	75		75		75		75	
§ TOTAL SPORE/m3		250		120		53		4,800

Comments:

* Most of these spore types are not seen with culturable methods (Andersen sampling), although some may appear as non-sporulating fungi.

Most of the basidiospores are "mushroom" spores while the rusts and smuts are plant pathogens.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

†† Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.

The Limit of Detection is the product of a raw count of 1 and 100 divided by the percent read. The analytical sensitivity (counts/m3) is the product of the Limit of Detection and 1000 divided by the sample volume.

‡ A "Version" greater than 1 indicates amended data.

§ Total Spores/m3 has been rounded to two significant figures to reflect analytical precision.
TestAmerica Environmental Microbiology Laboratory, Inc.

EMLab P&K

1150 Bayhill Drive, Suite 100, San Bruno, CA 94066
(650) 829-5800 Fax (650) 829-5852 www.emlab.com

Client: Biomax Environmental
C/O: Mr. Michael Polkabra
Re: 102908-01; DGS Building, 2014 BR, 6th Floor,
5th Floor

Date of Sampling: 10-29-2008
Date of Receipt: 10-30-2008
Date of Report: 10-31-2008

MoldRANGE™: Extended Outdoor Comparison**Outdoor Location: 14226819, Ambient, garage roof top**

Fungi Identified	Outdoor data	Typical Outdoor Data by Date†				Typical Outdoor Data by Location‡			
		Month: October				State: CA			
	spores/m3	low	med	high	freq %	low	med	high	freq %
Generally able to grow indoors*									
Alternaria	13	7	40	450	62	7	27	210	58
Bipolaris/Drechslera group	13	7	13	210	23	7	13	120	13
Chaetomium	40	7	13	130	14	7	13	120	19
Cladosporium	2,000	53	800	12,000	97	53	640	6,400	98
Curvularia	-	7	27	710	27	7	13	210	7
Nigrospora	27	7	13	230	25	7	13	170	8
Other brown	27	7	13	120	35	7	13	80	37
Penicillium/Aspergillus types	1,900	27	270	3,400	86	38	210	2,500	87
Stachybotrys	-	7	13	350	3	7	13	290	5
Torula	-	7	13	200	12	7	13	150	12
Seldom found growing indoors**									
Ascospores	440	13	170	4,700	82	13	110	1,800	72
Basidiospores	2,300	27	500	21,000	96	13	210	6,700	94
Rusts	-	7	22	410	29	7	13	250	28
Smuts, Periconia, Myxomycetes	80	8	53	880	79	8	40	480	70
TOTAL SPORES/M3	6,840								

† The Typical Outdoor Data by Date represents the typical outdoor spore levels across North America for the month indicated. The last column represents the frequency of occurrence. The low, medium, and high values represent the 2.5, 50, and 97.5 percentile values of the spore type when it is detected. For example, if the frequency of occurrence is 63% and the low value is 53, it would mean that the given spore type is detected 63% of the time and, when detected, 2.5% of the time it is present in levels above the detection limit and below 53 spores/m³. These values are updated periodically, and if enough data is not available to make a statistically meaningful assessment, it is indicated with a dash.

‡ The Typical Outdoor Data by Location represents the typical outdoor spore levels for the region indicated for the entire year. As with the Typical Outdoor Data by Date, the four columns represent the frequency of occurrence and the typical low, medium, and high concentration values for the spore type indicated. These values are updated periodically, and if enough data is not available to make a statistically meaningful assessment, it is indicated with a dash.

*The spores in this category are generally capable of growing on wet building materials in addition to growing outdoors. Building related growth is dependent upon the fungal type, moisture level, type of material, and other factors. *Cladosporium* is one of the predominant spore types worldwide and is frequently present in high numbers. *Penicillium/Aspergillus* species colonize both outdoor and indoor wet surfaces rapidly and are very easily dispersed. Other genera are usually present in lesser numbers.

**These fungi are generally not found growing on wet building materials. For example, the rusts and smuts are obligate plant pathogens. However, in each group there are notable exceptions. For example, agents of wood decay are members of the basidiomycetes and high counts of a single morphological type of basidiospore on an inside sample should be considered significant.

Interpretation of the data contained in this report is left to the client or the persons who conducted the field work. This report is provided for informational and comparative purposes only and should not be relied upon for any other purpose. "Typical outdoor data" are based on the results of the analysis of samples delivered to and analyzed by EMLab P&K and assumptions regarding the origins of those samples. Sampling techniques, contaminants infecting samples, unrepresentative samples and other similar or dissimilar factors may affect these results. In addition, EMLab P&K may not have received and tested a representative number of samples for every region or time period. EMLab P&K hereby disclaims any liability for any and all direct, indirect, punitive, incidental, special or consequential damages arising out of the use or interpretation of the data contained in, or any actions taken or omitted in reliance upon, this report.

EMLab P&K

1150 Bayhill Drive, Suite 100, San Bruno, CA 94066
(650) 829-5800 Fax (650) 829-5852 www.emlab.com

Client: Biomax Environmental
C/O: Mr. Michael Polkabila
Re: 102908-01; DGS Building, 2014 BR, 6th Floor,
5th Floor

Date of Sampling: 10-29-2008
Date of Receipt: 10-30-2008
Date of Report: 10-31-2008

MoldRANGE™: Extended Outdoor Comparison**Outdoor Location: 14226741, Ambient main entry**

Fungi Identified	Outdoor data	Typical Outdoor Data by Date†				Typical Outdoor Data by Location‡			
		Month: October				State: CA			
		spores/m3	low	med	high	freq %	low	med	high
Generally able to grow indoors*									
Alternaria	27	7	40	450	62	7	27	210	58
Bipolaris/Drechslera group	-	7	13	210	23	7	13	120	13
Chaetomium	-	7	13	130	14	7	13	120	19
Cladosporium	3,000	53	800	12,000	97	53	640	6,400	98
Curvularia	-	7	27	710	27	7	13	210	7
Nigrospora	13	7	13	230	25	7	13	170	8
Other brown	-	7	13	120	35	7	13	80	37
Penicillium/Aspergillus types	670	27	270	3,400	86	38	210	2,500	87
Stachybotrys	-	7	13	350	3	7	13	290	5
Torula	-	7	13	200	12	7	13	150	12
Seldom found growing indoors**									
Ascospores	230	13	170	4,700	82	13	110	1,800	72
Basidiospores	770	27	500	21,000	96	13	210	6,700	94
Rusts	-	7	22	410	29	7	13	250	28
Smuts, Periconia, Myxomycetes	53	8	53	880	79	8	40	480	70
TOTAL SPORES/M3	4,763								

† The Typical Outdoor Data by Date represents the typical outdoor spore levels across North America for the month indicated. The last column represents the frequency of occurrence. The low, medium, and high values represent the 2.5, 50, and 97.5 percentile values of the spore type when it is detected. For example, if the frequency of occurrence is 63% and the low value is 53, it would mean that the given spore type is detected 63% of the time and, when detected, 2.5% of the time it is present in levels above the detection limit and below 53 spores/m³. These values are updated periodically, and if enough data is not available to make a statistically meaningful assessment, it is indicated with a dash.

‡ The Typical Outdoor Data by Location represents the typical outdoor spore levels for the region indicated for the entire year. As with the Typical Outdoor Data by Date, the four columns represent the frequency of occurrence and the typical low, medium, and high concentration values for the spore type indicated. These values are updated periodically, and if enough data is not available to make a statistically meaningful assessment, it is indicated with a dash.

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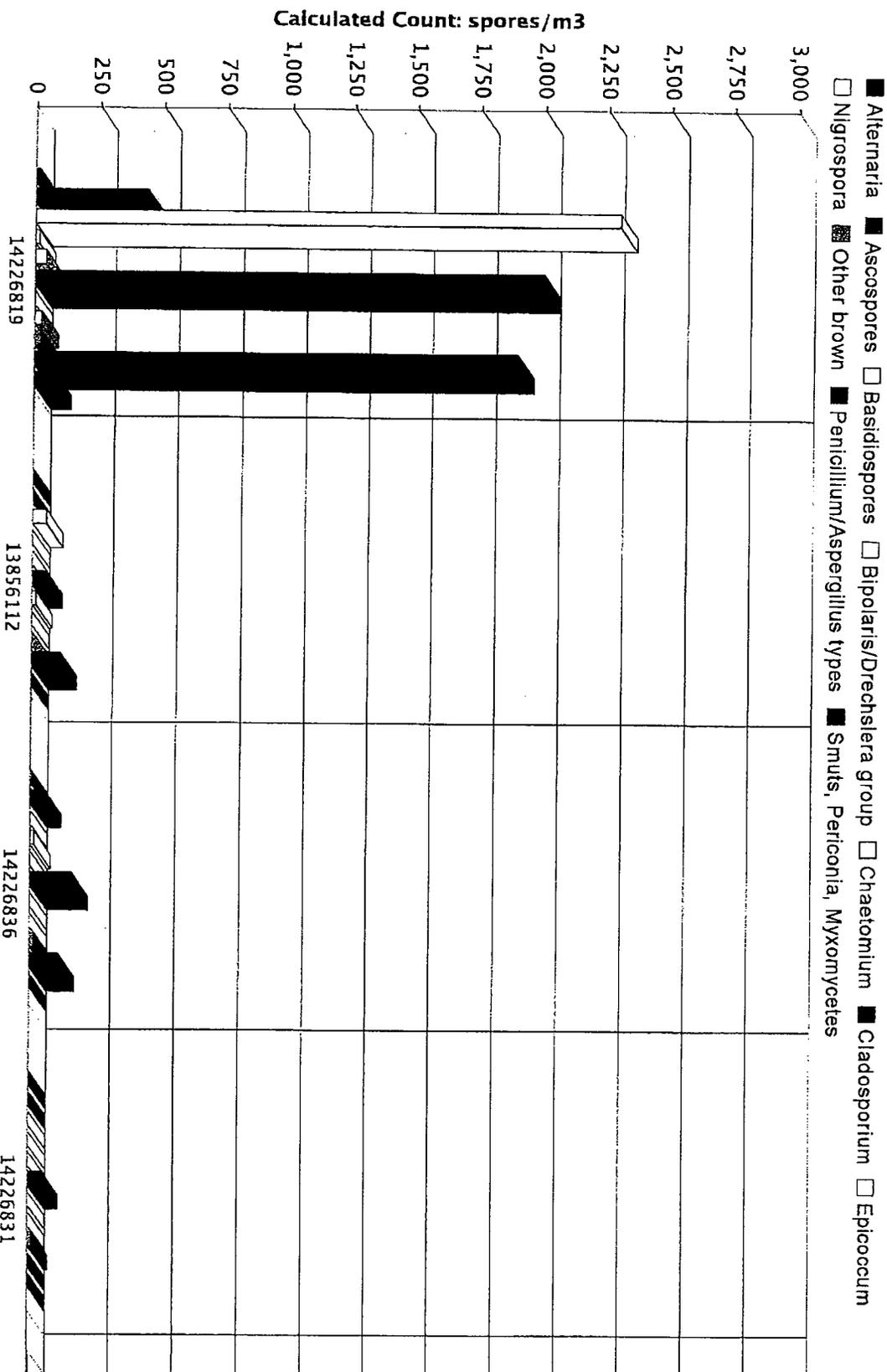
**These fungi are generally not found growing on wet building materials. For example, the rusts and smuts are obligate plant pathogens. However, in each group there are notable exceptions. For example, agents of wood decay are members of the basidiomycetes and high counts of a single morphological type of basidiospore on an inside sample should be considered significant.

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10-31-2008: 102908-01

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

EMLab P&K
 1150 Bayhill Drive, Suite 100, San Bruno, CA 94066
 (650) 829-5800 Fax (650) 829-5852 www.emlab.com



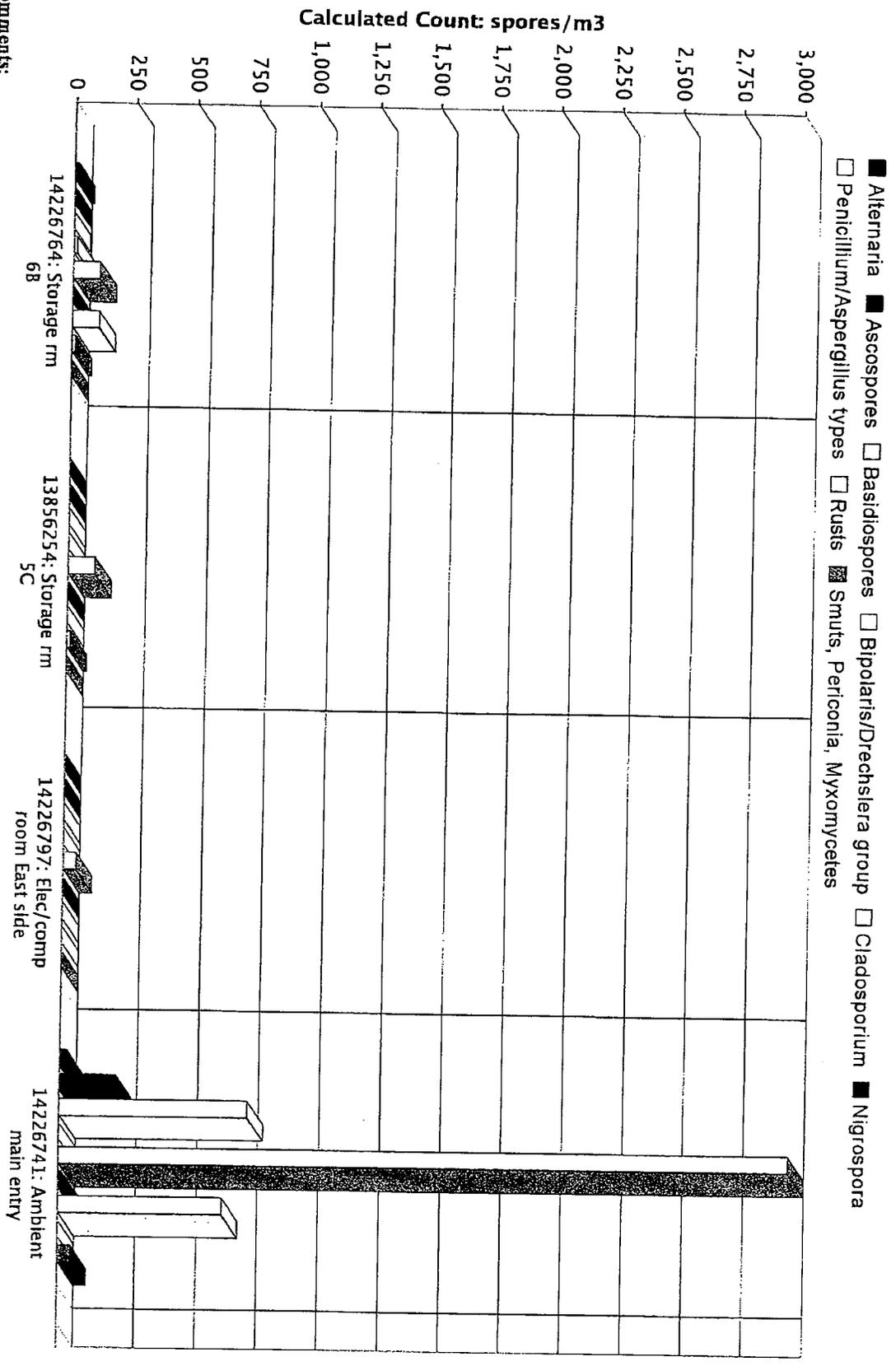
Comments:

Note: Graphical output may understate the importance of certain "marker" genera. TestAmerica Environmental Microbiology Laboratory, Inc.

10-31-2008: 102908-01

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

EMLab P&K
 1150 Bayhill Drive, Suite 100, San Bruno, CA 94066
 (650) 829-5800 Fax (650) 829-5852 www.emlab.com



Comments:

Note: Graphical output may understate the importance of certain "marker" genera. TestAmerica Environmental Microbiology Laboratory, Inc.

MICROBIAL SPORE TRAP AIR SAMPLING RECORD



000483560

Page 1 of 1

BioMax Environmental
775 San Pablo Ave.
Pinole, CA 94564

www.biomaxenvironmental.com

Phone: (510) 724-3100
Fax: (510) 724-3145
biomaxenv@aol.com

Location: <u>DGS Building</u> <u>2014 - BR</u> <u>6th Floor</u> <u>5th Floor</u>	Client: <u>DGS</u>
Date: <u>10/29/08</u>	Project #: <u>102908-01</u>
Collected by: <u>M.A. Poltack</u>	Laboratory: <u>EM Labs</u>
Signature: <u>[Signature]</u>	Req. Turn Around:
	Analysis (circle): <u>Fungal</u> <u>Particulate</u> AD/Quantification

Sample Number	Time	Location/Desc	Temp/RH
14226819	1120	Ambient - Garage Roof top	75° / 27%
13856112	1150	Occupied work area near 2014	75° / 28%
14226836	1205	Break Room 2014 (^{Behind} critical)	74° / 27%
14226831	1225	Storage Rm 6C	75 / 28
14226764	1240	Storage Rm 6B	72 / 29%
13856254	1255	Storage Rm 5C	73° / 28%
14226797	1320	Elec / Comp Room East Side	60° / 36%
14226741	1345	Ambient Main Entry	72° / 29%
Total Sample Time (min): <u>5</u>	Flow Rate (l/min): <u>15</u>	Total Sample Volume (liters): <u>75</u>	Ambient Conditions: <u>Clear / mild</u> <u>05 NW</u>
Comments:			

Please sign this form below acknowledging sample receipt and return executed form with laboratory reports. Fax, send, e-mail results to BioMax Environmental at (510) 724-3145 biomaxenv@aol.com
Other Instructions: _____

Relinquished by: <u>[Signature]</u>	Received By: <u>Ann Marisey</u>
Method of Transportation: <u>Fed Ex</u>	Time/Date Received: <u>10-30-08 9:15</u>
Time/Date Sent: <u>4:00 10/29/08</u>	

**EMLab P&K**

Report for:

Mr. Michael Polkabila
Biomax Environmental
775 San Pablo Ave.
Pinole, CA 94564

Regarding: Project: 110508-01; DGS
 EML ID: 485829

Approved by:

A handwritten signature in black ink, appearing to read "Dr. Kamashwaran Ramanathan".

Lab Manager
Dr. Kamashwaran Ramanathan

Dates of Analysis:

Quantitative spore count direct exam: 11-07-2008

Spore trap analysis: 11-07-2008

Spore trap analysis other particles-Supplement: 11-07-2008

Project SOPs: Quantitative spore count direct exam (I100006), Spore trap analysis (I100000), Spore trap analysis other particles-Supplement (100185)

This coversheet is included with your report in order to comply with AIHA and ISO accreditation requirements.

For clarity, we report the number of significant digits as calculated; but, due to the nature of this type of biological data, the number of significant digits that is used for interpretation should generally be one or two. All samples were received in acceptable condition unless noted in the Report Comments portion in the body of the report. Due to the nature of the analyses performed, field blank corrections of results is not a standard practice. The results relate only to the items tested.

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Document Number: 200091 - Revision Number: 5

EMLab P&K

1010 N Central Avenue, Glendale, CA 91202
(858) 569-5800 Fax (858) 569-5806 www.emlab.com

Client: Biomax Environmental
C/O: Mr. Michael Polkabila
Re: 110508-01; DGS

Date of Sampling: 11-05-2008
Date of Receipt: 11-06-2008
Date of Report: 11-07-2008

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	14226828: Ambient garage rooftop E		14226844: Room 5B storage		14226634: Hallway west 5B storage		14226789: Floor 22 east area	
Comments (see below)	None		None		None		None	
Lab ID-Version†:	2145311-1		2145312-1		2145313-1		2145314-1	
	raw ct.	spores/m3	raw ct.	spores/m3	raw ct.	spores/m3	raw ct.	spores/m3
Alternaria								
Arthrinium								
Ascospores*	4	170	1	13				
Aureobasidium								
Basidiospores*	62	3,100						
Bipolaris/Drechslera group								
Botrytis								
Chaetomium								
Cladosporium	98	5,200	1	53	4	210		
Curvularia								
Epicoccum	1	13						
Fusarium								
Myrothecium								
Nigrospora					1	13		
Other brown								
Other colorless								
Penicillium/Aspergillus types†	1	53						
Pithomyces								
Rusts*			2	27				
Smuts*, Periconia, Myxomycetes*	3	40						
Stachybotrys								
Stemphylium	1	13						
Torula								
Ulocladium			5	67	3	40		
Zygomycetes								
Background debris (1-4+)††	2+		2+		2+		1+	
Hyphal fragments/m3	40		< 13		13		< 13	
Pollen/m3	40		< 13		< 13		< 13	
Skin cells (1-4+)	< 1+		2+		1+		< 1+	
Sample volume (liters)	75		75		75		75	
§ TOTAL SPORE/m3		8,700		160		270		< 13

Comments:

* Most of these spore types are not seen with culturable methods (Andersen sampling), although some may appear as non-sporulating fungi. Most of the basidiospores are "mushroom" spores while the rusts and smuts are plant pathogens.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

†† Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.

The Limit of Detection is the product of a raw count of 1 and 100 divided by the percent read. The analytical sensitivity (counts/m3) is the product of the Limit of Detection and 1000 divided by the sample volume.

‡ A "Version" greater than 1 indicates amended data.

§ Total Spores/m3 has been rounded to two significant figures to reflect analytical precision.

TestAmerica Environmental Microbiology Laboratory, Inc.

EMLab ID: 485829, Page 1 of 2

EMLab P&K

1010 N Central Avenue, Glendale, CA 91202
 (858) 569-5800 Fax (858) 569-5806 www.emlab.com

Client: Biomax Environmental
 C/O: Mr. Michael Polkabila
 Re: 110508-01; DGS

Date of Sampling: 11-05-2008
 Date of Receipt: 11-06-2008
 Date of Report: 11-07-2008

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	14226790: Floor 22 north area		14226839: Floor 22 west		14226818: Floor 22 south area		14354900: Ambient garage roof-post	
Comments (see below)	None		None		None		None	
Lab ID-Version‡:	2145315-1		2145316-1		2145317-1		2145318-1	
	raw ct.	spores/m3	raw ct.	spores/m3	raw ct.	spores/m3	raw ct.	spores/m3
Alternaria								
Arthrimum								
Ascospores*							3	160
Aureobasidium								
Basidiospores*							77	4,000
Bipolaris/Drechslera group								
Botrytis								
Chaetomium								
Cladosporium							20	1,100
Curvularia								
Epicoccum								
Fusarium								
Myrothecium								
Nigrospora							6	80
Other brown					1	13	1	13
Other colorless								
Penicillium/Aspergillus types†								
Pithomyces								
Rusts*								
Smuts*, Periconia, Myxomycetes*							1	13
Stachybotrys								
Stemphylium								
Torula								
Ulocladium								
Zygomycetes								
Background debris (1-4+)††	2+		3+		> 4+		1+	
Hyphal fragments/m3	< 13		< 13		13		< 13	
Pollen/m3	< 13		< 13		< 13		< 13	
Skin cells (1-4+)	< 1+		< 1+		< 1+		< 1+	
Sample volume (liters)	75		75		75		75	
§ TOTAL SPORE/m3		< 13		< 13		13		5,300

Comments:

* Most of these spore types are not seen with culturable methods (Andersen sampling), although some may appear as non-sporulating fungi. Most of the basidiospores are "mushroom" spores while the rusts and smuts are plant pathogens.
 † The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.
 †† Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.
 The Limit of Detection is the product of a raw count of 1 and 100 divided by the percent read. The analytical sensitivity (counts/m3) is the product of the Limit of Detection and 1000 divided by the sample volume.
 ‡ A "Version" greater than 1 indicates amended data.
 § Total Spores/m3 has been rounded to two significant figures to reflect analytical precision.
 TestAmerica Environmental Microbiology Laboratory, Inc.

EMLab P&K

1010 N Central Avenue, Glendale, CA 91202
(858) 569-5800 Fax (858) 569-5806 www.emlab.com

Client: Biomax Environmental
C/O: Mr. Michael Polkabla
Re: 110508-01; DGS

Date of Sampling: 11-05-2008
Date of Receipt: 11-06-2008
Date of Report: 11-07-2008

MoldRANGE™: Extended Outdoor Comparison**Outdoor Location: 14226828, Ambient garage rooftop E**

Fungi Identified	Outdoor data	Typical Outdoor Data by Date†				Typical Outdoor Data by Location‡			
		Month: November				State: CA			
	spores/m3	low	med	high	freq %	low	med	high	freq %
Generally able to grow indoors*									
Alternaria	-	7	27	280	54	7	27	210	58
Bipolaris/Drechslera group	-	7	13	190	19	7	13	120	13
Chaetomium	-	7	13	210	12	7	13	120	19
Cladosporium	5,200	40	640	11,000	95	53	640	6,400	98
Curvularia	-	7	22	800	19	7	13	210	7
Epicoccum	13	7	13	210	25	7	13	160	20
Nigrospora	-	7	13	210	19	7	13	170	8
Other brown	-	7	13	110	36	7	13	80	37
Penicillium/Aspergillus types	53	27	270	3,100	87	38	210	2,500	87
Stachybotrys	-	7	13	260	4	7	13	290	5
Stemphylium	13	7	13	73	5	7	13	67	9
Torula	-	7	13	130	11	7	13	150	12
Seldom found growing indoors**									
Ascospores	170	13	120	2,700	75	13	110	1,800	72
Basidiospores	3,100	13	400	16,000	94	13	210	6,700	94
Rusts	-	7	13	280	25	7	13	250	28
Smuts, Periconia, Myxomycetes	40	7	50	730	75	8	40	480	70
TOTAL SPORES/M3	8,589								

† The Typical Outdoor Data by Date represents the typical outdoor spore levels across North America for the month indicated. The last column represents the frequency of occurrence. The low, medium, and high values represent the 2.5, 50, and 97.5 percentile values of the spore type when it is detected. For example, if the frequency of occurrence is 63% and the low value is 53, it would mean that the given spore type is detected 63% of the time and, when detected, 2.5% of the time it is present in levels above the detection limit and below 53 spores/m³. These values are updated periodically, and if enough data is not available to make a statistically meaningful assessment, it is indicated with a dash.

‡ The Typical Outdoor Data by Location represents the typical outdoor spore levels for the region indicated for the entire year. As with the Typical Outdoor Data by Date, the four columns represent the frequency of occurrence and the typical low, medium, and high concentration values for the spore type indicated. These values are updated periodically, and if enough data is not available to make a statistically meaningful assessment, it is indicated with a dash.

*The spores in this category are generally capable of growing on wet building materials in addition to growing outdoors. Building related growth is dependent upon the fungal type, moisture level, type of material, and other factors. *Cladosporium* is one of the predominant spore types worldwide and is frequently present in high numbers. *Penicillium/Aspergillus* species colonize both outdoor and indoor wet surfaces rapidly and are very easily dispersed. Other genera are usually present in lesser numbers.

**These fungi are generally not found growing on wet building materials. For example, the rusts and smuts are obligate plant pathogens. However, in each group there are notable exceptions. For example, agents of wood decay are members of the basidiomycetes and high counts of a single morphological type of basidiospore on an inside sample should be considered significant.

Interpretation of the data contained in this report is left to the client or the persons who conducted the field work. This report is provided for informational and comparative purposes only and should not be relied upon for any other purpose. "Typical outdoor data" are based on the results of the analysis of samples delivered to and analyzed by EMLab P&K and assumptions regarding the origins of those samples. Sampling techniques, contaminants infecting samples, unrepresentative samples and other similar or dissimilar factors may affect these results. In addition, EMLab P&K may not have received and tested a representative number of samples for every region or time period. EMLab P&K hereby disclaims any liability for any and all direct, indirect, punitive, incidental, special or consequential damages arising out of the use or interpretation of the data contained in, or any actions taken or omitted in reliance upon, this report.

EMLab P&K

1010 N Central Avenue, Glendale, CA 91202
(858) 569-5800 Fax (858) 569-5806 www.emlab.com

Client: Biomax Environmental
C/O: Mr. Michael Polkabl
Re: 110508-01; DGS

Date of Sampling: 11-05-2008
Date of Receipt: 11-06-2008
Date of Report: 11-07-2008

MoldRANGE™: Extended Outdoor Comparison**Outdoor Location: 14354900, Ambient garage roof-post**

Fungi Identified	Outdoor data	Typical Outdoor Data by Date†				Typical Outdoor Data by Location‡			
		Month: November				State: CA			
	spores/m3	low	med	high	freq %	low	med	high	freq %
Generally able to grow indoors*									
Alternaria	-	7	27	280	54	7	27	210	58
Bipolaris/Drechslera group	-	7	13	190	19	7	13	120	13
Chaetomium	-	7	13	210	12	7	13	120	19
Cladosporium	1,100	40	640	11,000	95	53	640	6,400	98
Curvularia	-	7	22	800	19	7	13	210	7
Epicoccum	-	7	13	210	25	7	13	160	20
Nigrospora	80	7	13	210	19	7	13	170	8
Other brown	13	7	13	110	36	7	13	80	37
Penicillium/Aspergillus types	-	27	270	3,100	87	38	210	2,500	87
Stachybotrys	-	7	13	260	4	7	13	290	5
Stemphylium	-	7	13	73	5	7	13	67	9
Torula	-	7	13	130	11	7	13	150	12
Seldom found growing indoors**									
Ascospores	160	13	120	2,700	75	13	110	1,800	72
Basidiospores	4,000	13	400	16,000	94	13	210	6,700	94
Rusts	-	7	13	280	25	7	13	250	28
Smuts, Periconia, Myxomycetes	13	7	50	730	75	8	40	480	70
TOTAL SPORES/M3	5,366								

† The Typical Outdoor Data by Date represents the typical outdoor spore levels across North America for the month indicated. The last column represents the frequency of occurrence. The low, medium, and high values represent the 2.5, 50, and 97.5 percentile values of the spore type when it is detected. For example, if the frequency of occurrence is 63% and the low value is 53, it would mean that the given spore type is detected 63% of the time and, when detected, 2.5% of the time it is present in levels above the detection limit and below 53 spores/m³. These values are updated periodically, and if enough data is not available to make a statistically meaningful assessment, it is indicated with a dash.

‡ The Typical Outdoor Data by Location represents the typical outdoor spore levels for the region indicated for the entire year. As with the Typical Outdoor Data by Date, the four columns represent the frequency of occurrence and the typical low, medium, and high concentration values for the spore type indicated. These values are updated periodically, and if enough data is not available to make a statistically meaningful assessment, it is indicated with a dash.

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**These fungi are generally not found growing on wet building materials. For example, the rusts and smuts are obligate plant pathogens. However, in each group there are notable exceptions. For example, agents of wood decay are members of the basidiomycetes and high counts of a single morphological type of basidiospore on an inside sample should be considered significant.

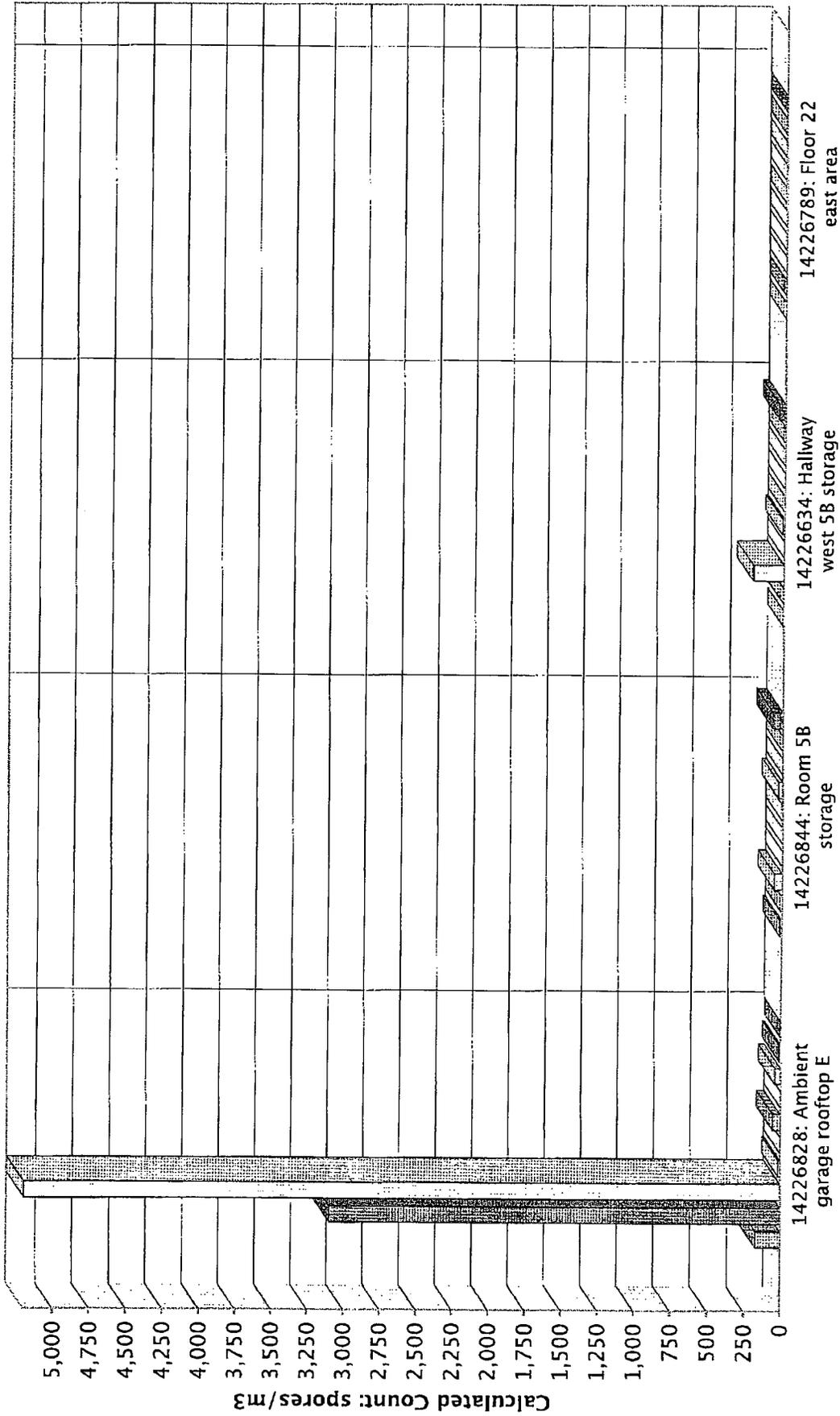
Interpretation of the data contained in this report is left to the client or the persons who conducted the field work. This report is provided for informational and comparative purposes only and should not be relied upon for any other purpose. "Typical outdoor data" are based on the results of the analysis of samples delivered to and analyzed by EMLab P&K and assumptions regarding the origins of those samples. Sampling techniques, contaminants infecting samples, unrepresentative samples and other similar or dissimilar factors may affect these results. In addition, EMLab P&K may not have received and tested a representative number of samples for every region or time period. EMLab P&K hereby disclaims any liability for any and all direct, indirect, punitive, incidental, special or consequential damages arising out of the use or interpretation of the data contained in, or any actions taken or omitted in reliance upon, this report.

11-07-2008: 110508-01

EMLab P&K
1010 N Central Avenue, Glendale, CA 91202
(858) 569-5800 Fax (858) 569-5806 www.emlab.com

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

- Ascospores Basidiospores Cladosporium Epicoccum Nigrospora Penicillium/Aspergillus types Rusts
- Smuts, Periconia, Myxomycetes Stemphylium Ulocladium

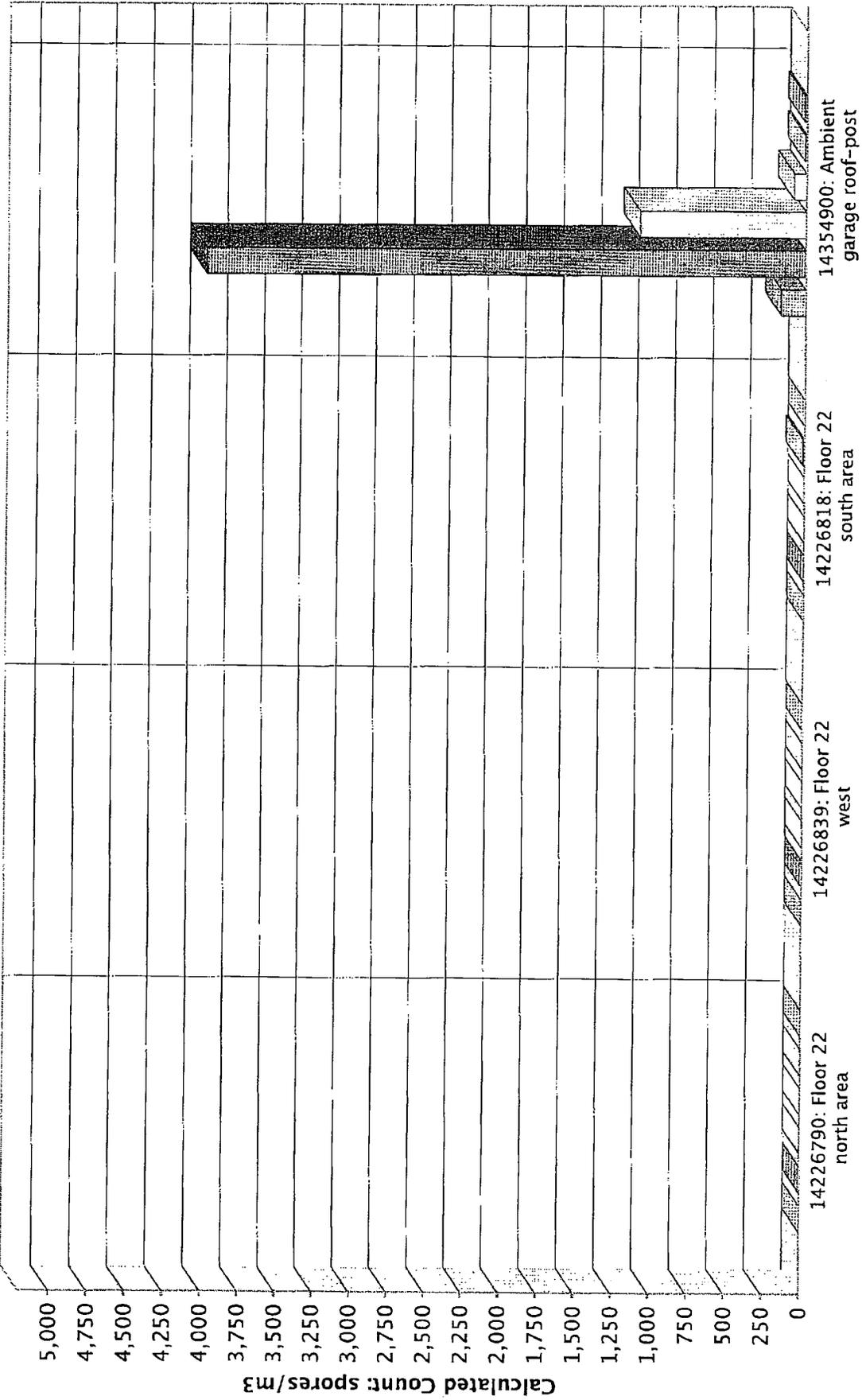


Comments:

Note: Graphical output may understate the importance of certain "marker" genera.
TestAmerica Environmental Microbiology Laboratory, Inc.

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Ascospores Basidiospores Cladosporium Nigrospora Other brown Smuts, Periconia, Myxomycetes



Comments:

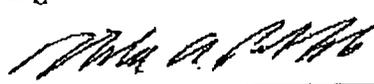
Note: Graphical output may understate the importance of certain "marker" genera.
TestAmerica Environmental Microbiology Laboratory, Inc.

MICROBIAL SPORE TRAP AIR SAMPLING RECORD

BioMax Environmental
775 San Pablo Ave.
Pinole, CA 94564

www.biomaxenvironmental.com

Phone: (510) 724-3100
Fax: (510) 724-3145
biomaxenv@aol.com

Location: 450 N. Street Sacramento, CA 5B Storage Rm.	Client: DGS Project #: 110508-01
Date: 11/5/08 Collected by: M. A. Polkable Signature: 	Laboratory: EMLabs Req. Turn Around: 24 HR Analysis (circle): <u>Fungal</u> <u>Particulate</u> <u>ID/Quantification.</u>

Sample Number	Time	Location/Desc.	Temp./RH
14226828	1320	Ambient Garage Rooftop E	60°/40%
14226844	1335	Room 5B Storage	72/28%
14226634	1355	Hallway w/d 5B Storage	72/30%
14226789	1405	Floor 22 East Area	76°/29%
14226790	1412	Floor 22 North Area	77°/31%
14226839	1420	Floor 22 west	76°/31%
14226818	1435	Floor 22 South Area	75°/32%
14354900	1455	Ambient Garage Roof Post	61°/38%
			
Total Sample Time (min): 5	Flow Rate (l/min): 15	Total Sample Volume (liters): 75 L	Ambient Conditions: Cool/mild NW 0-5

Please sign this form below acknowledging sample receipt and return executed form with laboratory reports. Fax, send, e-mail results to BioMax Environmental at (510) 724-3145 biomaxenv@aol.com
 Other Instructions: Fungal and particulate ID + Quantification

Relinquished by: <u>M. A. Polkable</u> Method of Transportation: <u>Fed Ex</u> Time/Date Sent: <u>3:40 11/5/08</u>	Received By: <u>Ann Marissay</u> Time/Date Received: <u>11-6-08 9:25</u>
--	---

Note - Fungal and particulate ID + Quantification
A.

**EMLab P&K**

Report for:

Mr. Michael Polkabila
Biomax Environmental
775 San Pablo Ave.
Pinole, CA 94564

Regarding: Project: BOE Building, 450 N Street, Sac, CA; Storage Rms, Electric and Telephone, Data Rooms
EML ID: 483563

Approved by:

A handwritten signature in black ink, appearing to read "Dr. Kamashwaran Ramanathan".

Lab Manager
Dr. Kamashwaran Ramanathan

Dates of Analysis:

Quantitative spore count direct exam: 10-31-2008

Project SOPs: Quantitative spore count direct exam (I100006)

This coversheet is included with your report in order to comply with AIHA and ISO accreditation requirements.

For clarity, we report the number of significant digits as calculated; but, due to the nature of this type of biological data, the number of significant digits that is used for interpretation should generally be one or two. All samples were received in acceptable condition unless noted in the Report Comments portion in the body of the report. Due to the nature of the analyses performed, field blank corrections of results is not a standard practice. The results relate only to the items tested.

EMLab P&K ("the Company") shall have no liability to the client or the client's customer with respect to decisions or recommendations made, actions taken or courses of conduct implemented by either the client or the client's customer as a result of or based upon the Test Results. In no event shall the Company be liable to the client with respect to the Test Results except for the Company's own willful misconduct or gross negligence nor shall the Company be liable for incidental or consequential damages or lost profits or revenues to the fullest extent such liability may be disclaimed by law, even if the Company has been advised of the possibility of such damages, lost profits or lost revenues. In no event shall the Company's liability with respect to the Test Results exceed the amount paid to the Company by the client therefor.

Document Number: 200091 - Revision Number: 5

EMLab P&K

1150 Bayhill Drive, Suite 100, San Bruno, CA 94066
 (650) 829-5800 Fax (650) 829-5852 www.emlab.com

Client: Biomax Environmental
 C/O: Mr. Michael Polkabila
 Re: BOE Building, 450 N Street, Sac, CA; Storage
 Rms, Electric and Telephone, Data Rooms

Date of Sampling: 10-29-2008
 Date of Receipt: 10-30-2008
 Date of Report: 10-31-2008

QUANTITATIVE SPORE COUNT REPORT

Location:	S01: Storage rm 6C, black smudges, North wall at 5' aft		S02: Storage rm 5C, left wall surface, N interior		S03: Electrical rm, East side, North end, fuzzy black material	
Comments (see below)	None		None		None	
Sample type	Tape sample		Tape sample		Tape sample	
Lab ID-Version‡:	2137818-1		2137819-1		2137820-1	
	raw ct.	spores/unit	raw ct.	spores/unit	raw ct.	spores/unit
Alternaria						
Arthrinium						
Ascospores*						
Aureobasidium						
Basidiospores*						
Bipolaris/Drechslera group						
Botrytis						
Chaetomium						
Cladosporium					106	23
Curvularia						
Epicoccum						
Fusarium						
Myrothecium						
Nigrospora						
Other brown					1	0.22
Other colorless						
Penicillium/Aspergillus types†			2,210	11,000		
Pithomyces						
Rusts*						
Smuts*, Periconia, Myxomycetes*						
Stachybotrys						
Stemphylium						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)††	1+		2+		4+	
Sample size	100		100		100	
Unit	1 mm2		1 mm2		1 mm2	
§ TOTAL SPORES/UNIT		< 0.01		11,000		23

Comments:

* Most of these spore types are not seen with culturable methods (Andersen sampling), although some may appear as nonsporulating colonies. Most of the basidiospores are 'mushroom' spores while the rusts and smuts are plant pathogens.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

†† Background debris is an indication of the amount of non-biological particulate matter present on the slide (dust in the air) and is graded from 1+ to 4+ with 4+ indicating the largest amounts. This background material is also an indication of visibility for the analyst and resultant difficulty reading the slide. For example, high background debris may obscure the small spores such as the *Penicillium/Aspergillus* group. Counts from areas with 4+ background debris should be regarded as minimal counts and may actually be higher than reported.

‡ A "Version" greater than 1 indicates amended data.

§ Total Spores/unit has been rounded to two significant figures to reflect analytical precision.

BULK / SURFACE SAMPLING RECORD BIOMAX ENVIRONMENTAL, LLC

775 San Pablo Avenue, Pinole, CA 94564

Phone: (510) 724-3100 Fax (510) 724-31435 biomaxenv@aol.com

Project Name and Location: *BOE Building - 450 N. Street, Sacramento, CA*
Storage Rms / Electric + telephone / Data Rooms

Analytical Laboratory: *EMUDAS* Date of Sampling: *10/29/08* Required Turn Around:

Analysis Requested: *Fungal ID - Microscopy* Sampled By: *MAFB/keble*

Sample ID	Sample Type B/S	Area/Volume Sampled	Location/Description
<i>S01</i>	<i>Surface</i>	<i>1 x 1 "</i>	<i>Storage Rm 6C - Black smudges</i> <i>North wall @ 5' off</i>
<i>S02</i>	<i>Surface</i>	<i>1 x 1 "</i>	<i>Storage Rm 5C</i> <i>Left wall surface - N, interior</i>
<i>S03</i>	<i>surf</i>	<i>1x1</i>	<i>Electrical Rm East Side - North end</i> <i>Fuzzy black material</i>



Instructions and Comments: *Fungal ID - Microscopy*

Please sign this form below acknowledging sample receipt and return executed form with laboratory reports. Fax, send and e-mail results to BioMax Environmental at (510) 724-3145 biomaxenv@aol.com

Relinquished by: <i>Maria A. ...</i>	Received By: <i>Am Morrissey</i>
Method of Transportation: <i>FedEx</i>	Time/Date Received: <i>10-30-08 9:15</i>
Time/Date Sent: <i>4:00 10/29/08</i>	

EMLab P&K

1150 Bayhill Drive, Suite 100, San Bruno, CA 94066
 (650) 829-5800 Fax (650) 829-5852 www.emlab.com

Client: Biomax Environmental
 C/O: Mr. Michael Polkabra
 Re: 110508-01; DGS

Date of Sampling: 11-05-2008
 Date of Receipt: 11-06-2008
 Date of Report: 11-07-2008

QUANTITATIVE SPORE COUNT REPORT

Location:	S01:		S02:	
	5B large black colonies on wall 6" afl		5B large brown staining on wall 6" afl	
Comments (see below)	None		None	
Sample type	Tape sample		Tape sample	
Lab ID-Version‡:	2145309-1		2145310-1	
	raw ct.	spores/unit	raw ct.	spores/unit
Alternaria				
Arthrinium				
Ascospores*				
Aureobasidium				
Basidiospores*				
Bipolaris/Drechslera group				
Botrytis				
Chaetomium				
Cladosporium				
Curvularia				
Epicoccum				
Fusarium				
Myrothecium				
Nigrospora				
Other colorless				
Penicillium/Aspergillus types†	48	240	430	2,200
Pithomyces				
Rusts*				
Smuts*, Periconia, Myxomycetes*				
Stachybotrys				
Stemphylium				
Torula				
Ulocladium	264	1,300	270	1,400
Zygomycetes				
Background debris (1-4+)††	2+		2+	
Sample size	100		100	
Unit	1 mm2		1 mm2	
§ TOTAL SPORES/UNIT		1.600		3.500

Comments:

* Most of these spore types are not seen with culturable methods (Andersen sampling), although some may appear as nonsporulating colonics. Most of the basidiospores are 'mushroom' spores while the rusts and smuts are plant pathogens.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

†† Background debris is an indication of the amount of non-biological particulate matter present on the slide (dust in the air) and is graded from 1+ to 4+ with 4+ indicating the largest amounts. This background material is also an indication of visibility for the analyst and resultant difficulty reading the slide. For example, high background debris may obscure the small spores such as the *Penicillium/Aspergillus* group. Counts from areas with 4+ background debris should be regarded as minimal counts and may actually be higher than reported.

‡ A "Version" greater than 1 indicates amended data.

§ Total Spores/unit has been rounded to two significant figures to reflect analytical precision.

BULK / SURFACE SAMPLING RECORD

BIOMAX ENVIRONMENTAL, LLC

775 San Pablo Avenue
Pinole, CA 94564

Phone: (510) 724-3100 Fax (510) 724-31435 biomaxenv@aol.com

Project Name and Location: *DGS / BOE Bid Storage Room 5B*

Client: *DGS*
450 N. Street
Sac. CA

Analysis Requested: *Fungal ID*

Analytical Laboratory: *FMLabs* Date of Sampling: *11/5/08*

Required Turn Around Time: *24 HR* Sampled By: *MA. Pollock*

Sample ID Number	Sample Type (Bulk/Surface)	Area/Volume Sampled	Location/Description
<i>501</i>	<i>Surf.</i>	<i>1 x 1</i>	<i>5B large black colonies on wall</i>
			<i>6" aER</i>
<i>502</i>	<i>Surf.</i>	<i>1 x 1</i>	<i>5B large Brown staining on wall</i>
			<i>6" aER</i>
			<i>< 4 SF concrete w/ mold</i>
			<i>2x paint stripping areas ~ 3 SF</i>

Instructions and Comments: *Fungal ID - Microscopy*

Please sign this form below acknowledging sample receipt and return executed form with laboratory reports. Fax, send and e-mail results to BioMax Environmental at (510) 724-3145 biomaxenv@aol.com

Relinquished by: <i>[Signature]</i>	Received By: <i>Ann Marissey</i>
Method of Transportation: <i>FedEx</i>	
Time/Date Sent: <i>4:00 11/5/08</i>	Time/Date Received: <i>11-6-08 9:25</i>

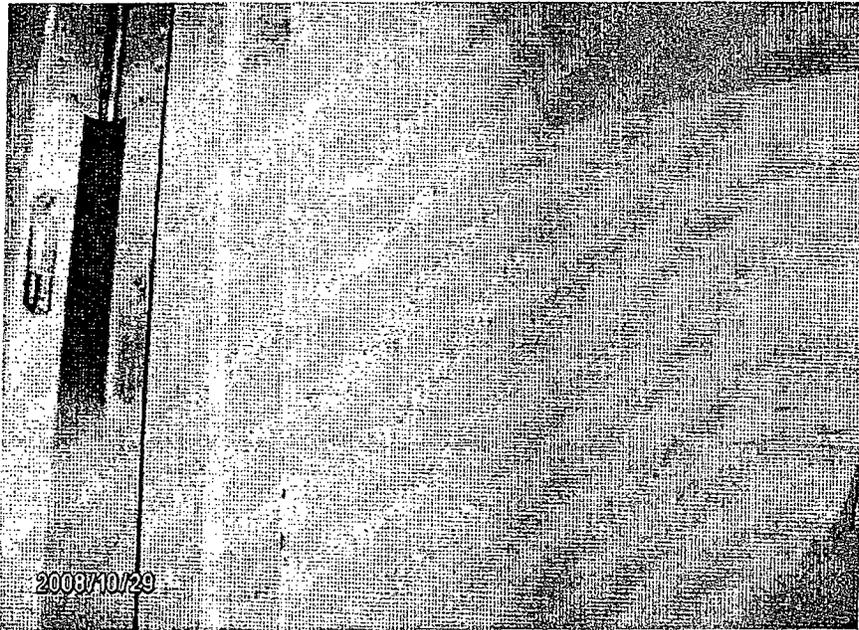


Attachment A: Digital Images

Page 1 of 5

October 29th, 2008BOE Building 5th and 6th Floor Storage Area Assessment
Sacramento, CA[Click here for color photos](#)

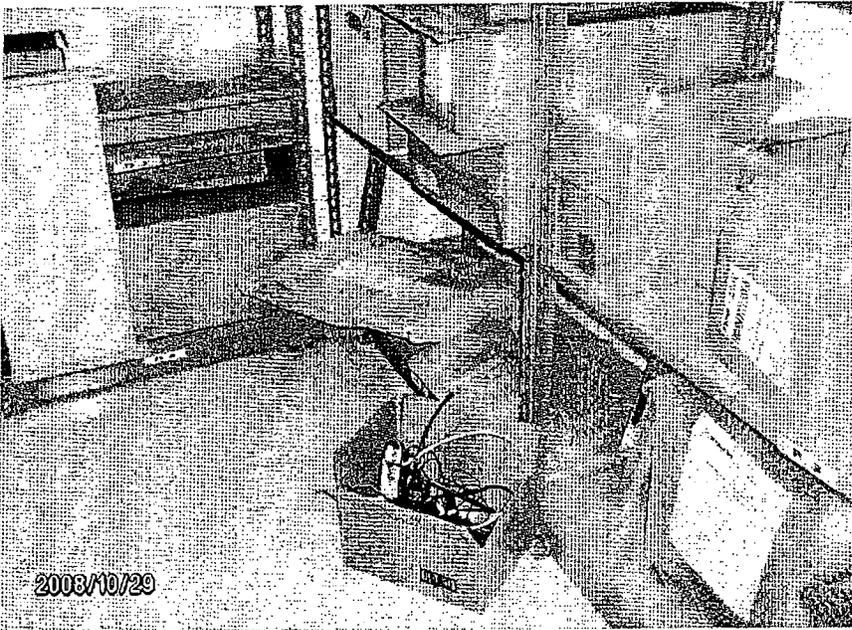
- 1) Condition of Storage Room 6C wherein extensive quantities of cubicle furniture was stored creating limited access for inspection and sampling during assessment.



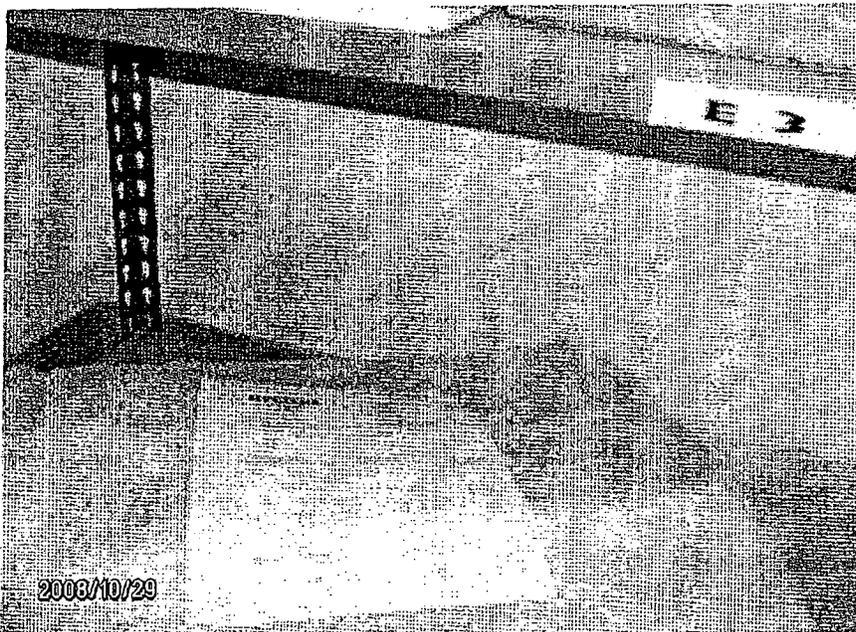
- 2) Image within Storage Room 6C of wall surface indicating suspect "smudges" which were sampled as part of surface assessment on 10/29/08.

October 29th, 2008
BOE Building 5th and 6th Floor Storage Area Assessment
Sacramento, CA

Page 2 of 5



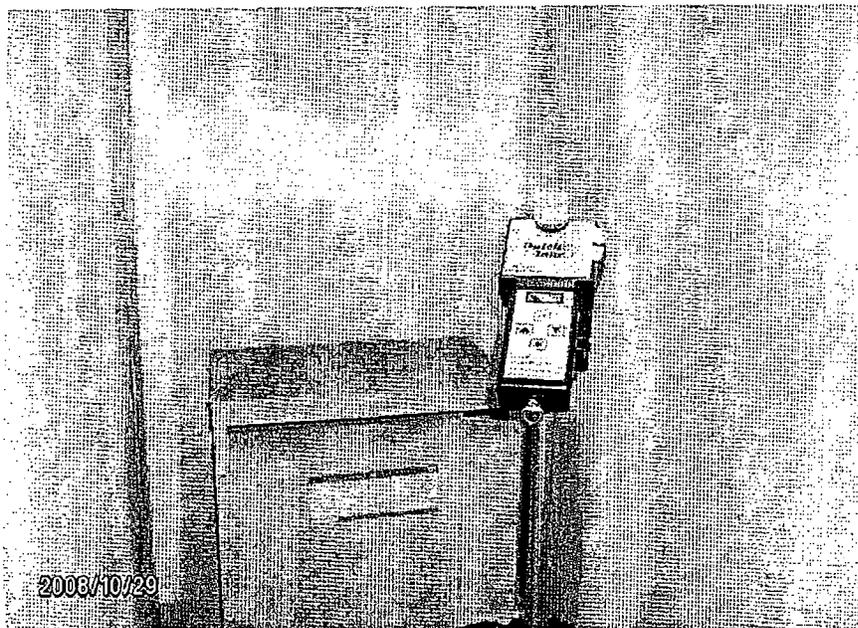
- 3) Image within Storage Room 5C wherein wall-bolted metal framed shelving had covered localized suspect mold-like growth at lower shelf area.



- 4) Close-up image of stained wall surface following removal of office equipment from lower shelf at time of assessment.

October 29th, 2008
BOE Building 5th and 6th Floor Storage Area Assessment
Sacramento, CA

Page 3 of 5



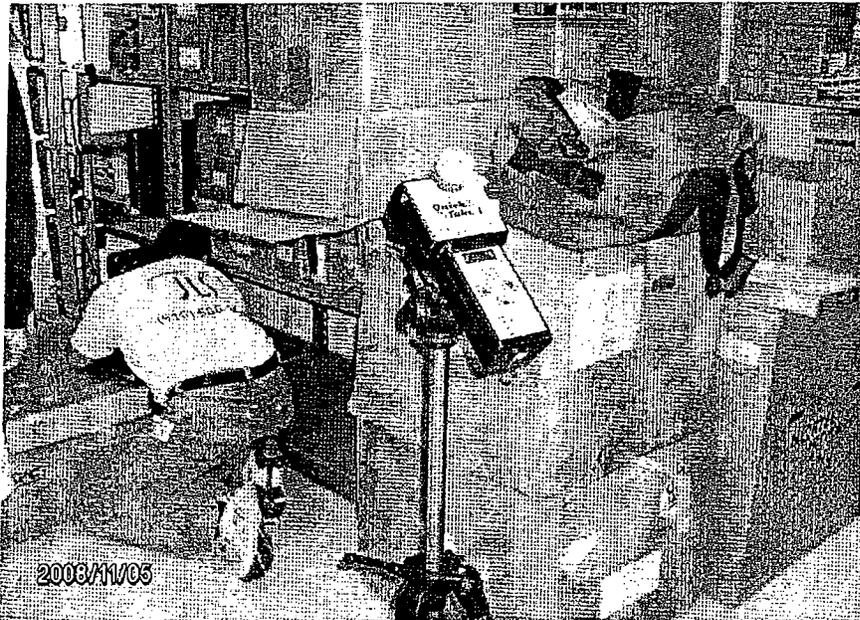
- 5) Image of air sampling and wall debris present within Electrical and Telephone Equipment Room located at northeastern access doorway of 5th floor.



- 6) Close-up image of black “fuzzy” material present on southern wall within 5th Floor Electrical and Telephone Equipment Room.

November 5th, 2008
BOE Building 5th Floor Storage Area Assessment Supplement
Sacramento, CA

Page 4 of 5



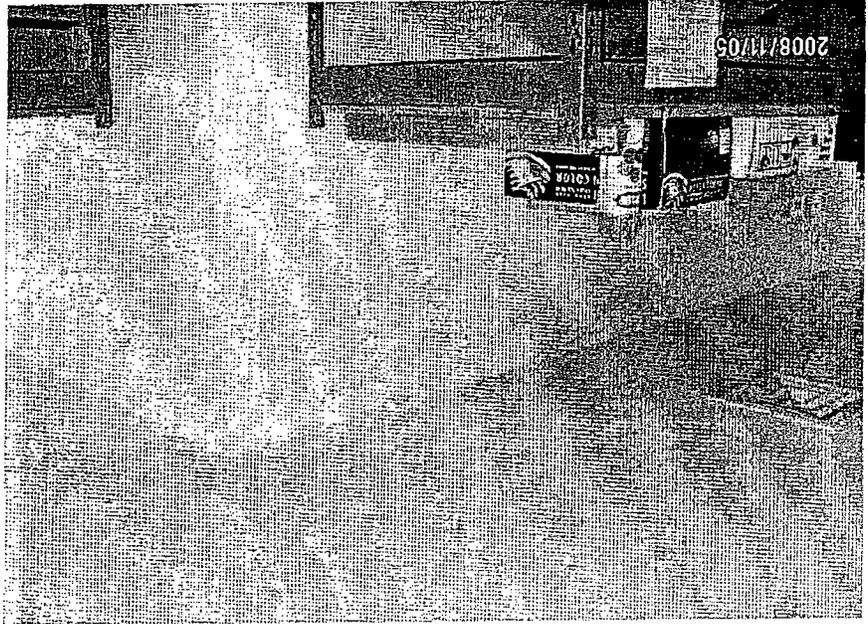
7) Image of air sampling and storage materials present within storage room 5B at time of follow-up airborne and surface assessment performed on 11/05/08.



8) Image of stored material covering stained wallboard material within storage Room 5B at time of assessment.



9) Close image of stained material on wallboard surface within storage Room 5B at time of assessment.



10) Image of "shuffed" paint material present on wall-to-ceiling interface within Storage Room 5B at area above and adjacent to visible wall staining.