

TAB 22

BioMax Environmental

Environmental Consulting and Industrial Hygiene Services

October 7th, 2008

Mr. Doug Button
Deputy Director
Real Estate Services Division
707 Third Street - 8th Floor
West Sacramento, CA 95605

Post Mitigation Assessment Report
Department of General Services
Board of Equalization Building, 450 N. Street
Break Room 707 and 706 Areas
Sacramento, California

Mr. Button,

BioMax Environmental, LLC (BioMax) is pleased to provide The Department of General Services (DGS) with this letter summary report detailing BioMax's findings and recommendations pertaining to our post mitigation microbial inspection and sampling assessment services provided within the noted break room and adjacent supply room areas within the Board of Equalization (BOE) building located at 450 N Street, Sacramento, California. BioMax understands that these post mitigation microbial inspection and sampling assessment services were contracted with BioMax, at your request, in an effort to review and verify the successful completion of microbial mitigative efforts performed by your restoration contractor, JLS Environmental, Inc., (JLS) within the previously identified areas located within the subject building.

Therefore, these post mitigation clearance assessment services are intended to assess the current site conditions wherein mitigative activities were performed by JLS to investigate and address (as needed) the prior moisture and mold related damages and impacts. Procedural recommendations pertaining to BioMax's review of historical and analytical data associated with the subject break room and adjacent interior areas have been summarized within our previously developed procedural assessment reports including those entitled:

- Mitigative and Clean Up Procedures for Interior Electrical/Data Rooms, Janitorial Rooms, Supply Rooms, Copy Rooms, Storage Rooms, and Rest Room Areas, dated May 7th, 2008.
- Microbial Assessment of Break Room Areas ("Building Wide"), dated July 11th, 2008

Additional historical reports and assessment data may also be obtained for further background and technical reference, as necessary.

Hence, these post mitigation microbial clearance assessment services, thereby, are intended to provide a professional evaluation verifying the physical conditions wherein the successful completion of microbial removal and decontamination within each of the affected areas has been achieved. Following the completion of the prescribed mitigative activities performed by your mitigation contractor, Mr. Michael A. Polkabila, CIH, REA of BioMax performed a detailed post mitigation site inspection and sampling assessment within each of the affected interior areas (and adjacent impacted areas as necessary) as noted in this report. BioMax's findings and conclusions pertaining to these post mitigation sampling assessment activities are, therefore, summarized herein.

SITE OBSERVATIONS

Site inspection and post mitigation assessment sampling activities were performed within the noted Break Room 707 and adjacent supply room 706 on October 1st, 2008. Site access into each of these contained areas was facilitated by site contractor and DGS personnel. On this date, Mr. Michael A. Polkabila, CIH, REA of BioMax performed a detailed visual site inspection within the noted containment system barriers associated with the noted break room and supply room areas. Following the performance of a detailed visual assessment within these areas (indicating acceptable visual post mitigation conditions), BioMax collected a series of airborne SporeTrap confirmation samples within and surrounding each of these areas as noted below.

On-site inspection and clearance sampling assessment activities were performed by Mr. Michael A. Polkabila, CIH, REA, of BioMax in accordance with currently recognized microbial assessment and sampling guideline procedures. Mr. Polkabila has been certified in the Comprehensive Practice of Industrial Hygiene by the American Board of Industrial Hygiene and holds the right to the designation "Certified Industrial Hygienist" (CIH) under certification number CP 7104. Mr. Polkabila is also certified by the California Environmental Protection Agency (Cal/EPA) as a Class I Registered Environmental Assessor (REA) under Cal/EPA certification number 05011. Previously established clearance criteria developed for these activities has been formalized in BioMax's Post Mitigation Clearance Assessment Protocols dated February 15th, 2008. Such protocols have been reviewed and approved by BOE's environmental consultant, Hygientech International, Inc. (HTI) prior to implementation. A summary of significant notations and observations gathered during BioMax's site inspection and post mitigation clearance assessment activities within the subject containment areas are compiled as follows:

1. At the time of our site inspection and clearance sampling assessment performed on October 1st, 2008 ambient outdoor conditions both prior to and following our interior assessment activities consisted of clear and mild conditions with an outdoor temperatures range between 77 and 79 degrees F and relative humidity of 28-28.5 %, respectively. Predominant winds were noted at approximately 0-5 knots from the westerly direction at the time of our

assessment. Interior environmental conditions within the sampled containment areas consisted of a temperature range between 73 and 74 degrees F with relative humidity of 29 percent.

2. At the time of this assessment, each of the observed interior containment barrier systems, whereby microbial mitigative and inspection activities were performed, were established and maintained within the impacted areas as per BioMax's protocols. Specific detail as noted on the "as built" construction site floor diagram documents may be reviewed for further reference as necessary. BioMax routinely performed regular and periodic inspections and review of records/conditions within and surrounding each of the noted containment areas during mitigative activities. A review of such information has indicated a preponderance of evidence verifying that the current barrier systems have provided appropriate protective controls for the duration and performance of the noted mitigative activities.
3. During the post mitigation inspection of each containment system, BioMax noted the absence of visible evidence of elevated residual moisture and/or microbial indicators (such as staining, delamination, etc.) within the remaining exposed interior walls, flooring, wall framing, and wall cavities following the performance of mitigative measures. Utilization of a TraMex hand-held inductive moisture meter indicated normal moisture content within all remaining walls and building materials inspected within the sampled containment areas at the time of our assessment.
4. As noted within the previously referenced assessment reports, the primary affected areas of visible moisture damage previously identified within the noted break room primarily included moisture staining and mold damaged cabinetry, adjacent flooring, and wallboard materials. According to BioMax's review of current evidence and available historical data, it is BioMax's opinion that such material damage was likely caused by a history of chronic plumbing deficiencies and water release events over an extended period of time.
5. Primary affected areas of visible moisture damage previously identified within the noted adjacent supply room area primarily consisted of moisture staining and mold damaged materials within the shared wallboard materials adjacent to the break room sink. According to BioMax's review of current evidence and available historical data, it is likely that such material damage were similarly caused by a history of chronic plumbing deficiencies and water release events over an extended period of time as no interior sources were identified within the supply room at the time of this assessment.
6. The establishment of containment system barriers encompassing each of the interior affected areas (including break room 707 and supply room 706) were observed and verified under appropriate posting and negative pressure differential at the time of this post mitigation assessment. Worker and equipment entry and exit chambers comprised of a series of zippered plastic access doorways were also observed attached to the noted containment barriers consistent with BioMax's previously noted mitigation protocols.

7. As verified during these assessment activities, all identified affected interior wallboard building materials had been removed from each of the noted interior areas of concern exposing interior wall cavity framing (metal) and underlayment wallboard siding materials present within each of the impacted materials and areas. Upon post mitigation inspection, all remaining exposed building materials associated within the break room and supply room areas exhibited no significant staining and/or elevated mold growth following the completion of prescribed physical material removal and chemical decontamination procedures performed by JLS within each impacted area.
8. Digital images and schematic records have been developed and maintained by JLS for the duration the performance of these mitigative removal activities indicating the extent and areas where visible staining and/or mold like indicators have been identified within the exposed wall cavities and wall cavity underlayment materials and subsequently removed within each containment area. Such records have been reviewed by BioMax as part of this clearance assessment and may be provided by JLS for additional review upon request.
9. Following the completion of visual inspections within each of these containment areas, BioMax collected series airborne samples within and outside the containment systems noted below for subsequent comparative analysis. Such samples collected within and surrounding each the interior containment system were performed in an effort to identify and quantify the presence of potential airborne mold spores present within (and surrounding) the containment systems following the completion of the prescribed mitigative effort. Findings associated with these verification sampling activities are noted below.
10. BioMax also collected a series of digital images during these post mitigative inspection and sampling assessment activities to document the conditions and significant site observations gathered at this time. Such images are provided as an attachment to this summary report for further reference, as necessary.

SAMPLING PROCEDURES

On-site inspection and sampling assessment activities were conducted by Mr. Michael A. Polkabila, CIH, REA, of BioMax Environmental within the noted break room and supply room areas on October 1st, 2008. All sampling equipment, supplies, calibration materials, and collection media were provided and maintained by BioMax as part of the performance of this scope of work. Sample collection procedures and methods were performed using standard industrial hygiene sampling methods following techniques prescribed by the contracted analytical laboratory.

Spore Trap Airborne Microbial and Particulate Sampling:

The collection of airborne Spore Trap microbial samples was achieved using Zefon Air-O-Cell sampling cassette collection devices placed in each of the areas identified in the tables below. Airborne Spore Trap samples were collected within and outside each of the containment area

locations at a height of approximately four feet above ground level using a tripod mounted Quick Take 15 air sampling pump manufactured by SKC. Samples were collected at a calibrated flow rate of 15 liters per minute for a total of five minutes per sample. Resultant total sample volumes, therefore, corresponded to 75 liters collected for each collected sample. Field calibration of the SKC air sampling pump was conducted using a field rotometer device calibrated with a Bios Drycal primary standard flow meter. All spore trap air sampling and analytical procedures were performed in accordance with prescribed manufacturer guidelines as well as applicable professional certified industrial hygiene indoor air quality microbial investigation procedures and certified industrial hygiene practices.

Additional exterior ambient samples were also similarly collected and analyzed in an effort to identify and quantify representative background microbial taxa (types), rank order, and corresponding airborne spore levels present within the ambient environment at the time of this assessment for comparative purposes. Sampling collection activities performed during this study included the collection of identifiable airborne microbial contaminants within the representative area locations noted in Table 1:

Table 1. Airborne Spore Trap Sampling Locations of Break Room 707 and Supply Room 706:

Air Sample Number	Spore Trap Air Sampling Location
14226774	Ambient Post Sample at Ground Level Main Entrance
14226731	7 th Floor Hallway outside Break Room 707 (outside containment)
14226603	Break Room 707 (inside containment)
14226653	Supply Room 706 (inside containment)
14226607	Ambient Post Sample at Ground Level NE Corner

At the conclusion of sampling activities, preparation and shipping of the collected samples were accomplished in accordance with standard industrial hygiene chain of custody (COC) documentation procedures and quality assurance/quality control practices. Once collected, labeled, and recorded, all samples were double sealed within airtight plastic Ziploc shipping containers and transported via Federal Express Priority Mail to Environmental Microbial Laboratories (EMLabs) in San Bruno, California. EMLabs holds current applicable analytical accreditation and specializes in microbial analytical procedures. Sampling and chain of custody records are provided as an attachment to this letter report for further reference.

ANALYTICAL FINDINGS AND CONCLUSIONS

Airborne Spore Trap Findings Break Room 707 and Supply Room 706:

Laboratory analytical methods for the identification and enumeration of microbial (mold) taxa and particulate contaminants were conducted in accordance with prescribed analytical procedures and quality control/assurance measures. Original laboratory results including the enumeration of recognizable microbial spore and particulate types are also attached to this letter report for further reference and detail. A summary of airborne Spore Trap microbial (mold) and particulate findings pertaining to each of the subject areas are presented in Table 2 below:

Table 2. Airborne Microbial and Particulate Findings – Break Room 707 and 706 Supply Room.

Location Desc.	Total Mold Spores (Cts/m3)	Background Debris (scale of 1-4)	Skin Cell Fragments (scale of 1-4)	Hyphal Fragments (units/m3)
Ambient Post Sample at Ground Level Main Entrance	1,200	3+	<1+	93
7 th Floor Hallway outside Break Room 707 (outside containment)	120	2+	1+	<13
Break Room 707 (inside containment)	160	3+	2+	13
Supply Room 706 (inside containment)	66	3+	1+	13
Ambient Post Sample at Ground Level NE Corner	2,200	3+	<1+	67

The analytical findings presented in Table 2 above clearly indicate the presence of significantly lower concentrations of total microbial (mold) spores measured within each of the interior samples collected both within and surrounding the subject break room and supply room containment areas when compared to the levels currently measured within the samples collected from the corresponding ambient outside environment. Analytical findings also indicate similar fungal taxa distribution (mold types) and rank order (predominant taxa) of molds identified within the mitigated areas as well as the adjacent hallway areas sampled (area noted as "Hallway" outside containment). Analysis of fungal hyphal fragments (vegetative fungal growth structures) also indicated fewer structures within the interior containment areas and adjacent interior spaces when compared to the corresponding levels found within the ambient outside environmental samples. Particularly worthy of note, was the absence of elevated levels of

hydrophilic (moisture loving) mold taxa following the performance of mitigative activities within each of the noted containment areas.

Although there are currently no regulatory standards or limits pertaining to allowable airborne fungal concentrations (for any mold taxa) present in indoor environments, there is a general consensus among indoor air quality experts that airborne microbial contamination found within "typical healthy" living and working spaces are generally similar in kind and present at levels which are below those found in the corresponding native outside environment. BioMax believes that the absence of visible staining resultant from moisture and/or residual mold, the absence of elevated residual moisture, absence of elevated hyphal (mold growth) structures, and relatively fewer total airborne mold levels with typical taxa and rank order distribution following mitigative clean-up activities are consistent with these generally acceptable interior working space conditions. BioMax, therefore, believes that these findings provide reasonable evidence indicating that current microbial removal and clean-up measures have successfully removed and contained mold contamination within the above noted mitigated areas and materials to normal representative levels.

Based on these findings, BioMax believes that the current physical site conditions present within each of the mitigated areas may be considered acceptable in meeting both the visual and analytical clearance criteria established for these activities. As such, BioMax's review and interpretation of the collected analytical data associated with each of the noted containment areas has been shown to meet the previously referenced clearance criteria established for these activities. Such clearance criteria has been presented in BioMax's Post Mitigation Clearance Assessment Protocols dated February 15th, 2008, and has been reviewed and approved by BOE's environmental consultant, HTI. Therefore, BioMax believes that the verified achievement of such criteria supports BioMax's determination and conclusion that the noted break room and supply room areas may be considered acceptable for reconstruction at this time.

Airborne Particulate Findings:

Analytical particulate findings also sampled and analyzed as part of this assessment identified, what BioMax believes to be, "unremarkable" levels present within the collected air samples. Such findings within and surrounding the noted containment areas also provide reasonable evidence indicating that current particulate clean-up and mitigative control measures have successfully controlled and contained particulate debris within the identified containment areas to acceptable post mitigation clean-up levels.

RECOMMENDATIONS

Based on BioMax's post mitigation assessment findings and conclusions presented in this report, BioMax believes that the current airborne microbial levels sampled and analyzed from within break room 707 and supply room 706 (adjacent to break room 707) provides no significant evidence of elevated residual microbial contamination or airborne contamination/migration following the completion of the prescribed microbial mitigative measures. BioMax understands

that parallel airborne and surface assessment sampling performed within each of these containment areas by BOE's consultant, HTI, also indicated acceptable airborne and surface microbial levels (as provided to BioMax verbally) following the completion of the mitigative effort. BioMax was provided this information verbally at the time of this writing and anticipates that HTI will be preparing a formal summary report of their parallel findings for appropriate distribution shortly.

Hence, based on current site observations, field measurements, and review of all available findings (both BioMax's and HTI's) at this time, BioMax believes that the mitigated areas within the noted containment areas may be considered acceptable for general reconstruction following prudent reconstruction practices. Therefore, based on our professional review and interpretation of these current referenced findings, BioMax provides the following recommendations for consideration as discussed below:

1. BioMax believes that current airborne microbial (mold) levels and mold taxa (types) identified within Break Room 707 and Supply Room 706 are currently consistent with generally acceptable conditions and industry standard parameters following the performance of the mitigative activities noted. Hence, BioMax recommends that no further airborne and/or surface microbial sampling activities are warranted within these specific noted containment areas and that the containment systems may be deactivated and considered as "acceptable" for reconstruction at this time.
2. During the performance of interior reconstruction activities, BioMax recommends that a qualified and experienced building inspector/contractor be utilized to verify the current compliance and functional integrity of all applicable building related plumbing, flashing, sealing, and drainage systems in accordance with current building codes and construction practices. Any identified deficiencies should be appropriately documented, corrected, and functionally verified (tested) prior to subsequent reconstruction. Certainly, the establishment and/or installation of any additional corrective measures or engineering controls (as identified through additional professional engineering consultation) should also be performed and implemented in accordance with applicable standards, building codes, and ordinances, as appropriate.
3. BioMax recommends that reconstruction of interior structural building materials within these areas should only be undertaken utilizing high quality, visibly clean (hand selected) construction grade building materials obtained from reputable commercial sources and which are verified through visual assessment to be free from elevated microbial contamination and/or elevated moisture content. Building materials, which are notably moist and/or visibly stained, should not be used during the reconstruction undertaken within the subject building.
4. BioMax also recommends that current plastic barriers (as established during this mitigative activity) should remain during any reconstruction activity so as to minimize the potential transmission of associated nuisance construction dust and debris as desired.

5. Reasonable additional assessment and investigative measures may also be required upon the identification of new or previously undiscovered materials and/or information related to moisture/microbial impacts within the noted structures and/or areas, as necessary. Any occurrence and/or re-occurrence of moisture intrusion following reconstruction within these areas should also be reviewed and addressed through additional professional consultation, as necessary. BioMax is certainly prepared to provide such professional consultation pertaining to these and any follow-up investigative measures upon request.

BioMax believes that the conclusions and recommendations provided above are consistent with standard industry microbial mitigative practices and prudent industrial hygiene hazard control and assessment methods. Please do not hesitate to contact me directly at (510) 724-3100 if you have any questions, comments, and/or require further assistance regarding this subject matter.

Sincerely,



Michael A. Polkabila, CIH, REA
Vice President, Principal



LIMITATIONS

Please note that the professional opinions presented in this review are intended for the sole use of the California State Department of General Services (DGS) and their designated beneficiaries. No other party should rely on the information contained herein without the prior written consent of BioMax Environmental and DGS. The professional opinions provided herein are based on BioMax's review and understanding of current site information and observed site conditions present within the areas inspected at the time these services were performed. Professional recommendations provided as part of this limited scope of work are intended for client consideration only and are not intended as a professional or regulatory mandate. Implementation of any of the above measures or recommendations does not, in any way, warrant the day-to-day health and/or safety of building occupants, residents, site workers, nor regulatory or building code compliance status during normal and changing environmental conditions. As microbial contamination, by nature, may change over time due to additional moisture intrusion, favorable growth conditions, and changing environments, the findings of this report are subject to change in the event that such conditions and/or environments arise. Also, the professional opinions expressed here are subject to revision in the event that new or previously undiscovered information is obtained or uncovered.

The information contained in this and any other applicable communication is for consideration purposes only. It is not intended, nor should it be construed as providing legal advice or warranting any level of safety or regulatory compliance. The sole purpose of such information is to assist with the anticipation, identification, evaluation and control of elevated and/or unnecessary health of physical hazards. Any action taken based on this information, including but not limited to opinions, suggestions and recommendations, whether implied or expressed, is the sole responsibility of the individual taking the action. The management of acceptable health and safety is criteria dependent and situation specific in nature, therefore requiring extensive knowledge and prudent value assessments so as to be properly determined and maintained.

These services were performed by BioMax in accordance with generally accepted professional industrial hygiene principals, practices, and standards of care. Under the existing Industrial Hygiene Definition and Registration Act, all reports, opinions or official documents prepared by a Certified Industrial Hygienist (CIH) constitutes an expression of professional opinion regarding those facts or findings which are subject of a certification and does not constitute a warranty or guarantee, either expressed or implied.

MICROBIAL SPORE TRAP AIR SAMPLING RECORD



000473110

Page 1 of 1

BioMax Environmental
775 San Pablo Ave.
Pinole, CA 94564

www.biomaxenvironmental.com

Phone: (510) 724-3100

Fax: (510) 724-3145

biomaxenv@aol.com

Location: Break Room 707 450 N. Street Sacramento, CA	Client: DGS Project #: 100108-01
Date: 10/1/08 Collected by: M.A. Falkobles Signature: 	Laboratory: EML Labs Req. Turn Around: 24 HR Analysis: <u>Fungal</u> <u>Particulate ID</u> <u>with Quantification</u>

Sample Number	Time	Location/Desc	Temp/RH
14226774	1110	Ambient Front Entry	77 / 28.5%
14226731	1125	7th Floor Hallway (OC)	73 / 29%
14226603	1135	Rm 707 Break Room (I.C.)	74 / 29%
14226653	1150	Rm 706 Supply Rm (I.C.)	73 / 29%
14226607	1205	Ambient Post NE Bld Corner	79 / 28%
Total Sample Time (min): 5	Flow Rate (l/min): 15	Total Sample Volume (liters): 75	Ambient Conditions: Clear - mild 0-5 w/sky Comments:

Please sign this form below acknowledging sample receipt and return executed form with laboratory reports. Fax, send, e-mail results to BioMax Environmental at (510) 724-3145 biomaxenv@aol.com
Other Instructions: _____

Relinquished by: Method of Transportation: Fed Ex Time/Date Sent: 4:00 10/1/08	Received By: Ann Morrissey Time/Date Received: 10-2-08 9:20
---	---



EMLab P&K

Report for:

Mr. Michael Polkabila
Biomax Environmental
775 San Pablo Ave.
Pinole, CA 94564

Regarding: Project: 100108-01
EML ID: 473110

Approved by:

Lab Manager
Dr. Kamashwaran Ramanathan

Dates of Analysis:
Spore trap analysis: 10-03-2008

Project SOPs: Spore trap analysis (I100000)

This coversheet is included with your report in order to comply with AIHA and ISO accreditation requirements.

For clarity, we report the number of significant digits as calculated; but, due to the nature of this type of biological data, the number of significant digits that is used for interpretation should generally be one or two. All samples were received in acceptable condition unless noted in the Report Comments portion in the body of the report. Due to the nature of the analyses performed, field blank corrections of results is not a standard practice. The results relate only to the items tested.

EMLab P&K ("the Company") shall have no liability to the client or the client's customer with respect to decisions or recommendations made, actions taken or courses of conduct implemented by either the client or the client's customer as a result of or based upon the Test Results. In no event shall the Company be liable to the client with respect to the Test Results except for the Company's own willful misconduct or gross negligence nor shall the Company be liable for incidental or consequential damages or lost profits or revenues to the fullest extent such liability may be disclaimed by law, even if the Company has been advised of the possibility of such damages, lost profits or lost revenues. In no event shall the Company's liability with respect to the Test Results exceed the amount paid to the Company by the client therefor.

Document Number: 200091 - Revision Number: 5

Client: Biomax Environmental
C/O: Mr. Michael Polkabla
Re: 100108-01

Date of Sampling: 10-01-2008
Date of Receipt: 10-02-2008
Date of Report: 10-03-2008

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	14226774: Ambient front entry		14226731: 7th floor hallway, OC		14226603: Room 707 break room, IC		14226653: Room 706 supply room, IC		14226607: Ambient post NE bld corner	
Comments (see below)	None		None		None		None		None	
Lab ID-Version‡:	2087796-1		2087797-1		2087798-1		2087799-1		2087800-1	
	raw ct.	spores/m3	raw ct.	spores/m3	raw ct.	spores/m3	raw ct.	spores/m3	raw ct.	spores/m3
Alternaria	2	27							4	53
Arthrinium										
Ascospores*	1	53	1	53						
Aureobasidium										
Basidiospores*	5	267	1	53	2	107	1	53	2	107
Bipolaris/Drechslera group	1	13								
Botrytis										
Chaetomium	9	120	1	13					6	80
Cladosporium	10	533							28	1,490
Curvularia										
Epicoccum										
Fusarium										
Myrothecium										
Nigrospora	1	13							2	27
Other brown					1	13	1	13	1	13
Other colorless									1	13
Penicillium/Aspergillus types†	2	107							5	267
Pithomyces										
Rusts*									1	13
Smuts*, Periconia, Myxomycetes*	6	80			3	40			11	147
Stachybotrys										
Stemphylium										
Torula										
Ulocladium										
Zygomycetes										
Background debris (1-4+)††	3+		2+		3+		3+		3+	
Hyphal fragments/m3	93		< 13		13		13		67	
Pollen/m3	40		< 13		< 13		< 13		67	
Skin cells (1-4+)	< 1+		1+		2+		1+		< 1+	
Sample volume (liters)	75		75		75		75		75	
§ TOTAL SPORE/m3		1,200		120		160		66		2,200

Comments:

* Most of these spore types are not seen with culturable methods (Andersen sampling), although some may appear as non-sporulating fungi. Most of the basidiospores are "mushroom" spores while the rusts and smuts are plant pathogens.
 † The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.
 †† Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.
 The Limit of Detection is the product of a raw count of 1 and 100 divided by the percent read. The analytical sensitivity (counts/m3) is the product of the Limit of Detection and 1000 divided by the sample volume.
 ‡ A "Version" greater than 1 indicates amended data.
 § Total Spores/m3 has been rounded to two significant figures to reflect analytical precision.
 TestAmerica Environmental Microbiology Laboratory, Inc.

Client: Biomax Environmental
C/O: Mr. Michael Polkabla
Re: 100108-01Date of Sampling: 10-01-2008
Date of Receipt: 10-02-2008
Date of Report: 10-03-2008**MoldRANGE™: Extended Outdoor Comparison**
Outdoor Location: 14226774, Ambient front entry

Fungi Identified	Outdoor data	Typical Outdoor Data by Date†				Typical Outdoor Data by Location‡			
		Month: October				State: CA			
	spores/m3	low	med	high	freq %	low	med	high	freq %
Generally able to grow indoors*									
Alternaria	27	7	33	430	61	7	27	210	59
Bipolaris/Drechslera group	13	7	13	200	23	7	13	120	13
Chaetomium	120	7	13	130	14	7	13	120	19
Cladosporium	533	53	800	11,000	97	53	640	6,400	98
Curvularia	-	7	27	680	27	7	13	200	7
Nigrospora	13	7	13	240	25	7	13	160	8
Other brown	-	7	13	110	36	7	13	80	37
Other colorless	-	7	13	210	6	7	13	93	6
Penicillium/Aspergillus types	107	27	270	3,500	87	38	210	2,500	87
Stachybotrys	-	7	13	200	3	7	13	280	5
Torula	-	7	13	200	13	7	13	150	13
Seldom found growing indoors**									
Ascospores	53	13	160	4,600	81	13	110	1,800	72
Basidiospores	267	27	480	20,000	96	13	230	6,700	94
Rusts	-	7	22	410	29	7	13	250	28
Smuts, Periconia, Myxomycetes	80	8	53	880	79	8	40	480	71
TOTAL SPORES/M3	1,213								

† The Typical Outdoor Data by Date represents the typical outdoor spore levels across North America for the month indicated. The last column represents the frequency of occurrence. The low, medium, and high values represent the 2.5, 50, and 97.5 percentile values of the spore type when it is detected. For example, if the frequency of occurrence is 63% and the low value is 53, it would mean that the given spore type is detected 63% of the time and, when detected, 2.5% of the time it is present in levels above the detection limit and below 53 spores/m³. These values are updated periodically, and if enough data is not available to make a statistically meaningful assessment, it is indicated with a dash.

‡ The Typical Outdoor Data by Location represents the typical outdoor spore levels for the region indicated for the entire year. As with the Typical Outdoor Data by Date, the four columns represent the frequency of occurrence and the typical low, medium, and high concentration values for the spore type indicated. These values are updated periodically, and if enough data is not available to make a statistically meaningful assessment, it is indicated with a dash.

*The spores in this category are generally capable of growing on wet building materials in addition to growing outdoors. Building related growth is dependent upon the fungal type, moisture level, type of material, and other factors. *Cladosporium* is one of the predominant spore types worldwide and is frequently present in high numbers. *Penicillium/Aspergillus* species colonize both outdoor and indoor wet surfaces rapidly and are very easily dispersed. Other genera are usually present in lesser numbers.

**These fungi are generally not found growing on wet building materials. For example, the rusts and smuts are obligate plant pathogens. However, in each group there are notable exceptions. For example, agents of wood decay are members of the basidiomycetes and high counts of a single morphological type of basidiospore on an inside sample should be considered significant.

Interpretation of the data contained in this report is left to the client or the persons who conducted the field work. This report is provided for informational and comparative purposes only and should not be relied upon for any other purpose. "Typical outdoor data" are based on the results of the analysis of samples delivered to and analyzed by EMLab P&K and assumptions regarding the origins of those samples. Sampling techniques, contaminants infecting samples, unrepresentative samples and other similar or dissimilar factors may affect these results. In addition, EMLab P&K may not have received and tested a representative number of samples for every region or time period. EMLab P&K hereby disclaims any liability for any and all direct, indirect, punitive, incidental, special or consequential damages arising out of the use or interpretation of the data contained in, or any actions taken or omitted in reliance upon, this report.

Client: Biomax Environmental
C/O: Mr. Michael Polkabla
Re: 100108-01Date of Sampling: 10-01-2008
Date of Receipt: 10-02-2008
Date of Report: 10-03-2008**MoldRANGE™: Extended Outdoor Comparison****Outdoor Location: 14226607, Ambient post NE bld corner**

Fungi Identified	Outdoor data	Typical Outdoor Data by Date†				Typical Outdoor Data by Location‡			
		Month: October				State: CA			
	spores/m3	low	med	high	freq %	low	med	high	freq %
Generally able to grow indoors*									
Alternaria	53	7	33	430	61	7	27	210	59
Bipolaris/Drechslera group	-	7	13	200	23	7	13	120	13
Chaetomium	80	7	13	130	14	7	13	120	19
Cladosporium	1,490	53	800	11,000	97	53	640	6,400	98
Curvularia	-	7	27	680	27	7	13	200	7
Nigrospora	27	7	13	240	25	7	13	160	8
Other brown	13	7	13	110	36	7	13	80	37
Other colorless	13	7	13	210	6	7	13	93	6
Penicillium/Aspergillus types	267	27	270	3,500	87	38	210	2,500	87
Stachybotrys	-	7	13	200	3	7	13	280	5
Torula	-	7	13	200	13	7	13	150	13
Seldom found growing indoors**									
Ascospores	-	13	160	4,600	81	13	110	1,800	72
Basidiospores	107	27	480	20,000	96	13	230	6,700	94
Rusts	13	7	22	410	29	7	13	250	28
Smuts, Periconia, Myxomycetes	147	8	53	880	79	8	40	480	71
TOTAL SPORES/M3	2,210								

† The Typical Outdoor Data by Date represents the typical outdoor spore levels across North America for the month indicated. The last column represents the frequency of occurrence. The low, medium, and high values represent the 2.5, 50, and 97.5 percentile values of the spore type when it is detected. For example, if the frequency of occurrence is 63% and the low value is 53, it would mean that the given spore type is detected 63% of the time and, when detected, 2.5% of the time it is present in levels above the detection limit and below 53 spores/m³. These values are updated periodically, and if enough data is not available to make a statistically meaningful assessment, it is indicated with a dash.

‡ The Typical Outdoor Data by Location represents the typical outdoor spore levels for the region indicated for the entire year. As with the Typical Outdoor Data by Date, the four columns represent the frequency of occurrence and the typical low, medium, and high concentration values for the spore type indicated. These values are updated periodically, and if enough data is not available to make a statistically meaningful assessment, it is indicated with a dash.

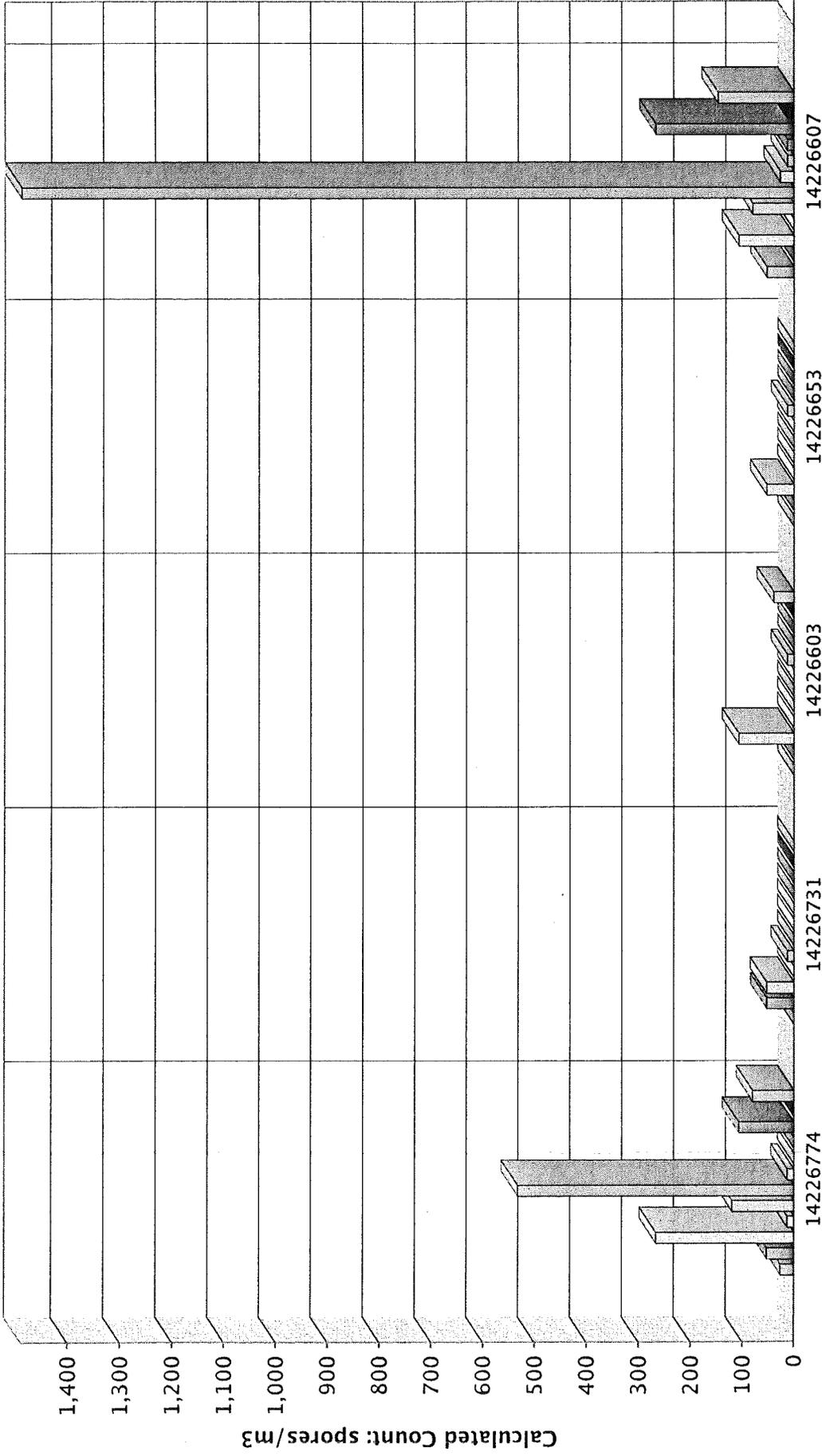
*The spores in this category are generally capable of growing on wet building materials in addition to growing outdoors. Building related growth is dependent upon the fungal type, moisture level, type of material, and other factors. *Cladosporium* is one of the predominant spore types worldwide and is frequently present in high numbers. *Penicillium/Aspergillus* species colonize both outdoor and indoor wet surfaces rapidly and are very easily dispersed. Other genera are usually present in lesser numbers.

**These fungi are generally not found growing on wet building materials. For example, the rusts and smuts are obligate plant pathogens. However, in each group there are notable exceptions. For example, agents of wood decay are members of the basidiomycetes and high counts of a single morphological type of basidiospore on an inside sample should be considered significant.

Interpretation of the data contained in this report is left to the client or the persons who conducted the field work. This report is provided for informational and comparative purposes only and should not be relied upon for any other purpose. "Typical outdoor data" are based on the results of the analysis of samples delivered to and analyzed by EMLab P&K and assumptions regarding the origins of those samples. Sampling techniques, contaminants infecting samples, unrepresentative samples and other similar or dissimilar factors may affect these results. In addition, EMLab P&K may not have received and tested a representative number of samples for every region or time period. EMLab P&K hereby disclaims any liability for any and all direct, indirect, punitive, incidental, special or consequential damages arising out of the use or interpretation of the data contained in, or any actions taken or omitted in reliance upon, this report.

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

- Alternaria Ascospores Basidiospores Bipolaris/Drechslera group Chaetomium Cladosporium Nigrospora
- Other brown Other colorless Penicillium/Aspergillus types Rusts Smuts, Periconia, Myxomycetes



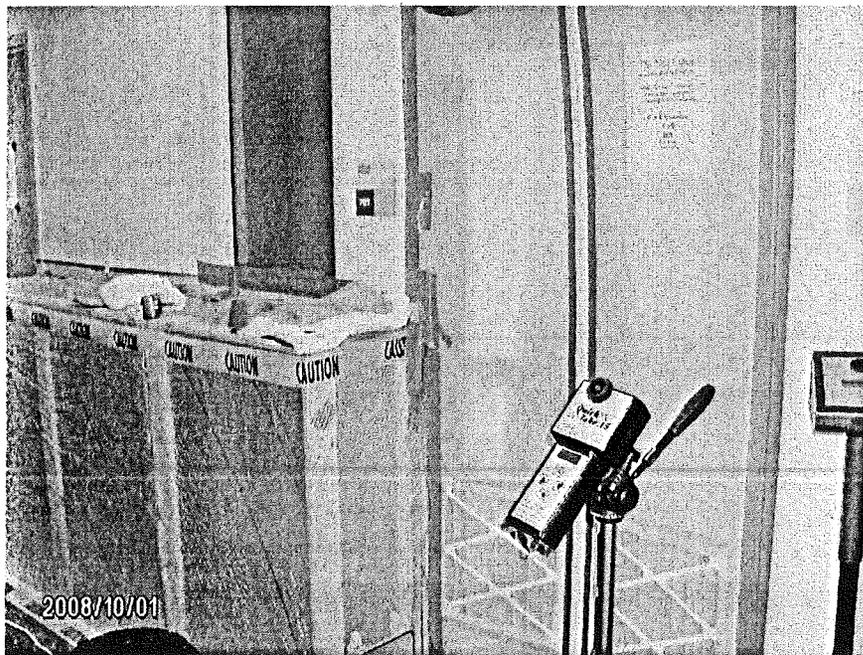
Comments:

Note: Graphical output may understate the importance of certain "marker" genera.
TestAmerica Environmental Microbiology Laboratory, Inc.

[Click here for color photos](#)



- 1) Image of hallway within BOE occupied working spaces located adjacent to 707 Break Room Area at time of assessment.



- 2) Image of Break Room 707 containment as viewed from occupied hallway at time of clearance assessment.



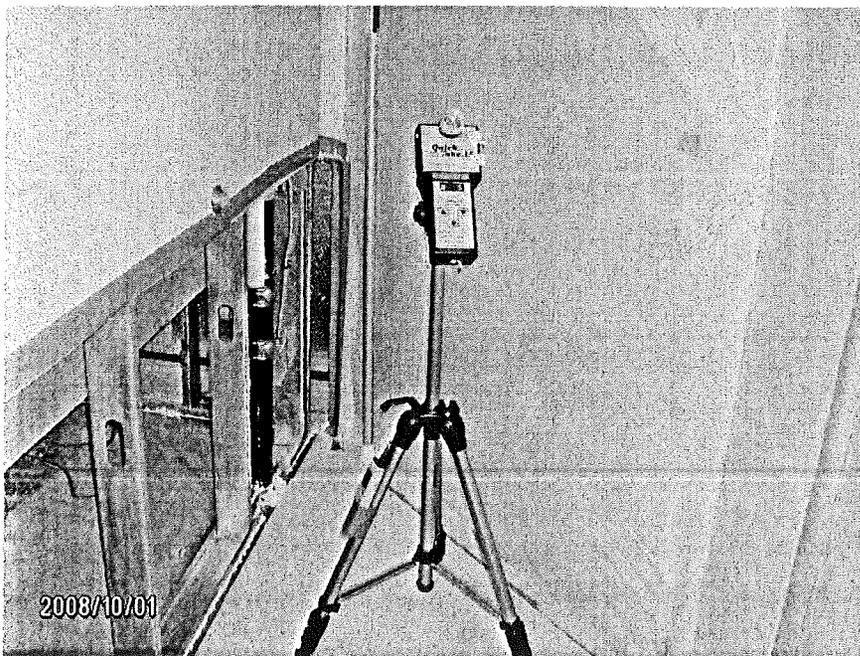
3) Image of air sampling equipment and wall removal within Break Room 707 area at time of assessment.



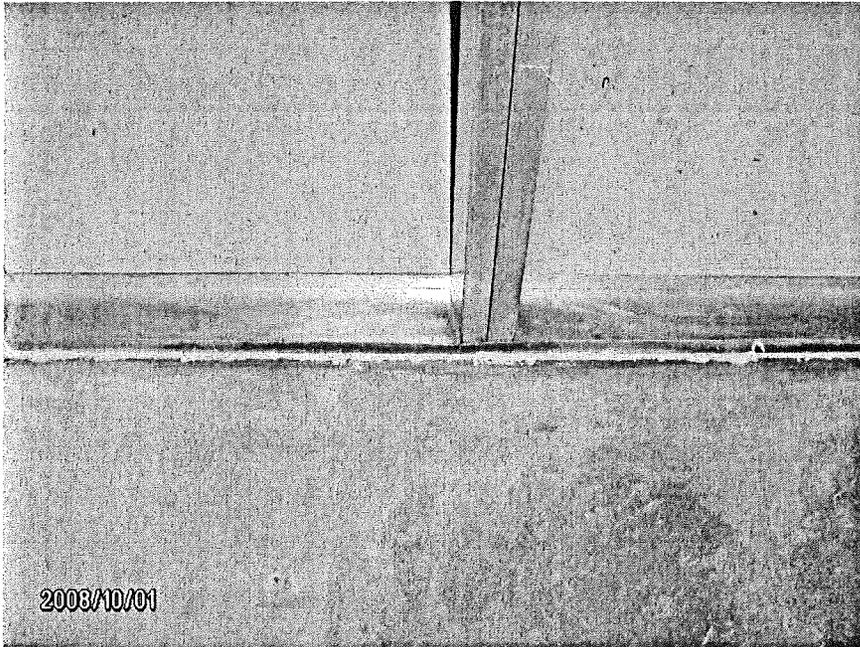
4) Image of exposed wall cavity, exposed plumbing, and delineation of wallboard removal within Break Room 707 at time of assessment.



5) Image of interior containment entry chamber and flex exhaust HEPA ducting within Break Room 707 at time of assessment.



6) Image of wallboard removal delineation within adjacent supply room area at time of assessment.



- 7) Close up image of exposed floor sill metal framing following mitigative cleaning as viewed from Break Room 707 at time of assessment.



- 8) Image of exposed metal wall framing and plumbing systems as viewed from adjacent supply room at time of assessment.



- 9) Close-up image of HTI technician collecting surface sample of metal framing surface within supply room at time of assessment.



- 10) Image of ambient air sampling equipment location performed at conclusion of interior sampling activities.

Client: Biomax Environmental
C/O: Mr. Michael Polkabla
Re: 100108-01

Date of Sampling: 10-01-2008
Date of Receipt: 10-02-2008
Date of Report: 10-03-2008

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	14226774: Ambient front entry		14226731: 7th floor hallway, OC		14226603: Room 707 break room, IC		14226653: Room 706 supply room, IC		14226607: Ambient post NE bld corner	
Comments (see below)	None		None		None		None		None	
Lab ID-Version‡:	2087796-1		2087797-1		2087798-1		2087799-1		2087800-1	
	raw ct.	spores/m3	raw ct.	spores/m3	raw ct.	spores/m3	raw ct.	spores/m3	raw ct.	spores/m3
Alternaria	2	27							4	53
Arthrinium										
Ascospores*	1	53	1	53						
Aureobasidium										
Basidiospores*	5	267	1	53	2	107	1	53	2	107
Bipolaris/Drechslera group	1	13								
Botrytis										
Chaetomium	9	120	1	13					6	80
Cladosporium	10	533							28	1,490
Curvularia										
Epicoccum										
Fusarium										
Myrothecium										
Nigrospora	1	13							2	27
Other brown					1	13	1	13	1	13
Other colorless									1	13
Penicillium/Aspergillus types†	2	107							5	267
Pithomyces										
Rusts*									1	13
Smuts*, Periconia, Myxomycetes*	6	80			3	40			11	147
Stachybotrys										
Stemphylium										
Torula										
Ulocladium										
Zygomycetes										
Background debris (1-4+)††	3+		2+		3+		3+		3+	
Hyphal fragments/m3	93		< 13		13		13		67	
Pollen/m3	40		< 13		< 13		< 13		67	
Skin cells (1-4+)	< 1+		1+		2+		1+		< 1+	
Sample volume (liters)	75		75		75		75		75	
§ TOTAL SPORE/m3		1,200		120		160		66		2,200

Comments:

* Most of these spore types are not seen with culturable methods (Andersen sampling), although some may appear as non-sporulating fungi. Most of the basidiospores are "mushroom" spores while the rusts and smuts are plant pathogens.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

†† Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.

The Limit of Detection is the product of a raw count of 1 and 100 divided by the percent read. The analytical sensitivity (counts/m3) is the product of the Limit of Detection and 1000 divided by the sample volume.

‡ A "Version" greater than 1 indicates amended data.

§ Total Spores/m3 has been rounded to two significant figures to reflect analytical precision.

TestAmerica Environmental Microbiology Laboratory, Inc.

Client: Biomax Environmental
C/O: Mr. Michael Polkabila
Re: 100108-01Date of Sampling: 10-01-2008
Date of Receipt: 10-02-2008
Date of Report: 10-03-2008**MoldRANGE™: Extended Outdoor Comparison****Outdoor Location: 14226774, Ambient front entry**

Fungi Identified	Outdoor data	Typical Outdoor Data by Date†				Typical Outdoor Data by Location‡			
		Month: October				State: CA			
	spores/m3	low	med	high	freq %	low	med	high	freq %
Generally able to grow indoors*									
Alternaria	27	7	33	430	61	7	27	210	59
Bipolaris/Drechslera group	13	7	13	200	23	7	13	120	13
Chaetomium	120	7	13	130	14	7	13	120	19
Cladosporium	533	53	800	11,000	97	53	640	6,400	98
Curvularia	-	7	27	680	27	7	13	200	7
Nigrospora	13	7	13	240	25	7	13	160	8
Other brown	-	7	13	110	36	7	13	80	37
Other colorless	-	7	13	210	6	7	13	93	6
Penicillium/Aspergillus types	107	27	270	3,500	87	38	210	2,500	87
Stachybotrys	-	7	13	200	3	7	13	280	5
Torula	-	7	13	200	13	7	13	150	13
Seldom found growing indoors**									
Ascospores	53	13	160	4,600	81	13	110	1,800	72
Basidiospores	267	27	480	20,000	96	13	230	6,700	94
Rusts	-	7	22	410	29	7	13	250	28
Smuts, Periconia, Myxomycetes	80	8	53	880	79	8	40	480	71
TOTAL SPORES/M3	1,213								

† The Typical Outdoor Data by Date represents the typical outdoor spore levels across North America for the month indicated. The last column represents the frequency of occurrence. The low, medium, and high values represent the 2.5, 50, and 97.5 percentile values of the spore type when it is detected. For example, if the frequency of occurrence is 63% and the low value is 53, it would mean that the given spore type is detected 63% of the time and, when detected, 2.5% of the time it is present in levels above the detection limit and below 53 spores/m³. These values are updated periodically, and if enough data is not available to make a statistically meaningful assessment, it is indicated with a dash.

‡ The Typical Outdoor Data by Location represents the typical outdoor spore levels for the region indicated for the entire year. As with the Typical Outdoor Data by Date, the four columns represent the frequency of occurrence and the typical low, medium, and high concentration values for the spore type indicated. These values are updated periodically, and if enough data is not available to make a statistically meaningful assessment, it is indicated with a dash.

*The spores in this category are generally capable of growing on wet building materials in addition to growing outdoors. Building related growth is dependent upon the fungal type, moisture level, type of material, and other factors. *Cladosporium* is one of the predominant spore types worldwide and is frequently present in high numbers. *Penicillium/Aspergillus* species colonize both outdoor and indoor wet surfaces rapidly and are very easily dispersed. Other genera are usually present in lesser numbers.

**These fungi are generally not found growing on wet building materials. For example, the rusts and smuts are obligate plant pathogens. However, in each group there are notable exceptions. For example, agents of wood decay are members of the basidiomycetes and high counts of a single morphological type of basidiospore on an inside sample should be considered significant.

Interpretation of the data contained in this report is left to the client or the persons who conducted the field work. This report is provided for informational and comparative purposes only and should not be relied upon for any other purpose. "Typical outdoor data" are based on the results of the analysis of samples delivered to and analyzed by EMLab P&K and assumptions regarding the origins of those samples. Sampling techniques, contaminants infecting samples, unrepresentative samples and other similar or dissimilar factors may affect these results. In addition, EMLab P&K may not have received and tested a representative number of samples for every region or time period. EMLab P&K hereby disclaims any liability for any and all direct, indirect, punitive, incidental, special or consequential damages arising out of the use or interpretation of the data contained in, or any actions taken or omitted in reliance upon, this report.

EMLab P&K

1150 Bayhill Drive, Suite 100, San Bruno, CA 94066
 (650) 829-5800 Fax (650) 829-5852 www.emlab.com

Client: Biomax Environmental
 C/O: Mr. Michael Polkabila
 Re: 100108-01

Date of Sampling: 10-01-2008
 Date of Receipt: 10-02-2008
 Date of Report: 10-03-2008

MoldRANGE™: Extended Outdoor Comparison

Outdoor Location: 14226607, Ambient post NE bld corner

Fungi Identified	Outdoor data	Typical Outdoor Data by Date†				Typical Outdoor Data by Location‡			
		Month: October				State: CA			
	spores/m3	low	med	high	freq %	low	med	high	freq %
Generally able to grow indoors*									
Alternaria	53	7	33	430	61	7	27	210	59
Bipolaris/Drechslera group	-	7	13	200	23	7	13	120	13
Chaetomium	80	7	13	130	14	7	13	120	19
Cladosporium	1,490	53	800	11,000	97	53	640	6,400	98
Curvularia	-	7	27	680	27	7	13	200	7
Nigrospora	27	7	13	240	25	7	13	160	8
Other brown	13	7	13	110	36	7	13	80	37
Other colorless	13	7	13	210	6	7	13	93	6
Penicillium/Aspergillus types	267	27	270	3,500	87	38	210	2,500	87
Stachybotrys	-	7	13	200	3	7	13	280	5
Torula	-	7	13	200	13	7	13	150	13