

TAB 3

BioMax Environmental

Environmental Consulting and Industrial Hygiene Services

March 31st, 2008

Mr. Doug Button
Deputy Director
Real Estate Services Division
707 Third Street - 8th Floor
West Sacramento, CA 95605

Post Mitigation Assessment Report
Department of General Services Board of Equalization Building
450 N. Street - 23rd Floor South and East Containment Areas
Sacramento, California

Mr. Button,

BioMax Environmental, LLC (BioMax) is pleased to provide The Department of General Services (DGS) with this letter summary report detailing BioMax's findings and recommendations pertaining to our post mitigation microbial inspection and sampling assessment services provided within the 23rd Floor containment areas present within south and eastern quadrants of the Board of Equalization (BOE) building (subject building) located at 450 N Street, Sacramento, California. BioMax understands that these post mitigation microbial inspection and clearance sampling assessment services were contracted with BioMax, at your request, in an effort to review and verify the successful completion of microbial mitigative efforts performed by your restoration contractor, JLS Environmental, Inc., within the previously identified mold damaged and moisture impacted areas within the 23rd floor of the subject building. These microbial clearance assessment services were requested following the previous discovery of moisture and mold damage materials primarily identified within the noted southern and eastern perimeter wall structures and areas of the subject building.

Hence, these post mitigation assessment services are intended to assess the current site conditions wherein mitigative activities were performed to address prior moisture and mold related damages and impacts. Observations, findings, and recommendations pertaining to BioMax's preliminary site assessments of the subject areas have been summarized within our previous Letter Summary Assessment Reports dated December 10th, 2007, December 13th, 2007, January 31st, 2008, and February 8th, 2008. These previous reports may be referenced for further historical information, as necessary. Following the completion of prescribed mitigative activities performed by your selected contractor, JLS, Mr. Michael A. Polkabila, CIH, REA of BioMax performed a post mitigation site inspection and sampling assessment within the affected areas of the subject building areas as noted below. BioMax's findings and conclusions pertaining to our post mitigation sampling assessment are summarized herein.

These post mitigation microbial clearance assessment services, thereby, are intended to provide a professional evaluation supported by technical sampling data verifying physical conditions wherein the successful completion of microbial removal and decontamination within the affected areas has been achieved.

SITE OBSERVATIONS

Site inspection and post mitigation assessment sampling activities were performed on Monday, March 10th, 2008 wherein site access into contained and non-contained areas was facilitated by Mr. Rick Boggs of JLS. On this day, Mr. Michael A. Polkabila, CIH, REA of BioMax performed a visual site inspection of each of the containment system barriers associated with the southern and eastern quadrant areas and collected a series of airborne samples within and surrounding the areas of concern the subject structures as noted below.

On-site inspection and clearance sampling assessment activities were performed by Mr. Michael A. Polkabila, CIH, REA, of BioMax in accordance with currently recognized microbial assessment and sampling guideline procedures. Mr. Polkabila has been certified in the Comprehensive Practice of Industrial Hygiene by the American Board of Industrial Hygiene and holds the right to the designation "Certified Industrial Hygienist" (CIH) under certification number CP 7104. Mr. Polkabila is also certified by the California Environmental Protection Agency (Cal/EPA) as a Class I Registered Environmental Assessor (REA) under Cal/EPA certification number 05011. A summary of significant notations and observations gathered during BioMax's site inspection and clearance assessment within the subject facility are compiled as follows:

1. At the time of our site inspection and clearance sampling assessment performed on March 10th, 2008 ambient outdoor conditions both prior to and following our interior assessment consisted of sunny and cold conditions with an outdoor temperatures range between 57 and 77 degrees F and relative humidity range between 30 and 65 %. Predominant winds were noted at approximately 0-5 knots from the northwesterly direction at the time of our assessment. Interior environmental conditions within the sampled 23rd Floor areas consisted of a temperature range between 72 and 86 degrees F with relative humidity range of 25 to 43 percent.
2. Interior containments whereby microbial mitigative activities were performed included the work areas noted associated with the building perimeter walls along the southern and eastern quadrants of the 23rd floor of the subject building. Within these areas, ceiling plastic barriers erected by JLS were erected and maintained within the impacted areas as noted on the "as built" construction site floor diagram documents. Such floor plan diagrams may be reviewed for further reference as supplied by the site mitigation contractor, JLS, as necessary. Based on our inspection and review of observations within and surrounding the noted containment areas, BioMax concluded that such systems provided evidence of appropriate control barriers and site protections during the mitigative activity through our post mitigation assessment.

3. Based on our post mitigation inspection within and surrounding the containment areas noted above, BioMax documented the absence of visible interior indications of elevated residual moisture and/or microbial indicators (such as staining, delamination, etc.) within the remaining exposed interior walls, wall framing, and wall cavities following the performance of mitigative measures. Utilization of a TraMex hand-held inductive moisture meter indicated normal moisture content within all remaining walls and building materials inspected within each of the sampled containment areas at the time of our assessment.
4. As noted within BioMax's previously referenced reports, the primary affected areas of visible moisture damage previously identified primarily included moisture and mold damaged wallboard materials and adjacent office furnishings located within the perimeter wall areas within the southern and eastern quadrants of the 23rd floor area.
5. Containment system encompassing each of the interior affected areas were observed and verified during multiple inspection dates under appropriate posting and negative pressure differential. Worker and equipment entry and exit chambers comprised of a series of zippered plastic access doorways were also observed attached to the noted containment barriers consistent with BioMax's previous written mitigation protocols.
6. As prescribed, all identified affected interior wallboard building materials had been removed from each of the affected areas exposing interior wall cavity framing (metal) as well as underlayment wallboard siding materials once present within the impacted containment areas. Upon post mitigation inspection, all remaining exposed building materials associated with the interior wall systems exhibited no significant staining and/or elevated mold growth following the completion of prescribed physical mold removal and chemical decontamination procedures performed by the selected mitigation contractor on the surfaces of such exposed building materials.
7. In conjunction with our visual inspection, BioMax collected series airborne samples within and outside each of the containment areas noted below for subsequent comparative analysis. Such samples collected within and surrounding each the interior containment areas were performed in an effort to identify and quantify the presence of any potential significant fugitive airborne mold spores present within (and surrounding) the containment systems following the completion of the prescribed mitigative effort.
8. BioMax also collected a series of digital images during these post mitigative inspection and sampling assessment activities to document the conditions and significant site observations gathered at this time. Such images are provided as an attachment to this summary report for further reference, as necessary.

SAMPLING PROCEDURES

On-site inspection and sampling assessment activities were conducted by Mr. Michael A. Polkabila, CIH, REA, of BioMax Environmental on March 10th, 2008. All sampling equipment, supplies, calibration materials, and collection media were provided by BioMax as part of the performance of this scope of work. Sample collection procedures and methods were performed using aseptic sampling methods following techniques prescribed by the contracted analytical laboratory.

Spore Trap Airborne Microbial and Particulate Sampling:

The collection of airborne Spore Trap microbial samples was achieved using Zefon Air-O-Cell sampling cassette collection devices placed in each of the areas identified in the tables below. Airborne Spore Trap samples were collected within and outside each of the containment areas at a height of approximately four feet above ground level using a tripod mounted Quick Take 15 air sampling pump manufactured by SKC. Samples were collected at a calibrated flow rate of 15 liters per minute for a total of five minutes per sample. Resultant total sample volumes, therefore, corresponded to 75 liters collected for each sample. Field calibration of the SKC air sampling pump was conducted and recorded prior to and following sampling activities using a field rotometer device calibrated with a Bios Drycal primary standard flow meter. All spore trap air sampling and analytical procedures were performed in accordance with prescribed manufacturer guidelines as well as applicable professional certified industrial hygiene indoor air quality microbial investigation procedures and certified industrial hygiene practices.

Additional exterior samples were also similarly collected and analyzed during the collection of interior samples in an effort to identify and quantify normal background microbial taxa (types), rank order, and corresponding airborne spore levels present at the time of this assessment. Efforts were made in the collection of airborne samples to capture such samples during conditions which were representative of the environmental conditions and activities within this floor so as to closely approximate air handling system ventilation conditions present within each of the subject areas at the time of this assessment. Sampling collection activities performed during this study included the collection of identifiable airborne microbial contaminants within the representative areas noted below:

Table 1. 23 South Area Airborne Spore Trap Sampling Locations:

Air Sample Number	Spore Trap Air Sampling Location
13430672	Ambient outside location (Main Entry Level)
13430716	Ambient outside sample from 23 rd Floor west balcony
13430719	Ambient outside sample from 12 th Floor SE balcony
13430730	Room 2311 (outside containment)

Air Sample Number	Spore Trap Air Sampling Location
13430691	Room 2305 south west side (inside containment)
13430698	Room 2305 south east side (inside containment)
13430693	Office 2307 (inside containment)
13430689	Equipment corridor (outside containment)

Table 2. 23 East Area Airborne Spore Trap Sampling Locations:

Air Sample Number	Spore Trap Air Sampling Location
13430731	Room 2339 (inside containment)
13430665	Room 2338 (inside containment)
13430726	Room 2337 (inside containment)
13430703	Room 2336 (inside containment)
13430700	Hallway (outside containment)
13430734	Room 2335 (inside containment)
13430684	Room 2334 (inside containment)
13430674	Room 2332 (inside containment)
13430676	Ambient outside sample from 23 rd Floor west balcony
13430688	Ambient outside location (Main Entry Level)

At the conclusion of sampling activities, preparation and shipping of the collected samples were accomplished in accordance with standard industrial hygiene chain of custody (COC) documentation procedures and quality assurance/quality control practices. Once collected, labeled, and recorded, all samples were double sealed within airtight plastic Ziploc shipping containers and transported via Federal Express Priority Mail to Environmental Microbial Laboratories (EMLabs) in San Bruno, California. EMLabs holds current applicable analytical accreditation and specializes in microbial analytical procedures. Sampling and chain of custody records are provided as an attachment to this letter report for further reference.

ANALYTICAL FINDINGS AND CONCLUSIONS

Airborne Spore Trap Findings:

Laboratory analytical methods for the identification and enumeration of microbial (mold) taxa and particulate contaminants were conducted in accordance with prescribed analytical procedures and quality control/assurance measures. Original laboratory results including the enumeration of recognizable microbial spore and particulate types are also attached to this letter report for further detail. Analytical comments provided by the microbial laboratory regarding relative levels are noted as a semi-quantitative assessment based on historical and regional data. BioMax has also provided a copy of a current analytical interpretive guideline as an attachment to this report for further reference. A summary of airborne Spore Trap microbial (mold) and particulate findings pertaining to each of the subject areas are presented in Table 3 and 4 below:

Table 3. Summary of Airborne Microbial and Particulate Findings – 23 South

Location Desc.	Total Mold Spores (Cts/m3)	Background Debris (scale of 1-4)	Skin Cell Fragments (scale of 1-4)
Ambient outside location (Main Entry Level)	2,080	3+	None
Ambient outside sample from 23 rd Floor west balcony	1,556	3+	None
Ambient outside sample from 12 th Floor SE balcony	2,850	3+	None
Room 2311 (outside containment)	587	3+	None
Room 2305 south west side (inside containment)	746	2+	<1+
Room 2305 south east side (inside containment)	107	3+	1+
Office 2307 (inside containment)	160	3+	1+
Equipment corridor (outside containment)	599	3+	1+

Table 4. Summary of Airborne Microbial and Particulate Findings – 23 South

Location Desc.	Total Mold Spores (Cts/m3)	Background Debris (scale of 1-4)	Skin Cell Fragments (scale of 1-4)
Room 2339 (inside containment)	214	2+	<1+
Room 2338 (inside containment)	67	2+	<1+
Room 2337 (inside containment)	53	2+	<1+
Room 2336 (inside containment)	53	2+	<1+
Hallway (outside containment)	66	2+	<1+
Room 2335 (inside containment)	53	2+	<1+
Room 2334 (inside containment)	79	2+	<1+
Room 2332 (inside containment)	53	2+	<1+
Ambient outside sample from 23 rd Floor west balcony	1,199	2+	<1+
Ambient outside location (Main Entry Level)	1,226	3+	<1+

The analytical findings presented in Tables 3 and 4 clearly indicate the presence of significantly lower concentrations of microbial (mold) spores measured within each of the interior samples collected both within and surrounding the subject areas when compared to the levels currently measured within the samples collected from the corresponding ambient outside environment. Analytical findings also indicate similar fungal taxa distribution (mold types) and rank order (predominant taxa) of molds identified within the mitigated areas as well as the adjacent worker equipment staging area samples (areas noted as “outside containment”). Analytical findings also indicated the absence of significantly elevated levels of unique fungal taxa present within these

noted adjacent support areas accessed by non-mitigation contractor staff and inspectors. Particularly worthy of note, was the absence of elevated levels of hydrophilic (moisture loving) mold taxa such as Penicillium, Aspergillus, and Stachybotrys which were previously identified present within the surface samples collected from within the noted containment areas as summarized in previous historical findings and assessment reports.

Although there are currently no regulatory standards or limits pertaining to allowable airborne fungal concentrations (for any mold taxa) present in indoor environments, there is a general consensus among indoor air quality experts that microbial contamination found within "typical healthy" living spaces are generally similar in kind and present at levels which are below those found in the corresponding native outside environment. BioMax believes that the absence of elevated moisture, absence of significant visible residual mold, and relatively fewer total airborne mold levels with typical taxa and rank order distribution following mitigative clean-up activities are consistent with these generally acceptable conditions. BioMax, therefore, believes that these findings provide reasonable evidence indicating that current microbial clean-up measures have successfully mitigated and contained mold contamination within previously affected areas and previously affected materials to normal representative levels.

Based on these findings, BioMax believes that the current site conditions present within the mitigated areas as well as the corresponding analytical data collected and evaluated, following the performance of the recommended mitigative procedures, meets the clearance criteria established for these activities as presented in BioMax's Post Mitigation Clearance Assessment Protocols dated February 15th, 2008 as reviewed and approved by BOE's environmental consultant, Hygientech. Therefore, BioMax believes that achievement of such criteria warrants our determination and recommendation that the previously impacted areas may be considered acceptable for reconstruction at this time.

Airborne Particulate Findings:

Analytical findings pertaining to the levels of airborne particulates debris identified within the collected air samples within and surrounding the previously impacted areas also provide reasonable evidence indicating that current particulate clean-up and mitigative control measures have successfully controlled and contained particulate debris within the identified containment areas.

Although, there are similarly no currently applicable regulatory standards pertaining to allowable particulate levels with which to compare, it is BioMax's professional opinion that interior particulate levels should continue to be minimized wherever possible. Therefore, additional (and ongoing) recommendations for optional particulate control measures have been provided at the end of this report for client consideration.

RECOMMENDATIONS

Based on the findings and conclusions presented in this report, BioMax believes that the current airborne microbial levels sampled and analyzed from within the identified 23rd Floor southern and eastern quadrant areas provides no significant evidence of elevated residual microbial contamination or airborne migration following the completion of prescribed microbial mitigative measures. Hence, based on our direct site observations, field measurements, and review of these findings at this time, BioMax believes that the previously affected areas may be considered acceptable for general reconstruction following prudent reconstruction practices with the implementation of the noted additional measures discussed below. Therefore, based on these findings, BioMax recommends the following post-mitigation measures and actions:

1. BioMax believes that current airborne microbial (mold) levels and mold types have been identified at levels which indicate that they reside within generally acceptable comparative ranges and parameters at present. Hence, BioMax recommends that no further airborne microbial sampling activities are warranted within the specific areas mitigated as part of the prescribed scope of work at this time. Certainly, due to the knowledge that microbial contamination, by nature, may change over time due to possible additional moisture intrusion, favorable growth conditions, and changing environments, these recommendations are subject to revision in the event that such conditions and/or environments arise.
2. During the performance of interior reconstruction activities, BioMax recommends that a qualified and experienced building inspector/contractor be utilized to verify the current functional integrity of all applicable plumbing, building flashing, sealing, and drainage systems in accordance with current building codes and construction practices. Any identified deficiencies should be appropriately documented, corrected, and then functionally verified (tested) prior to subsequent reconstruction and commercial use. Certainly, the establishment/installation of any additional engineering controls (as identified through additional professional engineering consultation) should also be performed and implemented in accordance with applicable standards, building codes, and ordinances, as necessary.
3. BioMax recommends that all reconstruction of interior structural building materials within these areas should only be undertaken utilizing high quality, visibly clean (hand selected) construction grade building materials obtained from reputable commercial sources and which are believed and visually free from elevated microbial contamination and/or elevated moisture content. Building materials, which are notably moist and/or visibly stained, should not be used during the reconstruction undertaken within the subject residence. BioMax recommends that all current plastic barriers (as established during this mitigation) should also remain during such reconstruction so as to minimize the potential transmission of associated construction dust and debris throughout the currently occupied areas of the subject structure.
4. As previously noted in is report, detectable levels of airborne particulates consisting of skin cell fragments and general debris particles were identified within the sampled interior areas surrounding the containment systems. Hence, and as an additional precautionary measure due to the presence of such materials, BioMax recommends that DGS considers the

performance of supplemental post reconstruction vacuuming and cleaning following the completion of interior renovation activities.

5. Following reconstruction, BioMax believes that any potential transmission and accumulation of the identified indoor airborne particulates may also be significantly reduced (if desired) on an immediate and ongoing basis through the use of routine HEPA filtered vacuuming and damp-wipe O&M cleaning methods employed by DGS maintenance personnel. BioMax's experience has indicated that these relatively simple and effective measures and methods have been shown to significantly reduce the accumulation of settled particulate debris on an immediate and ongoing basis if so desired.
6. Reasonable additional assessment and investigative measures may also be required upon the identification of new or previously undiscovered materials and/or information related to moisture/microbial impacts within the subject building structures, as necessary. Any occurrence and/or re-occurrence of moisture intrusion following routine O&M and/or general reconstruction within the subject building should also be reviewed and addressed through professional consultation, as necessary. BioMax is certainly prepared to provide such additional consultation pertaining to these and any follow-up investigative measures as necessary and upon request.

BioMax believes that the conclusions and recommendations outlined above are consistent with standard industry microbial mitigative practices and prudent industrial hygiene hazard control methods. Please do not hesitate to contact our offices directly at (510) 724-3100 if you have any additional questions, comments, or require further assistance regarding this matter.

Sincerely,



Michael A. Polkabila, CIH, REA
Vice President, Principal



LIMITATIONS

Please note that the professional opinions presented in this review are intended for the sole use of the California State Department of General Services (DGS) and their designated beneficiaries. No other party should rely on the information contained herein without the prior written consent of BioMax Environmental and DGS. The professional opinions provided herein are based on BioMax's review and understanding of current site information and observed site conditions present within the areas inspected at the time these services were performed. Professional recommendations provided as part of this limited scope of work are intended for client consideration only and are not intended as a professional or regulatory mandate. Implementation of any of the above measures or recommendations does not, in any way, warrant the day-to-day health and/or safety of building occupants, residents, site workers, nor regulatory or building code compliance status during normal and changing environmental conditions. As microbial contamination, by nature, may change over time due to additional moisture intrusion, favorable growth conditions, and changing environments, the findings of this report are subject to change in the event that such conditions and/or environments arise. Also, the professional opinions expressed here are subject to revision in the event that new or previously undiscovered information is obtained or uncovered.

The information contained in this and any other applicable communication is for consideration purposes only. It is not intended, nor should it be construed as providing legal advice or warranting any level of safety or regulatory compliance. The sole purpose of such information is to assist with the anticipation, identification, evaluation and control of elevated and/or unnecessary health of physical hazards. Any action taken based on this information, including but not limited to opinions, suggestions and recommendations, whether implied or expressed, is the sole responsibility of the individual taking the action. The management of acceptable health and safety is criteria dependent and situation specific in nature, therefore requiring extensive knowledge and prudent value assessments so as to be properly determined and maintained.

These services were performed by BioMax in accordance with generally accepted professional industrial hygiene principals, practices, and standards of care. Under the existing Industrial Hygiene Definition and Registration Act, all reports, opinions or official documents prepared by a Certified Industrial Hygienist (CIH) constitutes an expression of professional opinion regarding those facts or findings which are subject of a certification and does not constitute a warranty or guarantee, either expressed or implied.



EMLab P&K

Report for:

Mr. Michael Polkabila
Biomax Environmental
775 San Pablo Ave.
Pinole, CA 94564

Regarding: Project: 031008-01, 23 South; 450 N Street, Sacramento, CA, 23 South
EML ID: 398394

Approved by:



Lab Manager
Magzoub Ismail

Dates of Analysis:
Spore trap analysis: 03-13-2008

Project SOPs: Spore trap analysis (1100000)

This coversheet is included with your report in order to comply with AIHA and ISO accreditation requirements.

For clarity, we report the number of significant digits as calculated; but, due to the nature of this type of biological data, the number of significant digits that is used for interpretation should generally be one or two. All samples were received in acceptable condition unless noted in the Report Comments portion in the body of the report. Due to the nature of the analyses performed, field blank corrections of results is not a standard practice. The results relate only to the items tested.

EMLab P&K ("the Company") shall have no liability to the client or the client's customer with respect to decisions or recommendations made, actions taken or courses of conduct implemented by either the client or the client's customer as a result of or based upon the Test Results. In no event shall the Company be liable to the client with respect to the Test Results except for the Company's own willful misconduct or gross negligence nor shall the Company be liable for incidental or consequential damages or lost profits or revenues to the fullest extent such liability may be disclaimed by law, even if the Company has been advised of the possibility of such damages, lost profits or lost revenues. In no event shall the Company's liability with respect to the Test Results exceed the amount paid to the Company by the client therefor.

Document Number: 200091 - Revision Number: 5

Client: Biomax Environmental
 C/O: Mr. Michael Polkabila
 Re: 031008-01, 23 South; 450 N Street, Sacramento,
 CA, 23 South

Date of Sampling: 03-10-2008
 Date of Receipt: 03-12-2008
 Date of Report: 03-13-2008

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	13430672: Ambient front entry		13430716: 23rd floor balcony, West, amb		13430719: 12th floor ambient at intake area, SE		13430730: 23 SSW, rm 2311, outside cont	
Comments (see below)	None		None		None		None	
Lab ID-Version†:	1749978-1		1749979-1		1749980-1		1749981-1	
	raw ct.	spores/m3	raw ct.	spores/m3	raw ct.	spores/m3	raw ct.	spores/m3
Alternaria					2	27		
Arthrinium								
Ascospores*	5	227						
Aureobasidium								
Basidiospores*	17	627	6	320	4	213		
Bipolaris/Drechslera group								
Botrytis								
Chaetomium								
Cladosporium	7	373	4	213	22	1,170	2	107
Curvularia								
Epicoccum								
Fusarium								
Myrothecium								
Nigrospora								
Other colorless								
Penicillium/Aspergillus types†	16	853	19	1,010	27	1,440	9	480
Pithomyces								
Rusts*			1	13				
Smuts*, Periconia, Myxomycetes*								
Stachybotrys								
Stemphylium								
Torula								
Ulocladium								
Zygomycetes								
Background debris (1-4+)††	3+		3+		3+		3+	
Hyphal fragments/m3	< 13		< 13		80		< 13	
Pollen/m3	320		240		413		< 13	
Skin cells (1-4+)	None		None		None		None	
Sample volume (liters)	75		75		75		75	
TOTAL SPORE/m3		2,080		1,556		2,850		587

Comments:

* Most of these spore types are not seen with culturable methods (Andersen sampling), although some may appear as non-sporulating fungi. Most of the basidiospores are "mushroom" spores while the rusts and smuts are plant pathogens.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

†† Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.

The Limit of Detection is the product of a raw count of 1 and 100 divided by the percent read. The analytical sensitivity (counts/m3) is the product of the Limit of Detection and 1000 divided by the sample volume.

‡ A "Version" greater than 1 indicates amended data.

Client: Biomax Environmental
 C/O: Mr. Michael Polkabla
 Re: 031008-01, 23 South; 450 N Street, Sacramento,
 CA, 23 South

Date of Sampling: 03-10-2008
 Date of Receipt: 03-12-2008
 Date of Report: 03-13-2008

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	13430691: 23 SSW, in containment		13430698: 23 SSE, in containment		13430693: 23 S, office 2307		13430689: 23 S, equip corridor	
Comments (see below)	None		None		None		None	
Lab ID-Version‡:	1749982-1		1749983-1		1749984-1		1749985-1	
	raw ct.	spores/m3	raw ct.	spores/m3	raw ct.	spores/m3	raw ct.	spores/m3
Alternaria							1	13
Arthrinium								
Ascospores*								
Aureobasidium								
Basidiospores*							1	53
Bipolaris/Drechslera group								
Botrytis								
Chaetomium								
Cladosporium	4	213					1	53
Curvularia								
Epicoccum								
Fusarium								
Myrothecium								
Nigrospora								
Other colorless								
Penicillium/Aspergillus types†	10	533	2	107	3	160	9	480
Pithomyces								
Rusts*								
Smuts*, Periconia, Myxomycetes*								
Stachybotrys								
Stemphylium								
Torula								
Ulocladium								
Zygomycetes								
Background debris (1-4+)††	2+		3+		3+		3+	
Hyphal fragments/m3	< 13		< 13		< 13		< 13	
Pollen/m3	< 13		< 13		< 13		< 13	
Skin cells (1-4+)	< 1+		1+		1+		1+	
Sample volume (liters)	75		75		75		75	
TOTAL SPORE/m3		746		107		160		599

Comments:

* Most of these spore types are not seen with culturable methods (Andersen sampling), although some may appear as non-sporulating fungi. Most of the basidiospores are "mushroom" spores while the rusts and smuts are plant pathogens.
 † The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.
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 ‡ A "Version" greater than 1 indicates amended data.

Client: Biomax Environmental
 C/O: Mr. Michael Polkabila
 Re: 031008-01, 23 South; 450 N Street, Sacramento,
 CA, 23 South

Date of Sampling: 03-10-2008
 Date of Receipt: 03-12-2008
 Date of Report: 03-13-2008

MoldRANGE™: Extended Outdoor Comparison
Outdoor Location: 13430672, Ambient front entry

Fungi Identified	Outdoor data	Typical Outdoor Data by Date†				Typical Outdoor Data by Location‡			
		Month: March				State: CA			
	spores/m3	low	med	high	freq %	low	med	high	freq %
Generally able to grow indoors*									
Alternaria	-	7	27	210	43	7	27	230	60
Bipolaris/Drechslera group	-	7	13	120	12	7	13	120	14
Chaetomium	-	7	13	120	8	7	13	110	19
Cladosporium	373	27	320	4,300	91	53	640	6,500	98
Curvularia	-	7	13	210	7	7	13	210	7
Nigrospora	-	7	13	110	7	7	13	170	8
Penicillium/Aspergillus types	853	27	160	1,600	82	40	210	2,500	88
Stachybotrys	-	7	13	310	3	7	13	330	5
Torula	-	7	13	170	8	7	13	150	13
Seldom found growing indoors**									
Ascospores	227	13	130	2,000	74	13	110	1,800	73
Basidiospores	627	13	320	5,700	90	13	270	6,900	95
Rusts	-	7	13	320	17	7	13	270	29
Smuts, Periconia, Myxomycetes	-	7	27	310	54	8	40	470	71
TOTAL SPORES/M3	2,080								

† The Typical Outdoor Data by Date represents the typical outdoor spore levels across North America for the month indicated. The last column represents the frequency of occurrence. The low, medium, and high values represent the 2.5, 50, and 97.5 percentile values of the spore type when it is detected. For example, if the frequency of occurrence is 63% and the low value is 53, it would mean that the given spore type is detected 63% of the time and, when detected, 2.5% of the time it is present in levels above the detection limit and below 53 spores/m3. These values are updated periodically, and if enough data is not available to make a statistically meaningful assessment, it is indicated with a dash.

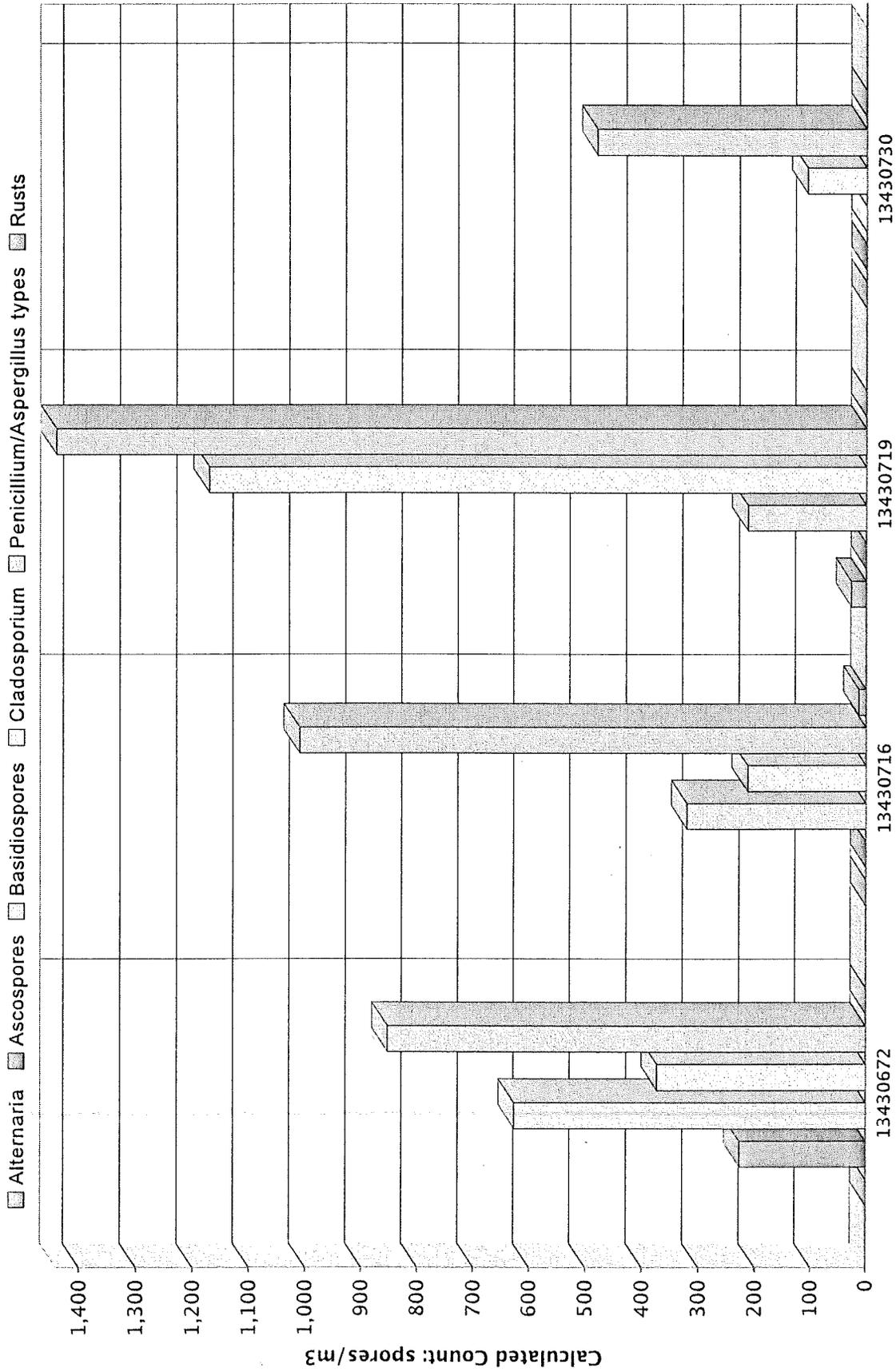
‡ The Typical Outdoor Data by Location represents the typical outdoor spore levels for the region indicated for the entire year. As with the Typical Outdoor Data by Date, the four columns represent the frequency of occurrence and the typical low, medium, and high concentration values for the spore type indicated. These values are updated periodically, and if enough data is not available to make a statistically meaningful assessment, it is indicated with a dash.

*The spores in this category are generally capable of growing on wet building materials in addition to growing outdoors. Building related growth is dependent upon the fungal type, moisture level, type of material, and other factors. *Cladosporium* is one of the predominant spore types worldwide and is frequently present in high numbers. *Penicillium/Aspergillus* species colonize both outdoor and indoor wet surfaces rapidly and are very easily dispersed. Other genera are usually present in lesser numbers.

**These fungi are generally not found growing on wet building materials. For example, the rusts and smuts are obligate plant pathogens. However, in each group there are notable exceptions. For example, agents of wood decay are members of the basidiomycetes and high counts of a single morphological type of basidiospore on an inside sample should be considered significant.

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SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

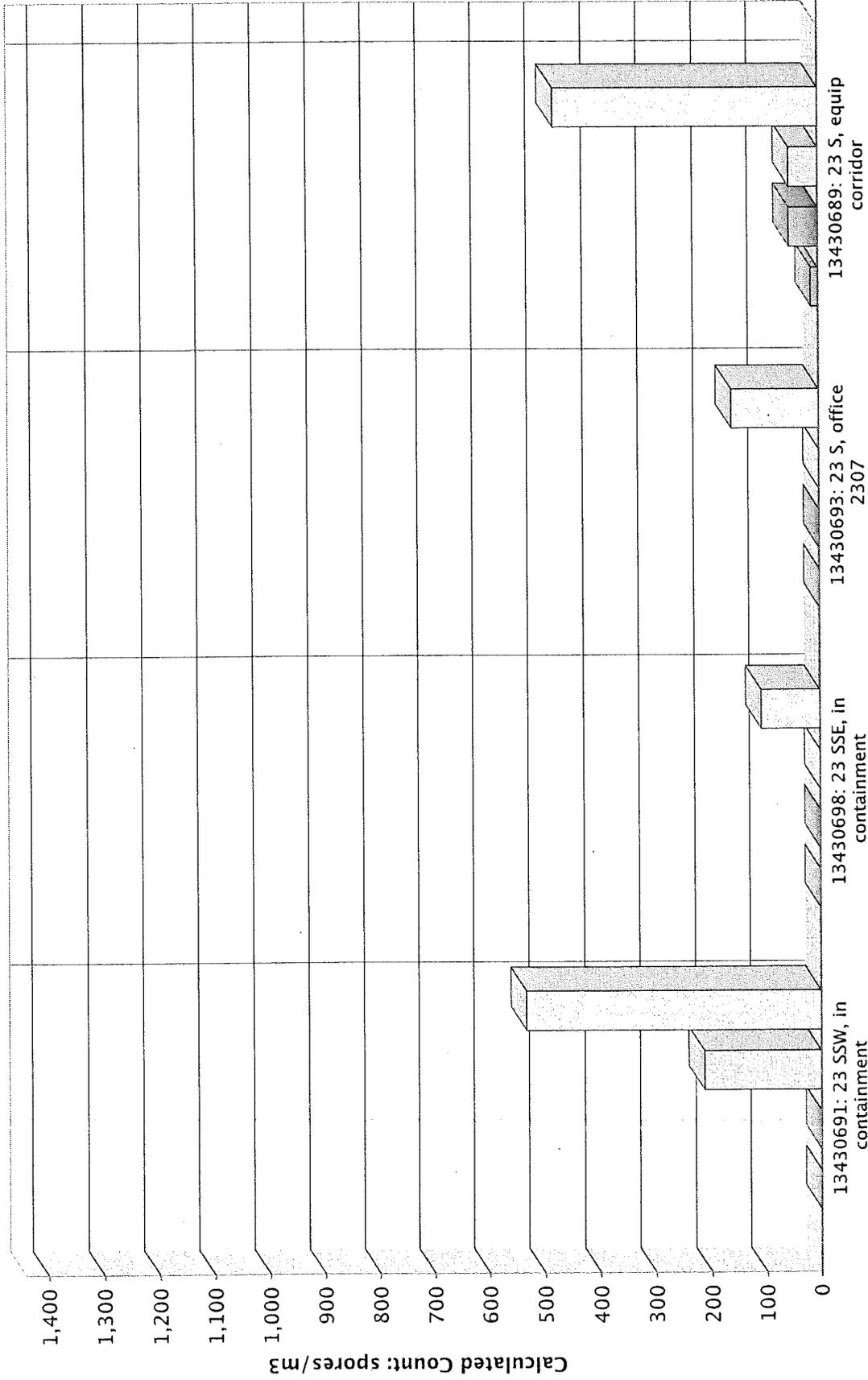


Comments:

Note: Graphical output may understate the importance of certain "marker" genera.

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

■ Alternaria ■ Basidiospores ■ Cladosporium ■ Penicillium/Aspergillus types



Comments:

Note: Graphical output may understate the importance of certain "marker" genera.

MICROBIAL SPORE TRAP AIR SAMPLING RECORD



000398394

Page 1 of 1

BioMax Environmental
775 San Pablo Ave.
Pinole, CA 94564

www.biomaxenvironmental.com

Phone: (510) 724-3100
Fax: (510) 724-3145
biomaxenv@aol.com

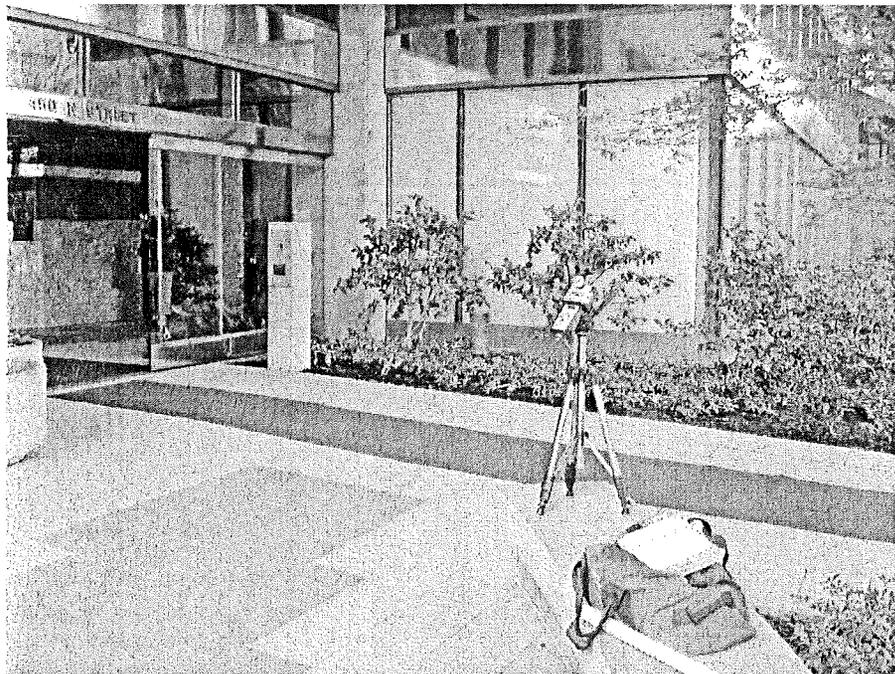
Location: 450 N Street Sacramento, CA 23 South	Client: DGS
Date: 3/10/08	Project #: 031008-01 23 South
Collected by: M. A. Polkable, CIH Signature: <i>Michael A Polkable</i>	Laboratory: EMLabs / PK Req. Turn Around: 24 HR Analysis: Fungal -Particulate-ID with Quantification.

Sample Number	Time	Location/Desc.	Temp / RH
13430672	1010	Ambient Front Entry	57.5° / 65%
13430716	1040	23rd Floor Balcony West (Amb)	60° / 52%
13430719	1100	12th Floor Ambient @ intake area SE	74° / 25%
13430730	1120	23SSW RM 2311 (outside cont.)	72° / 30%
13430691	1130	23SSW (in containment)	77 / 28%
13430698	1140	23SSE (in containment)	77 / 28%
13430693	1150	23 S office 2307	77 / 29%
13430689	1200	23S Equip corridor	75 / 30%
Total Sample Time (min): 5 min	Flow Rate (l/min): 15 l/min	Total Sample Volume (liters): 75 L	Ambient Conditions: Am - clear / mild Comments:

Please sign this form below acknowledging sample receipt and return executed form with laboratory reports. Fax, send, e-mail results to BioMax Environmental at (510) 724-3145 biomaxenv@aol.com
Other Instructions: _____

Relinquished by: <i>Michael Polkable</i>	Received By: <i>Ann Morrissey</i>
Method of Transportation: <i>Foot x Drop Box</i>	Time/Date Received: <i>3-12-08 9am</i>
Time/Date Sent: <i>3/10/08 @ 1800</i>	

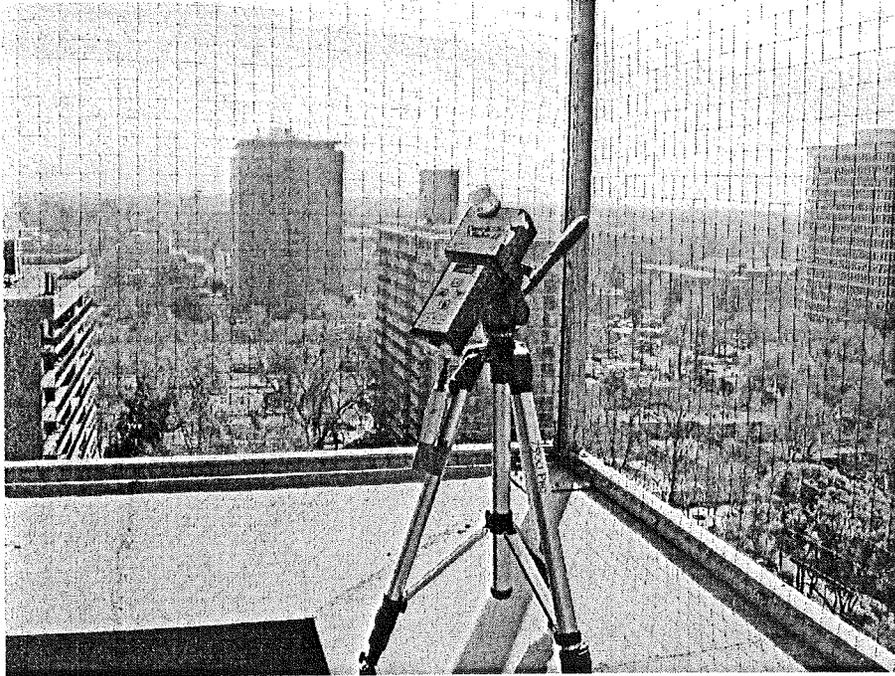
[Click here for color photos](#)



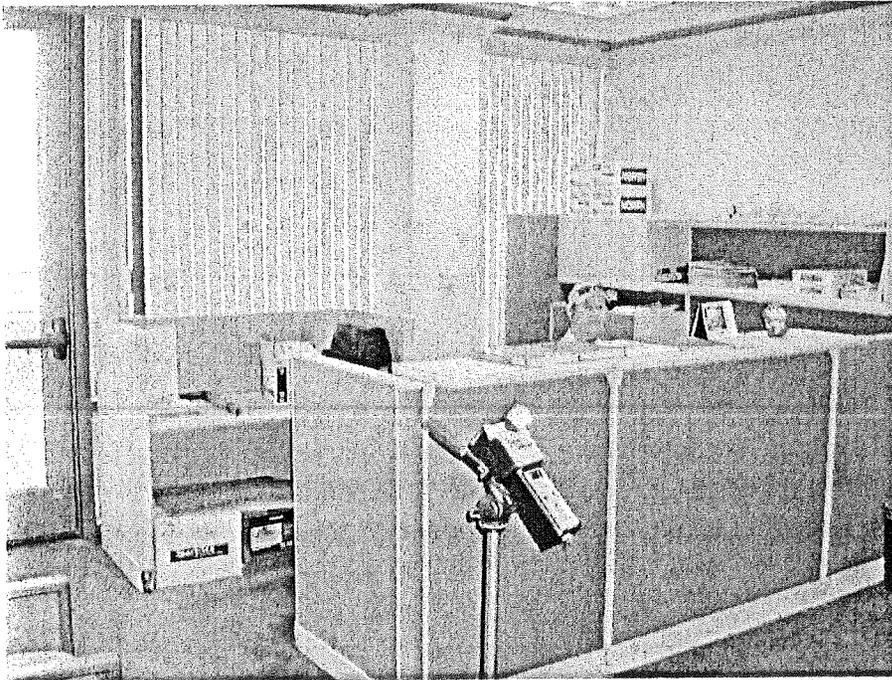
- 1) Image of ambient air sampling location at front entry of BOE Building (Subject Building) located at 450 N Street, Sacramento, California at time of assessment.



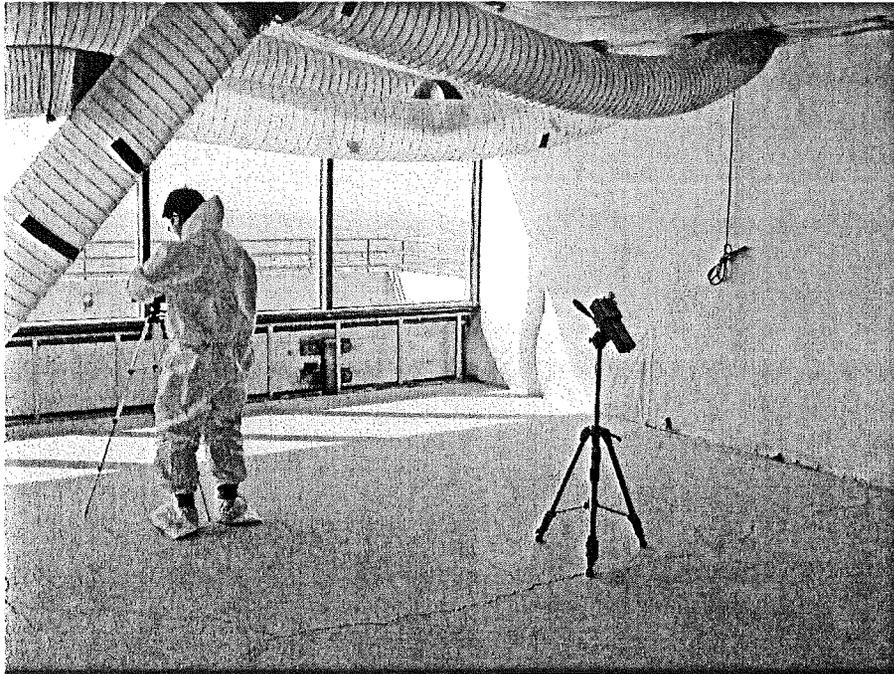
- 2) Image of Ambient air sampling location on balcony of 23 south with HTI technician collecting parallel samples.



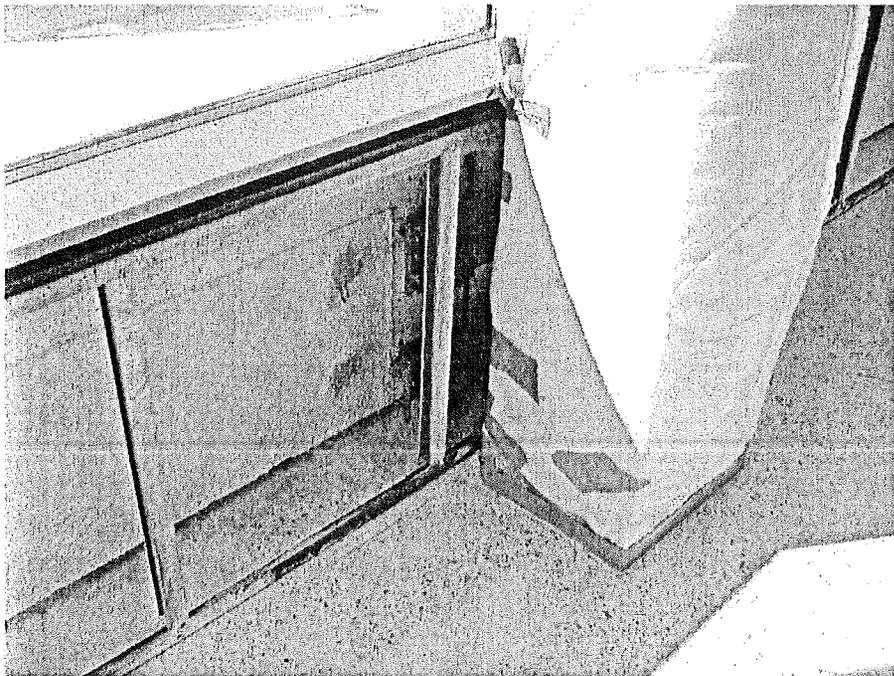
3) Image of 12 Floor southeastern balcony during ambient air collection procedures at time of assessment.



4) Image of hallway air sampling within construction offices location (outside containment) adjacent to 23 South containment area at time of assessment.



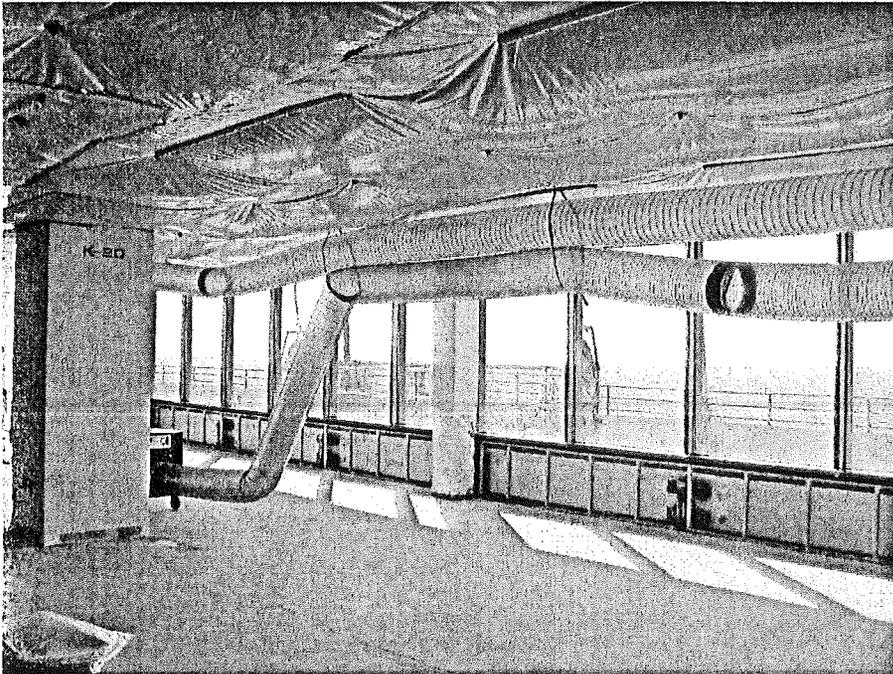
- 5) Image within 23rd Floor south containment area indicating location of air sampling equipment and “parallel” HTI technician activities.



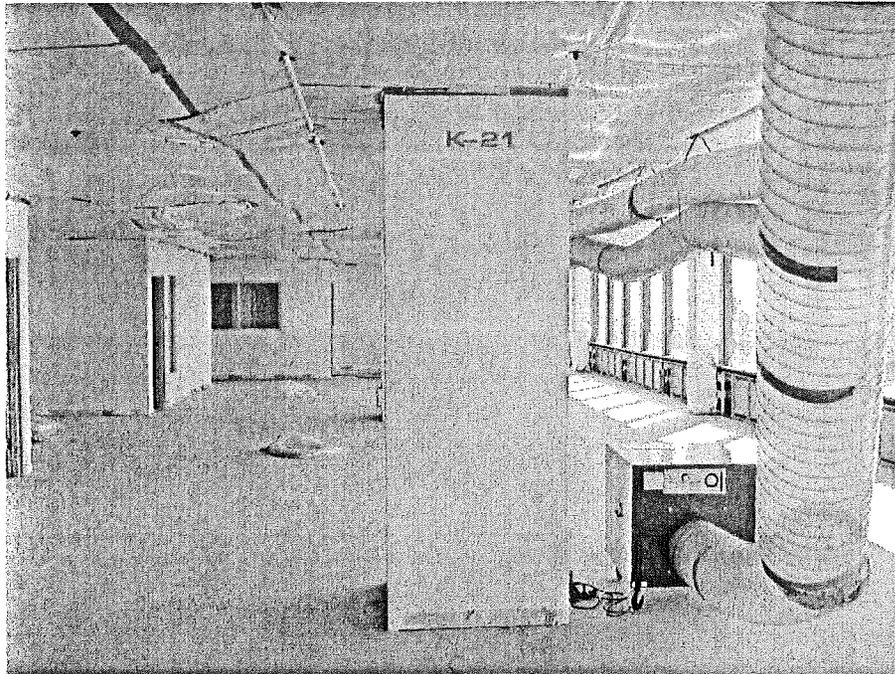
- 6) Image of encased column located at exterior perimeter wall within 23 South containment area following wallboard removal.



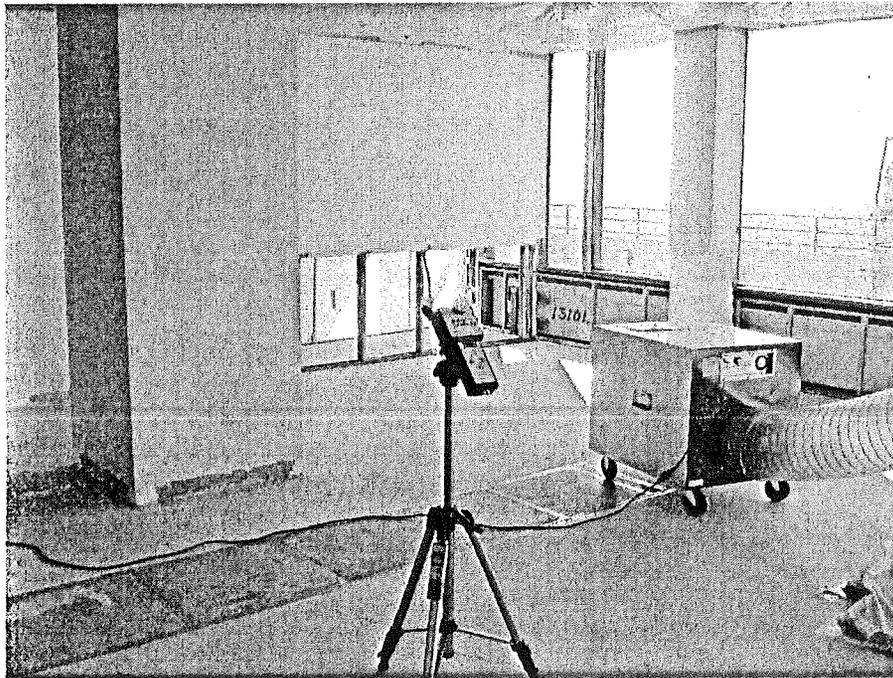
- 7) Close-up image of perimeter wall structures indicating evidence of prior water intrusion (corrosion on floor sill metal) following removal of damaged wallboard materials.



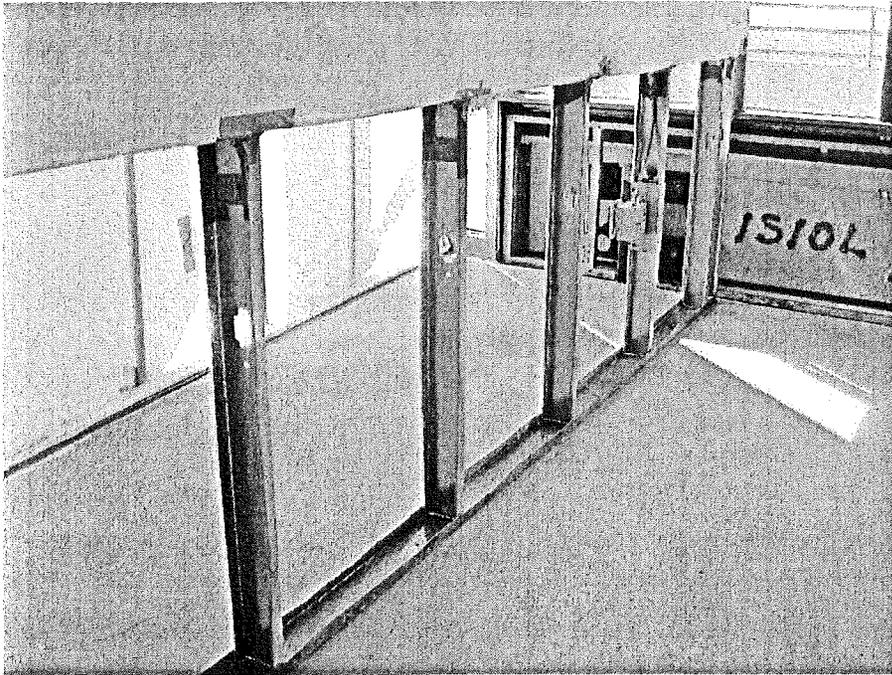
- 8) Wide angle image of 23South containment space indicating ventilation ducting and contained ceiling structures at time of assessment.



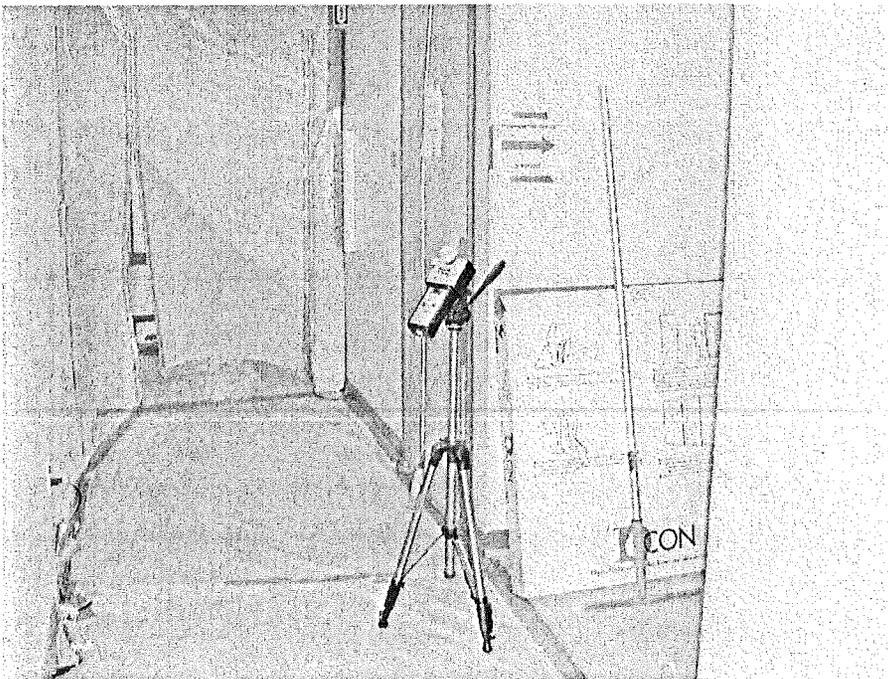
- 9) Additional image of air handling equipment location within 23 south containment area and exposed concrete subflooring present at time of assessment.



- 10) Image of air sampling equipment within eastern corner of 23 South containment area.



11) Additional image of exposed wall metal framing located at eastern corner of 23 South containment area.



12) Image of air sampling equipment located at exterior hallway of 23 South containment. Sample collected outside containment structures.



EMLab P&K

Report for:

Mr. Michael Polkabila
Biomax Environmental
775 San Pablo Ave.
Pinole, CA 94564

Regarding: Project: 031008.02, 23 East; 450 N Street, Sacramento, CA, 23 East
EML ID: 398389

Approved by:


Lab Manager
Magzoub Ismail

Dates of Analysis:
Spore trap analysis: 03-13-2008

Project SOPs: Spore trap analysis (I100000)

This coversheet is included with your report in order to comply with AIHA and ISO accreditation requirements.

For clarity, we report the number of significant digits as calculated; but, due to the nature of this type of biological data, the number of significant digits that is used for interpretation should generally be one or two. All samples were received in acceptable condition unless noted in the Report Comments portion in the body of the report. Due to the nature of the analyses performed, field blank corrections of results is not a standard practice. The results relate only to the items tested.

EMLab P&K ("the Company") shall have no liability to the client or the client's customer with respect to decisions or recommendations made, actions taken or courses of conduct implemented by either the client or the client's customer as a result of or based upon the Test Results. In no event shall the Company be liable to the client with respect to the Test Results except for the Company's own willful misconduct or gross negligence nor shall the Company be liable for incidental or consequential damages or lost profits or revenues to the fullest extent such liability may be disclaimed by law, even if the Company has been advised of the possibility of such damages, lost profits or lost revenues. In no event shall the Company's liability with respect to the Test Results exceed the amount paid to the Company by the client therefor.

Document Number: 200091 - Revision Number: 5

Client: Biomax Environmental
 C/O: Mr. Michael Polkabila
 Re: 031008.02, 23 East; 450 N Street, Sacramento,
 CA, 23 East

Date of Sampling: 03-10-2008
 Date of Receipt: 03-12-2008
 Date of Report: 03-13-2008

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	13430731: Rm 2339 in containment		13430665: Rm 2338 in containment		13430726: Rm 2337 in containment		13430703: Rm 2336 in cont	
Comments (see below)	None		None		None		None	
Lab ID-Version‡:	1750101-1		1750102-1		1750103-1		1750104-1	
	raw ct.	spores/m3	raw ct.	spores/m3	raw ct.	spores/m3	raw ct.	spores/m3
Alternaria								
Arthrinium								
Ascospores*								
Aureobasidium								
Basidiospores*	2	107	2	67	1	53		
Bipolaris/Drechslera group								
Botrytis								
Chaetomium								
Cladosporium							1	53
Curvularia								
Epicoccum								
Fusarium								
Myrothecium								
Nigrospora								
Oidium								
Other brown								
Other colorless								
Penicillium/Aspergillus types†	2	107						
Pithomyces								
Rusts*								
Smuts*, Periconia, Myxomycetes*								
Stachybotrys								
Stemphylium								
Torula								
Ulocladium								
Background debris (1-4+)††	2+		2+		2+		2+	
Hyphal fragments/m3	< 13		< 13		< 13		< 13	
Pollen/m3	13		< 13		< 13		< 13	
Skin cells (1-4+)	< 1+		< 1+		< 1+		< 1+	
Sample volume (liters)	75		75		75		75	
TOTAL SPORE/m3		214		67		53		53

Comments:

* Most of these spore types are not seen with culturable methods (Andersen sampling), although some may appear as non-sporulating fungi. Most of the basidiospores are "mushroom" spores while the rusts and smuts are plant pathogens.
 † The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.
 †† Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.
 The Limit of Detection is the product of a raw count of 1 and 100 divided by the percent read. The analytical sensitivity (counts/m3) is the product of the Limit of Detection and 1000 divided by the sample volume.
 ‡ A "Version" greater than 1 indicates amended data.

Client: Biomax Environmental
 C/O: Mr. Michael Polkabra
 Re: 031008.02, 23 East; 450 N Street, Sacramento,
 CA, 23 East

Date of Sampling: 03-10-2008
 Date of Receipt: 03-12-2008
 Date of Report: 03-13-2008

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	13430700: Hallway outside containment		13430734: Rm 2335 in containment		13430684: Rm 2334 in containment		13430674: Rm 2332 in containment	
Comments (see below)	None		None		None		None	
Lab ID-Version‡:	1750105-1		1750106-1		1750107-1		1750108-1	
	raw ct.	spores/m3	raw ct.	spores/m3	raw ct.	spores/m3	raw ct.	spores/m3
Alternaria								
Arthrinium								
Ascospores*								
Aureobasidium								
Basidiospores*					1	53		
Bipolaris/Drechslera group								
Botrytis								
Chaetomium								
Cladosporium	1	53	1	53				
Curvularia								
Epicoccum								
Fusarium								
Myrothecium								
Nigrospora								
Oidium								
Other brown	1	13			1	13		
Other colorless								
Penicillium/Aspergillus types†							1	53
Pithomyces								
Rusts*								
Smuts*, Periconia, Myxomycetes*								
Stachybotrys					1	13		
Stemphylium								
Torula								
Ulocladium								
Background debris (1-4+)††	2+		2+		2+		2+	
Hyphal fragments/m3	< 13		< 13		< 13		< 13	
Pollen/m3	< 13		< 13		< 13		< 13	
Skin cells (1-4+)	< 1+		< 1+		< 1+		< 1+	
Sample volume (liters)	75		75		75		75	
TOTAL SPORE/m3		66		53		79		53

Comments:

* Most of these spore types are not seen with culturable methods (Andersen sampling), although some may appear as non-sporulating fungi. Most of the basidiospores are "mushroom" spores while the rusts and smuts are plant pathogens.
 † The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.
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Client: Biomax Environmental
 C/O: Mr. Michael Polkabra
 Re: 031008.02, 23 East; 450 N Street, Sacramento,
 CA, 23 East

Date of Sampling: 03-10-2008
 Date of Receipt: 03-12-2008
 Date of Report: 03-13-2008

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	13430676: Amb West balcony 23		13430688: Ambient front entry area	
Comments (see below)	None		None	
Lab ID-Version‡:	1750109-1		1750110-1	
	raw ct.	spores/m3	raw ct.	spores/m3
Alternaria	2	27	2	27
Arthrinium				
Ascospores*	3	120	2	107
Aureobasidium				
Basidiospores*	6	200	6	240
Bipolaris/Drechslera group				
Botrytis				
Chaetomium	4	53	1	13
Cladosporium	13	693	10	533
Curvularia				
Epicoccum				
Fusarium				
Myrothecium				
Nigrospora				
Oidium			1	13
Other brown	3	40	2	27
Other colorless	1	13		
Penicillium/Aspergillus types†			4	213
Pithomyces				
Rusts*				
Smuts*, Periconia, Myxomycetes*	3	40	1	13
Stachybotrys	1	13	3	40
Stemphylium				
Torula				
Ulocladium				
Zygomycetes				
Background debris (1-4+)††	2+		3+	
Hyphal fragments/m3	13		< 13	
Pollen/m3	253		320	
Skin cells (1-4+)	< 1+		< 1+	
Sample volume (liters)	75		75	
TOTAL SPORE/m3		1,199		1,226

Comments:

* Most of these spore types are not seen with culturable methods (Andersen sampling), although some may appear as non-sporulating fungi. Most of the basidiospores are "mushroom" spores while the rusts and smuts are plant pathogens.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

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Client: Biomax Environmental
 C/O: Mr. Michael Polkabila
 Re: 031008.02, 23 East; 450 N Street, Sacramento,
 CA, 23 East

Date of Sampling: 03-10-2008
 Date of Receipt: 03-12-2008
 Date of Report: 03-13-2008

MoldRANGE™: Extended Outdoor Comparison

Outdoor Location: 13430676, Amb West balcony 23

Fungi Identified	Outdoor data	Typical Outdoor Data by Date†				Typical Outdoor Data by Location‡			
		Month: March				State: CA			
	spores/m3	low	med	high	freq %	low	med	high	freq %
Generally able to grow indoors*									
Alternaria	27	7	27	210	43	7	27	230	60
Bipolaris/Drechslera group	-	7	13	120	12	7	13	120	14
Chaetomium	53	7	13	120	8	7	13	110	19
Cladosporium	693	27	320	4,300	91	53	640	6,500	98
Curvularia	-	7	13	210	7	7	13	210	7
Nigrospora	-	7	13	110	7	7	13	170	8
Other brown	40	7	13	80	34	7	13	80	37
Other colorless	13	7	13	120	9	7	13	93	7
Penicillium/Aspergillus types	-	27	160	1,600	82	40	210	2,500	88
Stachybotrys	13	7	13	310	3	7	13	330	5
Torula	-	7	13	170	8	7	13	150	13
Seldom found growing indoors**									
Ascospores	120	13	130	2,000	74	13	110	1,800	73
Basidiospores	200	13	320	5,700	90	13	270	6,900	95
Oidium	-	7	13	330	14	7	13	200	20
Rusts	-	7	13	320	17	7	13	270	29
Smuts, Periconia, Myxomycetes	40	7	27	310	54	8	40	470	71
TOTAL SPORES/M3	1,199								

† The Typical Outdoor Data by Date represents the typical outdoor spore levels across North America for the month indicated. The last column represents the frequency of occurrence. The low, medium, and high values represent the 2.5, 50, and 97.5 percentile values of the spore type when it is detected. For example, if the frequency of occurrence is 63% and the low value is 53, it would mean that the given spore type is detected 63% of the time and, when detected, 2.5% of the time it is present in levels above the detection limit and below 53 spores/m3. These values are updated periodically, and if enough data is not available to make a statistically meaningful assessment, it is indicated with a dash.

‡ The Typical Outdoor Data by Location represents the typical outdoor spore levels for the region indicated for the entire year. As with the Typical Outdoor Data by Date, the four columns represent the frequency of occurrence and the typical low, medium, and high concentration values for the spore type indicated. These values are updated periodically, and if enough data is not available to make a statistically meaningful assessment, it is indicated with a dash.

*The spores in this category are generally capable of growing on wet building materials in addition to growing outdoors. Building related growth is dependent upon the fungal type, moisture level, type of material, and other factors. *Cladosporium* is one of the predominant spore types worldwide and is frequently present in high numbers. *Penicillium/Aspergillus* species colonize both outdoor and indoor wet surfaces rapidly and are very easily dispersed. Other genera are usually present in lesser numbers.

**These fungi are generally not found growing on wet building materials. For example, the rusts and smuts are obligate plant pathogens. However, in each group there are notable exceptions. For example, agents of wood decay are members of the basidiomycetes and high counts of a single morphological type of basidiospore on an inside sample should be considered significant.

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Client: Biomax Environmental
 C/O: Mr. Michael Polkabila
 Re: 031008.02, 23 East; 450 N Street, Sacramento,
 CA, 23 East

Date of Sampling: 03-10-2008

Date of Receipt: 03-12-2008

Date of Report: 03-13-2008

MoldRANGE™: Extended Outdoor Comparison**Outdoor Location: 13430688, Ambient front entry area**

Fungi Identified	Outdoor data	Typical Outdoor Data by Date†				Typical Outdoor Data by Location‡			
		Month: March				State: CA			
	spores/m3	low	med	high	freq %	low	med	high	freq %
Generally able to grow indoors*									
Alternaria	27	7	27	210	43	7	27	230	60
Bipolaris/Drechslera group	-	7	13	120	12	7	13	120	14
Chaetomium	13	7	13	120	8	7	13	110	19
Cladosporium	533	27	320	4,300	91	53	640	6,500	98
Curvularia	-	7	13	210	7	7	13	210	7
Nigrospora	-	7	13	110	7	7	13	170	8
Other brown	27	7	13	80	34	7	13	80	37
Other colorless	-	7	13	120	9	7	13	93	7
Penicillium/Aspergillus types	213	27	160	1,600	82	40	210	2,500	88
Stachybotrys	40	7	13	310	3	7	13	330	5
Torula	-	7	13	170	8	7	13	150	13
Seldom found growing indoors**									
Ascospores	107	13	130	2,000	74	13	110	1,800	73
Basidiospores	240	13	320	5,700	90	13	270	6,900	95
Oidium	13	7	13	330	14	7	13	200	20
Rusts	-	7	13	320	17	7	13	270	29
Smuts, Periconia, Myxomycetes	13	7	27	310	54	8	40	470	71
TOTAL SPORES/M3	1,226								

† The Typical Outdoor Data by Date represents the typical outdoor spore levels across North America for the month indicated. The last column represents the frequency of occurrence. The low, medium, and high values represent the 2.5, 50, and 97.5 percentile values of the spore type when it is detected. For example, if the frequency of occurrence is 63% and the low value is 53, it would mean that the given spore type is detected 63% of the time and, when detected, 2.5% of the time it is present in levels above the detection limit and below 53 spores/m3. These values are updated periodically, and if enough data is not available to make a statistically meaningful assessment, it is indicated with a dash.

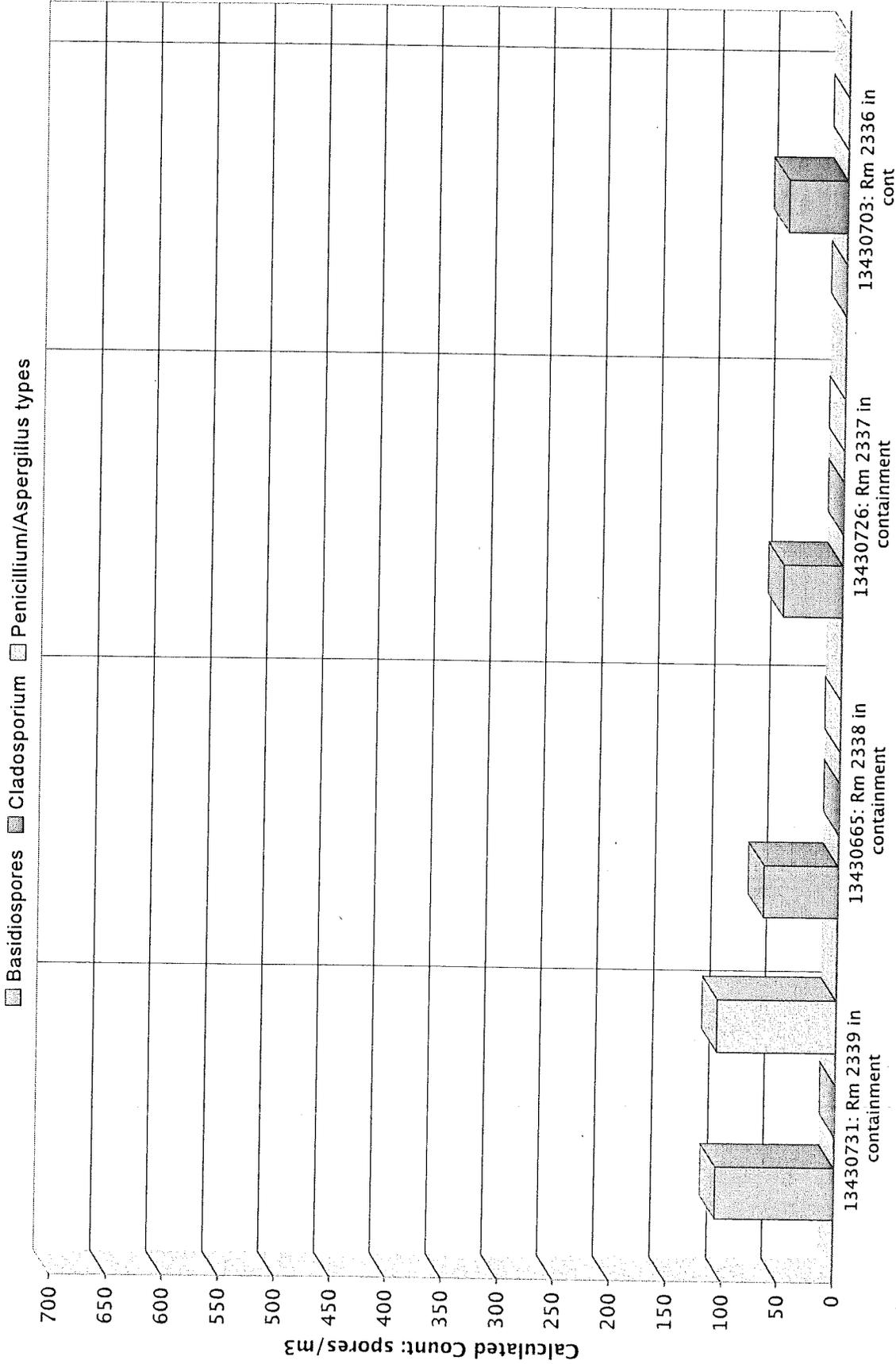
‡ The Typical Outdoor Data by Location represents the typical outdoor spore levels for the region indicated for the entire year. As with the Typical Outdoor Data by Date, the four columns represent the frequency of occurrence and the typical low, medium, and high concentration values for the spore type indicated. These values are updated periodically, and if enough data is not available to make a statistically meaningful assessment, it is indicated with a dash.

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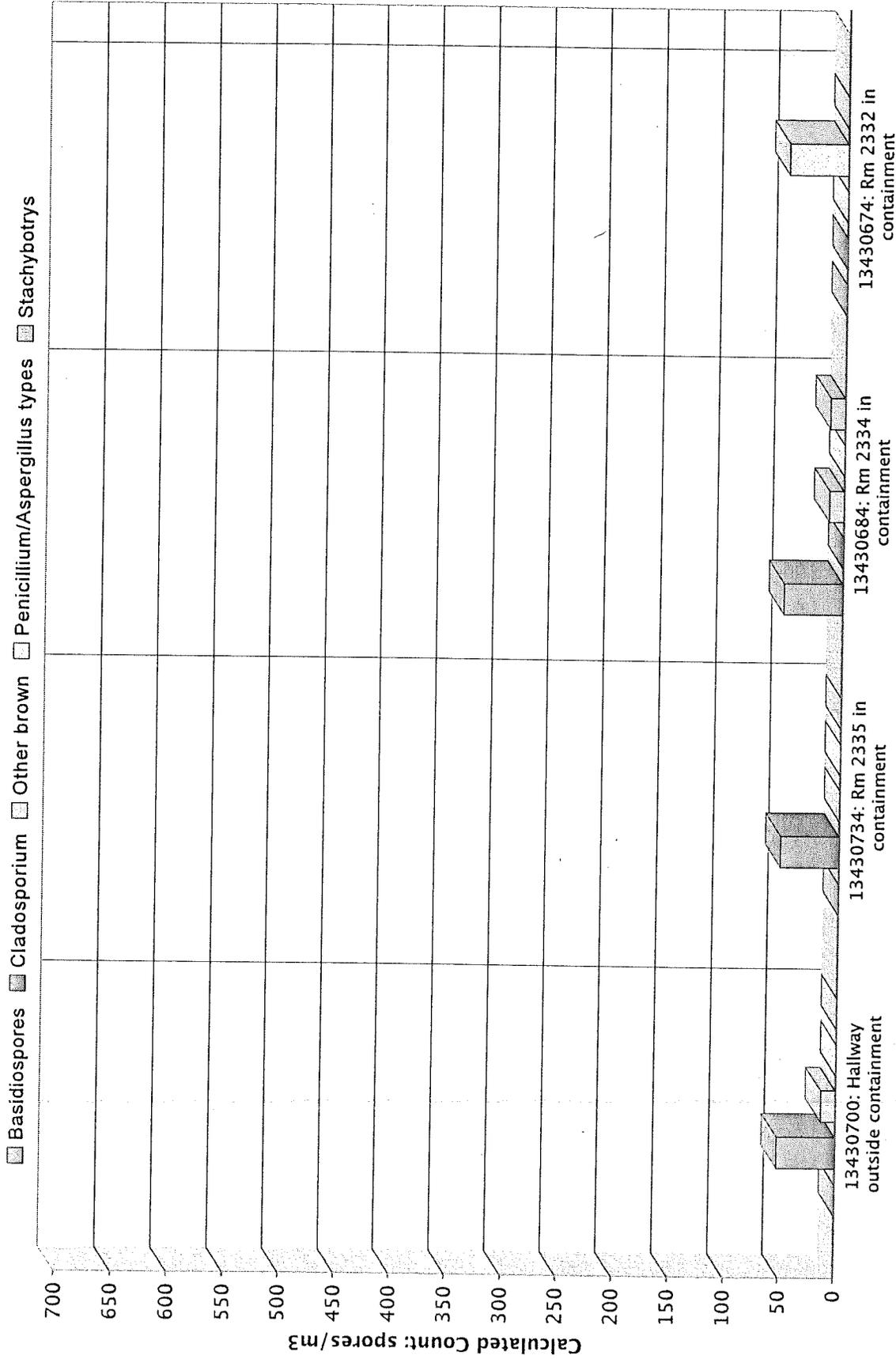
SPORE TRAP REPORT: NON-VIABLE METHODOLOGY



Comments:

Note: Graphical output may understate the importance of certain "marker" genera.

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

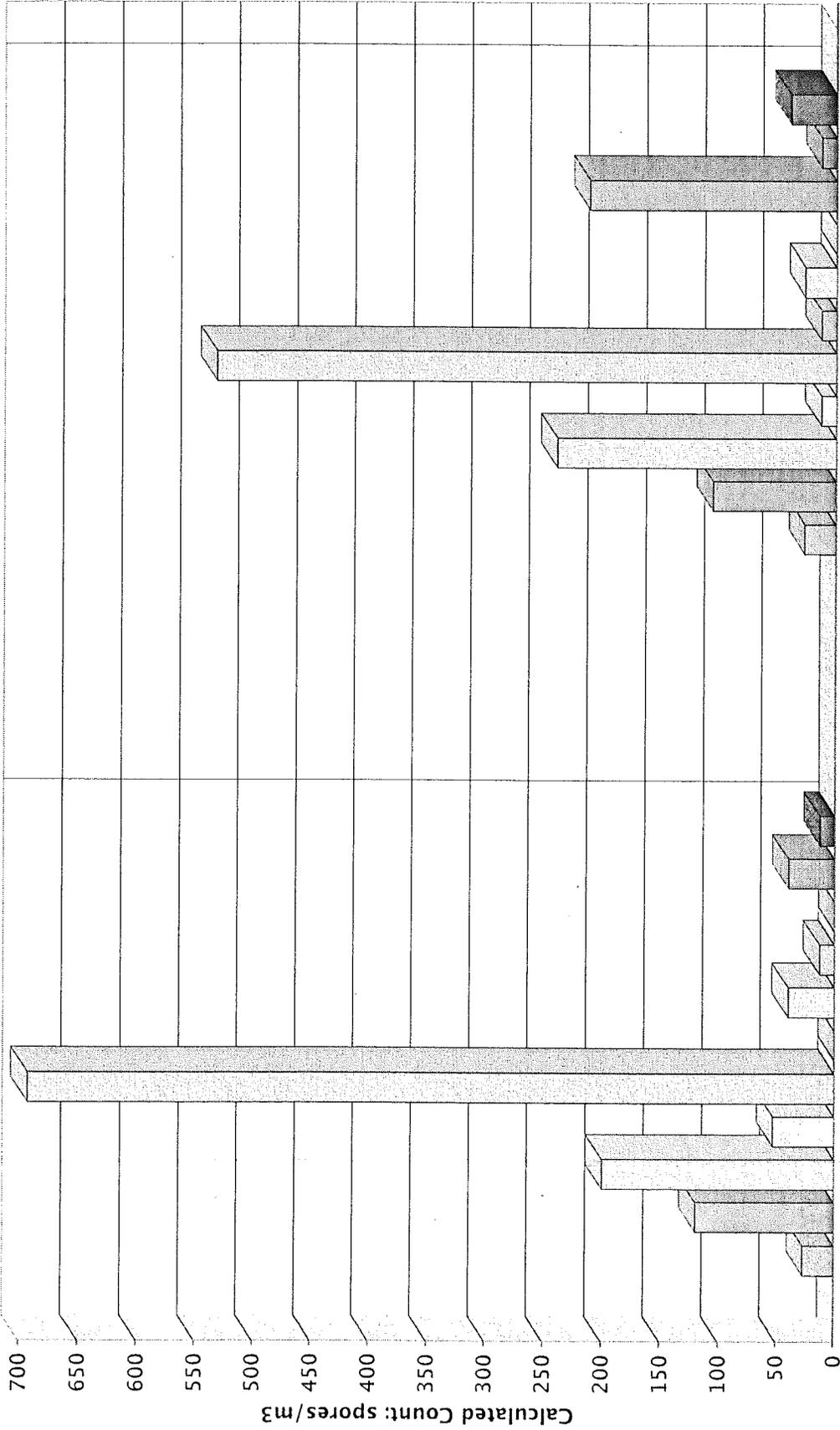


Comments:

Note: Graphical output may understate the importance of certain "marker" genera.

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

- Alternaria Ascospores Basidiospores Chaetomium Cladosporium Oidium Other brown Other colorless
- Penicillium/Aspergillus types Smuts, Periconia, Myxomycetes Stachybotrys



13430676: Amb West balcony 23

13430688: Ambient front entry area

Comments:

Note: Graphical output may understate the importance of certain "marker" genera.

MICROBIAL SPORE TRAP AIR SAMPLING RECORD



000398389

Page 1 of 1

BioMax Environmental
775 San Pablo Ave.
Pinole, CA 94564

www.biomaxenvironmental.com

Phone: (510) 724-3100
Fax: (510) 724-3145
biomaxenv@aol.com

Location: 450 N. Street Sacramento, CA 23 East	Client: DGS
Date: 3/10/08	Project #: 031008-02 23 East
Collected by: M. A. Palkobk	Laboratory: EML Labs
Signature: <i>[Signature]</i>	Req. Turn Around: 24 h.
	Analysis (circle): Fungal / Particulate ID / Quantification.

	Sample Number	Time	Location/Desc.	Temp / RH
I.	13430731	1320	Rm 2339 in Containment.	86° / 26%
II.	13430665	1330	Rm 2338 in Containment.	86° / 25%
III.	13430726	1335	Rm 2337 in Containment.	85° / 25%
IV.	13430703	1345	Rm 2336 in Cont.	85° / 27%
V.	13430700	1350	Hallway outside Containment.	76° / 43%
VI.	13430734	1515	Rm 2335 in Containment	80° / 28%
VII.	13430684	1525	Rm 2334 in Containment	80° / 28%
VIII.	13430674	1530	Rm 2332 in Containment	79° / 28%
IX.	13430676	1545	Dark West Balcony 23	77° / 30%
X.	13430688	1555	Ambient Front Entry Area	72° / 36%
	Total Sample Time (min): 5 min	Flow Rate (l/min): 15 l/min	Total Sample Volume (liters): 75 L	Ambient Conditions: clear / mild
	Comments:			

Please sign this form below acknowledging sample receipt and return executed form with laboratory reports. Fax, send, e-mail results to BioMax Environmental at (510) 724-3145 biomaxenv@aol.com
Other Instructions: _____

Relinquished by: <i>[Signature]</i>	Received By: Ann Morrissey
Method of Transportation: FedEx Drop Box	Time/Date Received: 3-12-08 <i>ann</i>
Time/Date Sent: 3/10/08 1800	

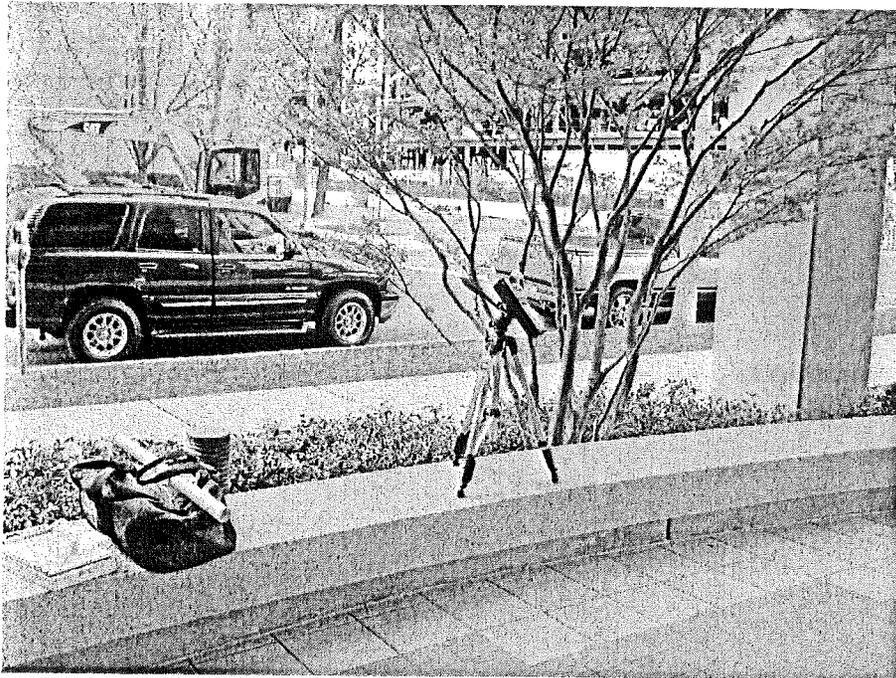
Received by: *[Signature]*
3/13/08 10:00 AM

Attachment A: Digital Images

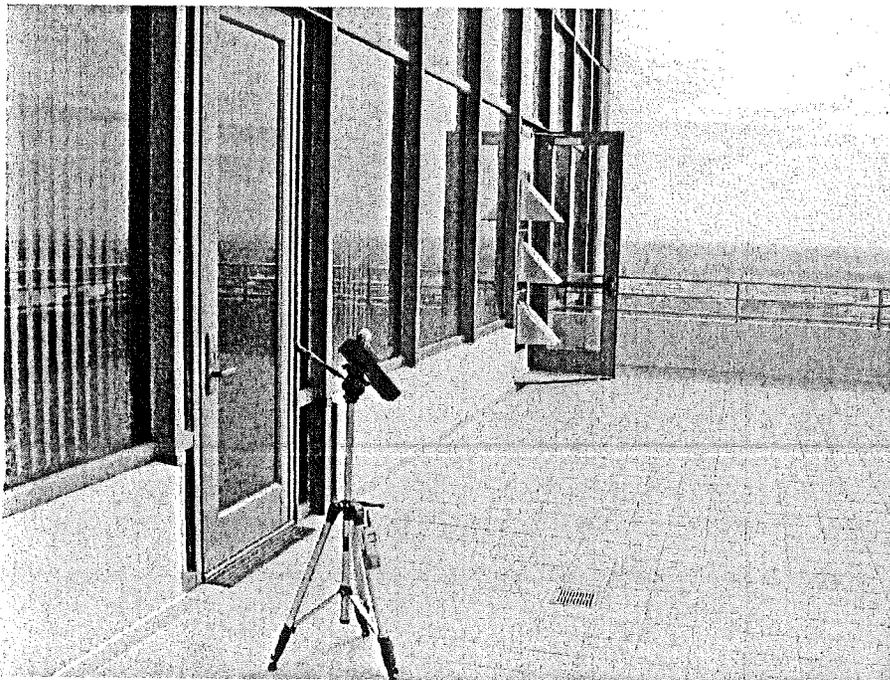
March 10th, 2008

BOE Building 23rd Floor East
Sacramento, CA

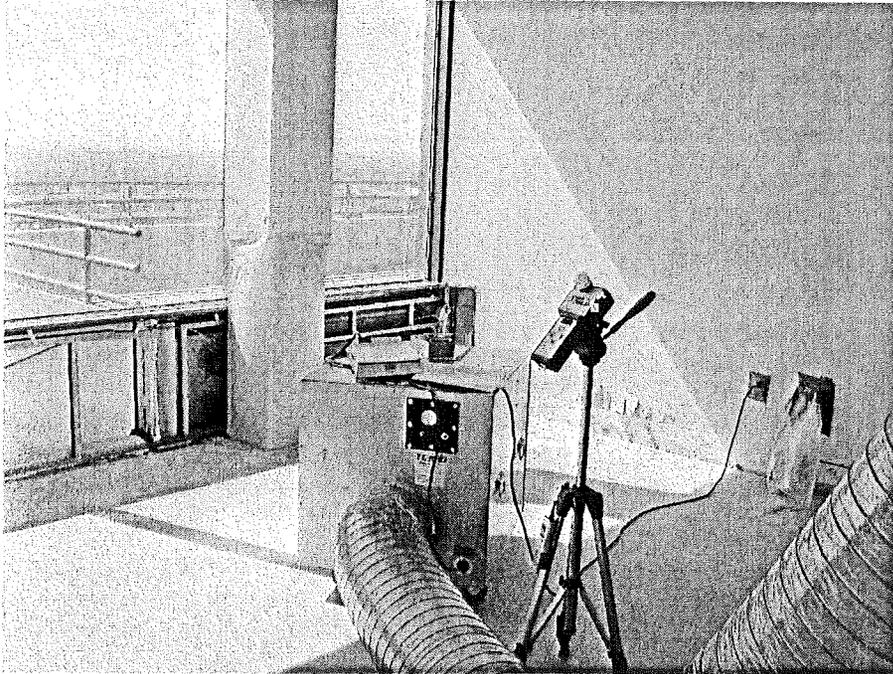
[Click here for color photos](#)



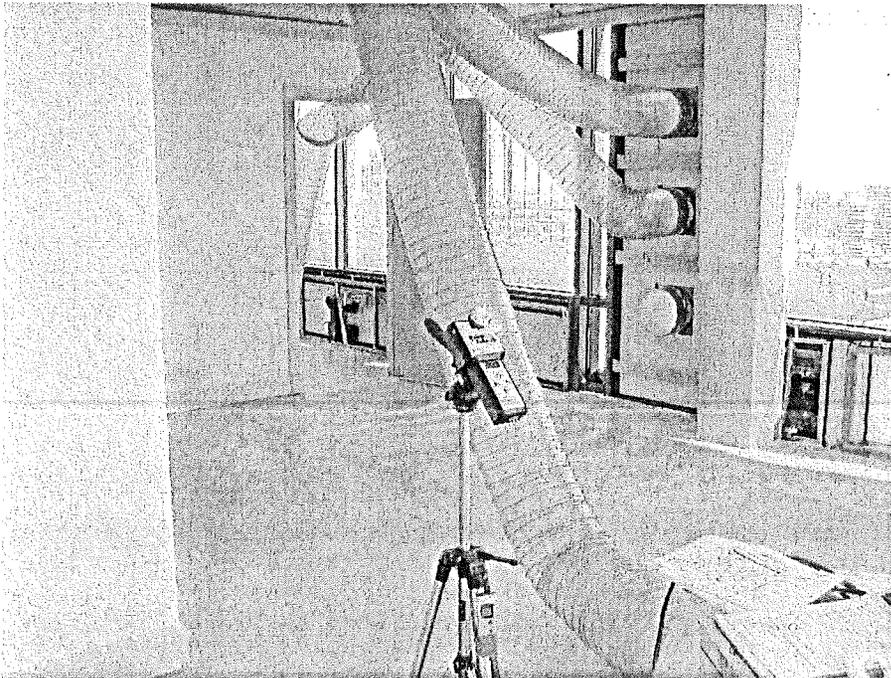
- 1) Image of ambient air sampling location at front entry of BOE Building (Subject Building) located at 450 N Street, Sacramento, California at time of assessment.



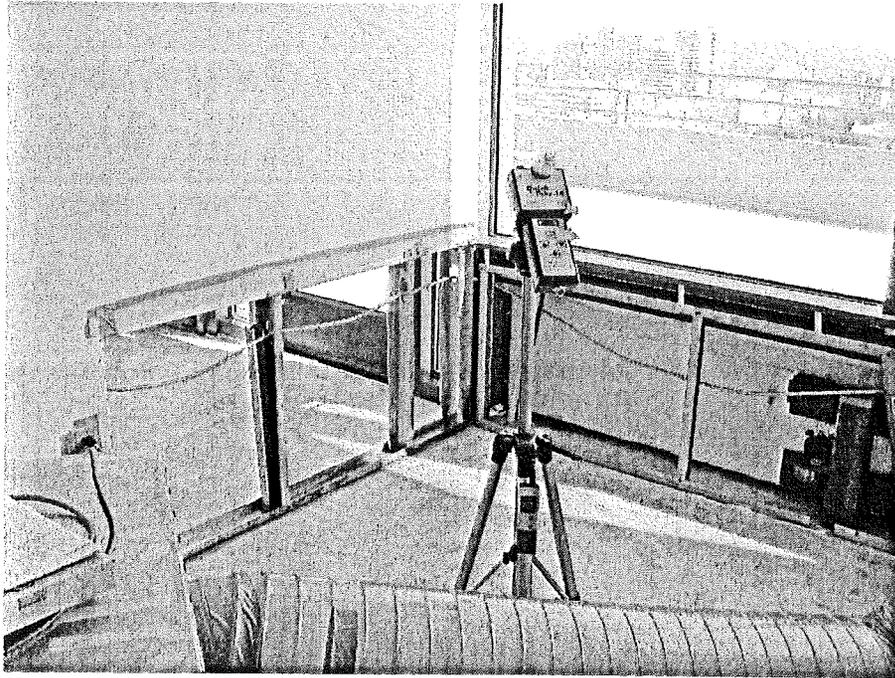
- 2) Image of Ambient air sampling location on balcony of 23 South at time of assessment.



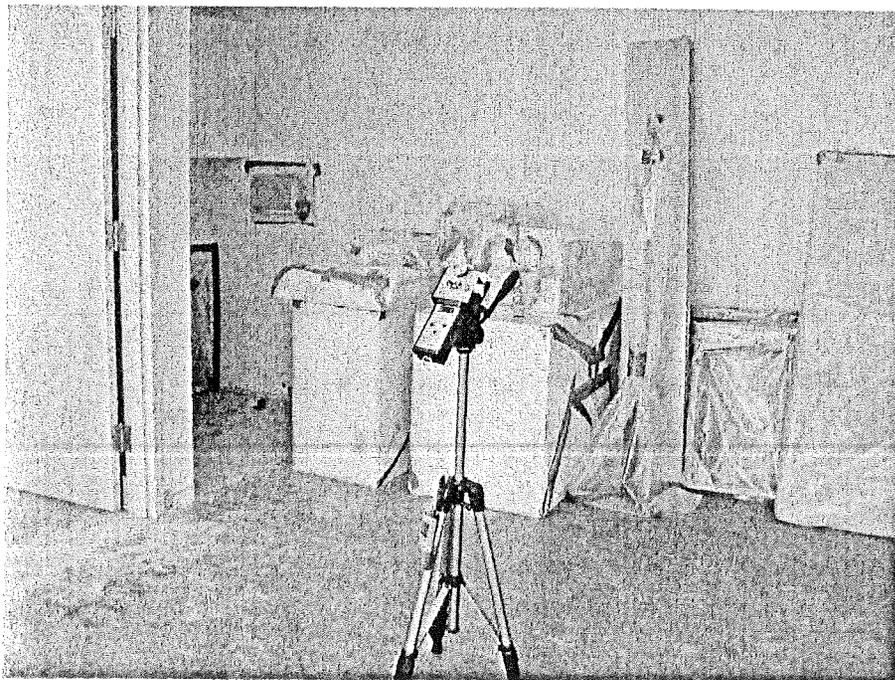
- 3) Image within 23 Eastern containment system indicating location of air sampling devices and air handling equipment at time of assessment.



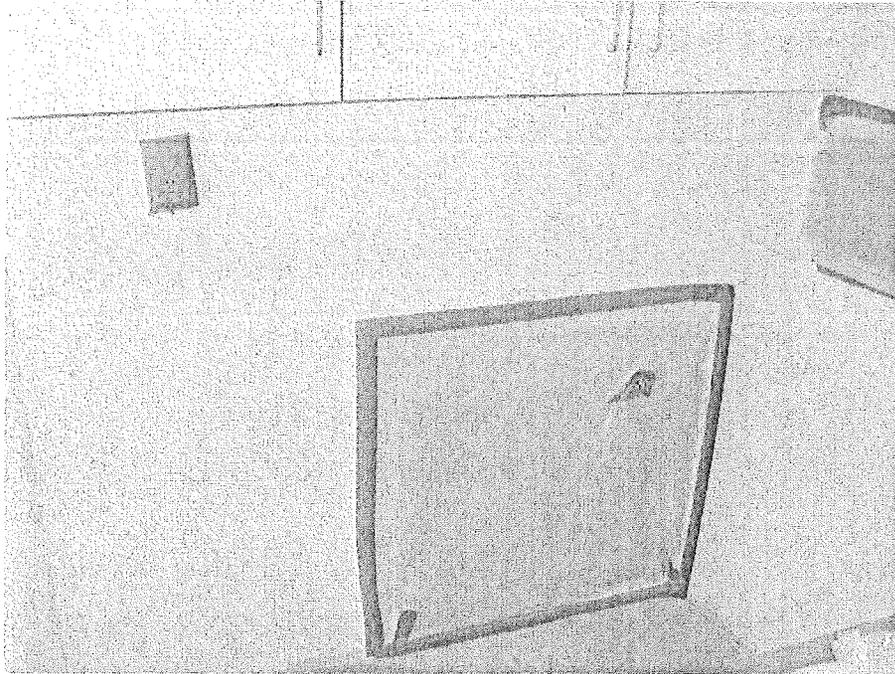
- 4) Additional image of air sampling and air handling equipment within 23 East containment area at time of assessment.



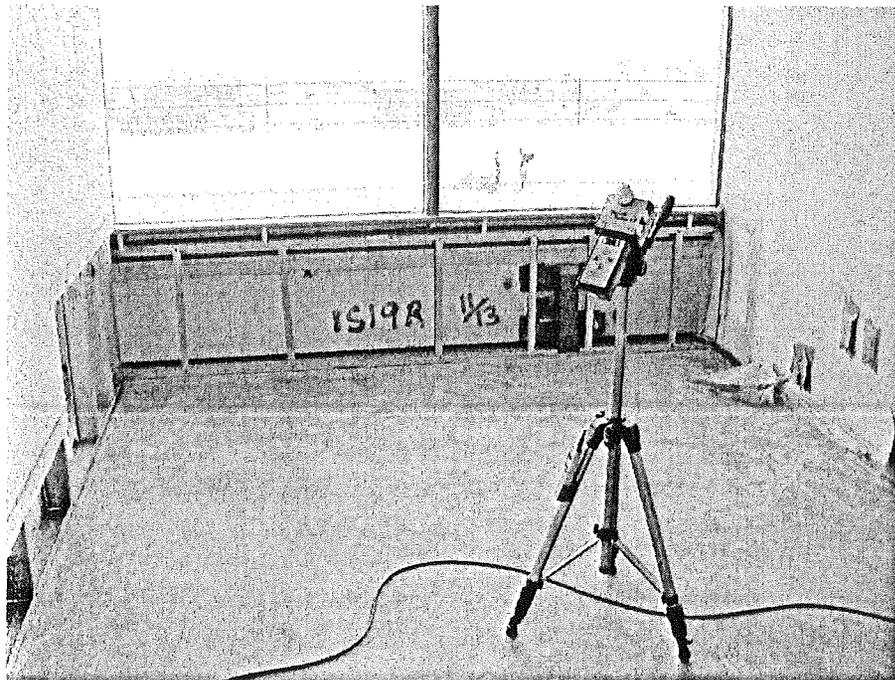
- 5) Image within 23rd Floor east containment area indicating location of air sampling equipment and extent of perimeter and interior wall removal.



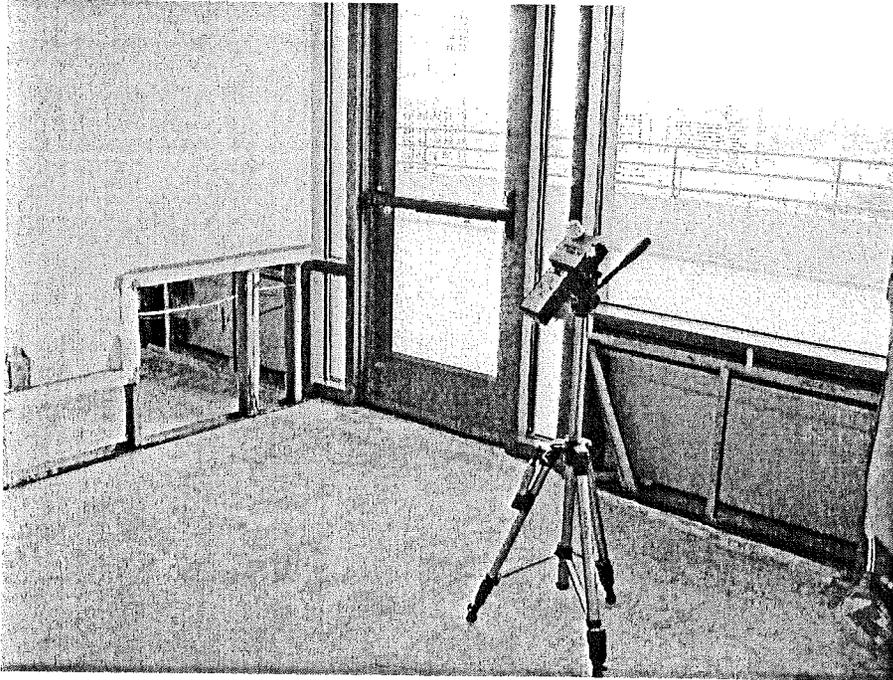
- 6) Image of protected office furnishings located within interior office where air sampling was performed as part of this assessment.



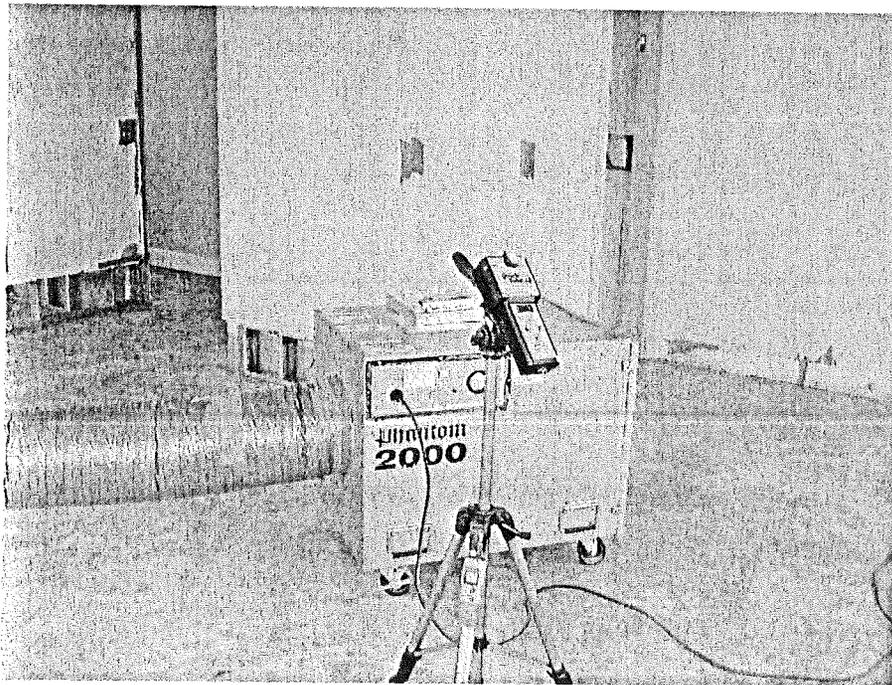
7) Close-up image of mini coffee bar cabinet location indicating wallboard and cabinet removal prior to clearance assessment within 23 East containment area.



8) Air sampling within 23 East containment perimeter office space indicating extent of exterior and interior wallboard removal.



9) Additional image of air handling equipment location within 23 East perimeter office at doorway location.



10) Image of air sampling equipment within 23 East containment area.

EMLab P&K

4401 Atlantic Avenue, Suite 200, Long Beach, CA 90807
 (858) 569-5800 Fax (858) 569-5806 www.emlab.com

Client: Biomax Environmental
 C/O: Mr. Michael Polkabla
 Re: 031008.02, 23 East; 450 N Street, Sacramento,
 CA, 23 East

Date of Sampling: 03-10-2008
 Date of Receipt: 03-12-2008
 Date of Report: 03-13-2008

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	13430731: Rm 2339 in containment		13430665: Rm 2338 in containment		13430726: Rm 2337 in containment		13430703: Rm 2336 in cont	
Comments (see below)	None		None		None		None	
Lab ID-Version‡:	1750101-1		1750102-1		1750103-1		1750104-1	
	raw ct.	spores/m3	raw ct.	spores/m3	raw ct.	spores/m3	raw ct.	spores/m3
Alternaria								
Arthrimum								
Ascospores*								
Aureobasidium								
Basidiospores*	2	107	2	67	1	53		
Bipolaris/Drechslera group								
Botrytis								
Chaetomium								
Cladosporium							1	53
Curvularia								
Epicoccum								
Fusarium								
Myrothecium								
Nigrospora								
Oidium								
Other brown								
Other colorless								
Penicillium/Aspergillus types†	2	107						
Pithomyces								
Rusts*								
Smuts*, Periconia, Myxomycetes*								
Stachybotrys								
Stemphylium								
Torula								
Ulocladium								
Background debris (1-4+)††	2+		2+		2+		2+	
Hyphal fragments/m3	< 13		< 13		< 13		< 13	
Pollen/m3	13		< 13		< 13		< 13	
Skin cells (1-4+)	< 1+		< 1+		< 1+		< 1+	
Sample volume (liters)	75		75		75		75	
TOTAL SPORE/m3		214		67		53		53

Comments:

* Most of these spore types are not seen with culturable methods (Andersen sampling), although some may appear as non-sporulating fungi. Most of the basidiospores are "mushroom" spores while the rusts and smuts are plant pathogens.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

†† Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.

The Limit of Detection is the product of a raw count of 1 and 100 divided by the percent read. The analytical sensitivity (counts/m3) is the product of the Limit of Detection and 1000 divided by the sample volume.

‡ A "Version" greater than 1 indicates amended data.

Client: Biomax Environmental
 C/O: Mr. Michael Polkabila
 Re: 031008.02, 23 East; 450 N Street, Sacramento,
 CA, 23 East

Date of Sampling: 03-10-2008
 Date of Receipt: 03-12-2008
 Date of Report: 03-13-2008

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	13430700: Hallway outside containment		13430734: Rm 2335 in containment		13430684: Rm 2334 in containment		13430674: Rm 2332 in containment	
Comments (see below)	None		None		None		None	
Lab ID-Version‡:	1750105-1		1750106-1		1750107-1		1750108-1	
	raw ct.	spores/m3	raw ct.	spores/m3	raw ct.	spores/m3	raw ct.	spores/m3
Alternaria								
Arthrinium								
Ascospores*								
Aureobasidium								
Basidiospores*					1	53		
Bipolaris/Drechslera group								
Botrytis								
Chaetomium								
Cladosporium	1	53	1	53				
Curvularia								
Epicoccum								
Fusarium								
Myrothecium								
Nigrospora								
Oidium								
Other brown	1	13			1	13		
Other colorless								
Penicillium/Aspergillus types†							1	53
Pithomyces								
Rusts*								
Smuts*, Periconia, Myxomycetes*								
Stachybotrys					1	13		
Stemphylium								
Torula								
Ulocladium								
Background debris (1-4+)††	2+		2+		2+		2+	
Hyphal fragments/m3	< 13		< 13		< 13		< 13	
Pollen/m3	< 13		< 13		< 13		< 13	
Skin cells (1-4+)	< 1+		< 1+		< 1+		< 1+	
Sample volume (liters)	75		75		75		75	
TOTAL SPORE/m3		66		53		79		53

Comments:

* Most of these spore types are not seen with culturable methods (Andersen sampling), although some may appear as non-sporulating fungi. Most of the basidiospores are "mushroom" spores while the rusts and smuts are plant pathogens.
 † The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.
 †† Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.
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 ‡ A "Version" greater than 1 indicates amended data.

Client: Biomax Environmental
C/O: Mr. Michael Polkabila
Re: 031008.02, 23 East; 450 N Street, Sacramento,
CA, 23 East

Date of Sampling: 03-10-2008
Date of Receipt: 03-12-2008
Date of Report: 03-13-2008

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	13430676: Amb West balcony 23		13430688: Ambient front entry area	
Comments (see below)	None		None	
Lab ID-Version‡:	1750109-1		1750110-1	
	raw ct.	spores/m3	raw ct.	spores/m3
Alternaria	2	27	2	27
Arthrinium				
Ascospores*	3	120	2	107
Aureobasidium				
Basidiospores*	6	200	6	240
Bipolaris/Drechslera group				
Botrytis				
Chaetomium	4	53	1	13
Cladosporium	13	693	10	533
Curvularia				
Epicoccum				
Fusarium				
Myrothecium				
Nigrospora				
Oidium			1	13
Other brown	3	40	2	27
Other colorless	1	13		
Penicillium/Aspergillus types†			4	213
Pithomyces				
Rusts*				
Smuts*, Periconia, Myxomycetes*	3	40	1	13
Stachybotrys	1	13	3	40
Stemphylium				
Torula				
Ulocladium				
Zygomycetes				
Background debris (1-4+)††	2+		3+	
Hyphal fragments/m3	13		< 13	
Pollen/m3	253		320	
Skin cells (1-4+)	< 1+		< 1+	
Sample volume (liters)	75		75	
TOTAL SPORE/m3		1,199		1,226

Comments:

* Most of these spore types are not seen with culturable methods (Andersen sampling), although some may appear as non-sporulating fungi. Most of the basidiospores are "mushroom" spores while the rusts and smuts are plant pathogens.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

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Client: Biomax Environmental
 C/O: Mr. Michael Polkabla
 Re: 031008.02, 23 East; 450 N Street, Sacramento,
 CA, 23 East

Date of Sampling: 03-10-2008
 Date of Receipt: 03-12-2008
 Date of Report: 03-13-2008

MoldRANGE™: Extended Outdoor Comparison**Outdoor Location: 13430676, Amb West balcony 23**

Fungi Identified	Outdoor data	Typical Outdoor Data by Date†				Typical Outdoor Data by Location‡			
		Month: March				State: CA			
	spores/m3	low	med	high	freq %	low	med	high	freq %
Generally able to grow indoors*									
Alternaria	27	7	27	210	43	7	27	230	60
Bipolaris/Drechslera group	-	7	13	120	12	7	13	120	14
Chaetomium	53	7	13	120	8	7	13	110	19
Cladosporium	693	27	320	4,300	91	53	640	6,500	98
Curvularia	-	7	13	210	7	7	13	210	7
Nigrospora	-	7	13	110	7	7	13	170	8
Other brown	40	7	13	80	34	7	13	80	37
Other colorless	13	7	13	120	9	7	13	93	7
Penicillium/Aspergillus types	-	27	160	1,600	82	40	210	2,500	88
Stachybotrys	13	7	13	310	3	7	13	330	5
Torula	-	7	13	170	8	7	13	150	13
Seldom found growing indoors**									
Ascospores	120	13	130	2,000	74	13	110	1,800	73
Basidiospores	200	13	320	5,700	90	13	270	6,900	95
Oidium	-	7	13	330	14	7	13	200	20
Rusts	-	7	13	320	17	7	13	270	29
Smuts, Periconia, Myxomycetes	40	7	27	310	54	8	40	470	71
TOTAL SPORES/M3	1,199								

† The Typical Outdoor Data by Date represents the typical outdoor spore levels across North America for the month indicated. The last column represents the frequency of occurrence. The low, medium, and high values represent the 2.5, 50, and 97.5 percentile values of the spore type when it is detected. For example, if the frequency of occurrence is 63% and the low value is 53, it would mean that the given spore type is detected 63% of the time and, when detected, 2.5% of the time it is present in levels above the detection limit and below 53 spores/m³. These values are updated periodically, and if enough data is not available to make a statistically meaningful assessment, it is indicated with a dash.

‡ The Typical Outdoor Data by Location represents the typical outdoor spore levels for the region indicated for the entire year. As with the Typical Outdoor Data by Date, the four columns represent the frequency of occurrence and the typical low, medium, and high concentration values for the spore type indicated. These values are updated periodically, and if enough data is not available to make a statistically meaningful assessment, it is indicated with a dash.

*The spores in this category are generally capable of growing on wet building materials in addition to growing outdoors. Building related growth is dependent upon the fungal type, moisture level, type of material, and other factors. *Cladosporium* is one of the predominant spore types worldwide and is frequently present in high numbers. *Penicillium/Aspergillus* species colonize both outdoor and indoor wet surfaces rapidly and are very easily dispersed. Other genera are usually present in lesser numbers.

**These fungi are generally not found growing on wet building materials. For example, the rusts and smuts are obligate plant pathogens. However, in each group there are notable exceptions. For example, agents of wood decay are members of the basidiomycetes and high counts of a single morphological type of basidiospore on an inside sample should be considered significant.

Interpretation of the data contained in this report is left to the client or the persons who conducted the field work. This report is provided for informational and comparative purposes only and should not be relied upon for any other purpose. "Typical outdoor data" are based on the results of the analysis of samples delivered to and analyzed by EMLab P&K and assumptions regarding the origins of those samples. Sampling techniques, contaminants infecting samples, unrepresentative samples and other similar or dissimilar factors may affect these results. In addition, EMLab P&K may not have received and tested a representative number of samples for every region or time period. EMLab P&K hereby disclaims any liability for any and all direct, indirect, punitive, incidental, special or consequential damages arising out of the use or interpretation of the data contained in, or any actions taken or omitted in reliance upon, this report.

EMLab P&K

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Client: Biomax Environmental
 C/O: Mr. Michael Polkabla
 Re: 031008.02, 23 East; 450 N Street, Sacramento,
 CA, 23 East

Date of Sampling: 03-10-2008

Date of Receipt: 03-12-2008

Date of Report: 03-13-2008

MoldRANGE™: Extended Outdoor Comparison**Outdoor Location: 13430688, Ambient front entry area**

Fungi Identified	Outdoor data	Typical Outdoor Data by Date†				Typical Outdoor Data by Location‡			
		Month: March				State: CA			
	spores/m3	low	med	high	freq %	low	med	high	freq %
Generally able to grow indoors*									
Alternaria	27	7	27	210	43	7	27	230	60
Bipolaris/Drechslera group	-	7	13	120	12	7	13	120	14
Chaetomium	13	7	13	120	8	7	13	110	19
Cladosporium	533	27	320	4,300	91	53	640	6,500	98
Curvularia	-	7	13	210	7	7	13	210	7
Nigrospora	-	7	13	110	7	7	13	170	8
Other brown	27	7	13	80	34	7	13	80	37
Other colorless	-	7	13	120	9	7	13	93	7
Penicillium/Aspergillus types	213	27	160	1,600	82	40	210	2,500	88
Stachybotrys	40	7	13	310	3	7	13	330	5
Torula	-	7	13	170	8	7	13	150	13
Seldom found growing indoors**									
Ascospores	107	13	130	2,000	74	13	110	1,800	73
Basidiospores	240	13	320	5,700	90	13	270	6,900	95
Oidium	13	7	13	330	14	7	13	200	20
Rusts	-	7	13	320	17	7	13	270	29
Smuts, Periconia, Myxomycetes	13	7	27	310	54	8	40	470	71
TOTAL SPORES/M3	1,226								

† The Typical Outdoor Data by Date represents the typical outdoor spore levels across North America for the month indicated. The last column represents the frequency of occurrence. The low, medium, and high values represent the 2.5, 50, and 97.5 percentile values of the spore type when it is detected. For example, if the frequency of occurrence is 63% and the low value is 53, it would mean that the given spore type is detected 63% of the time and, when detected, 2.5% of the time it is present in levels above the detection limit and below 53 spores/m³. These values are updated periodically, and if enough data is not available to make a statistically meaningful assessment, it is indicated with a dash.

‡ The Typical Outdoor Data by Location represents the typical outdoor spore levels for the region indicated for the entire year. As with the Typical Outdoor Data by Date, the four columns represent the frequency of occurrence and the typical low, medium, and high concentration values for the spore type indicated. These values are updated periodically, and if enough data is not available to make a statistically meaningful assessment, it is indicated with a dash.

*The spores in this category are generally capable of growing on wet building materials in addition to growing outdoors. Building related growth is dependent upon the fungal type, moisture level, type of material, and other factors. *Cladosporium* is one of the predominant spore types worldwide and is frequently present in high numbers. *Penicillium/Aspergillus* species colonize both outdoor and indoor wet surfaces rapidly and are very easily dispersed. Other genera are usually present in lesser numbers.

**These fungi are generally not found growing on wet building materials. For example, the rusts and smuts are obligate plant pathogens. However, in each group there are notable exceptions. For example, agents of wood decay are members of the basidiomycetes and high counts of a single morphological type of basidiospore on an inside sample should be considered significant.

Interpretation of the data contained in this report is left to the client or the persons who conducted the field work. This report is provided for informational and comparative purposes only and should not be relied upon for any other purpose. "Typical outdoor data" are based on the results of the analysis of samples delivered to and analyzed by EMLab P&K and assumptions regarding the origins of those samples. Sampling techniques, contaminants infecting samples, unrepresentative samples and other similar or dissimilar factors may affect these results. In addition, EMLab P&K may not have received and tested a representative number of samples for every region or time period. EMLab P&K hereby disclaims any liability for any and all direct, indirect, punitive, incidental, special or consequential damages arising out of the use or interpretation of the data contained in, or any actions taken or omitted in reliance upon, this report.

Client: Biomax Environmental
C/O: Mr. Michael Polkabila
Re: 031008-01, 23 South; 450 N Street, Sacramento,
CA, 23 South

Date of Sampling: 03-10-2008
Date of Receipt: 03-12-2008
Date of Report: 03-13-2008

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	13430672: Ambient front entry		13430716: 23rd floor balcony, West, amb		13430719: 12th floor ambient at intake area, SE		13430730: 23 SSW, rm 2311, outside cont	
Comments (see below)	None		None		None		None	
Lab ID-Version†:	1749978-1		1749979-1		1749980-1		1749981-1	
	raw ct.	spores/m3	raw ct.	spores/m3	raw ct.	spores/m3	raw ct.	spores/m3
Alternaria					2	27		
Arthrinium								
Ascospores*	5	227						
Aureobasidium								
Basidiospores*	17	627	6	320	4	213		
Bipolaris/Drechslera group								
Botrytis								
Chaetomium								
Cladosporium	7	373	4	213	22	1,170	2	107
Curvularia								
Epicoccum								
Fusarium								
Myrothecium								
Nigrospora								
Other colorless								
Penicillium/Aspergillus types†	16	853	19	1,010	27	1,440	9	480
Pithomyces								
Rusts*			1	13				
Smuts*, Periconia, Myxomycetes*								
Stachybotrys								
Stemphylium								
Torula								
Ulocladium								
Zygomycetes								
Background debris (1-4+)††	3+		3+		3+		3+	
Hyphal fragments/m3	< 13		< 13		80		< 13	
Pollen/m3	320		240		413		< 13	
Skin cells (1-4+)	None		None		None		None	
Sample volume (liters)	75		75		75		75	
TOTAL SPORE/m3		2,080		1,556		2,850		587

Comments:

* Most of these spore types are not seen with culturable methods (Andersen sampling), although some may appear as non-sporulating fungi. Most of the basidiospores are "mushroom" spores while the rusts and smuts are plant pathogens.
 † The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.
 †† Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.
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 ‡ A "Version" greater than 1 indicates amended data.

EMLab P&K

5473 Kearny Villa Road, Suite 130, San Diego, CA 92123
(858) 569-5800 Fax (858) 569-5806 www.emlab.com

Client: Biomax Environmental

Date of Sampling: 03-10-2008

C/O: Mr. Michael Polkabla

Date of Receipt: 03-12-2008

Re: 031008-01, 23 South; 450 N Street, Sacramento, CA, 23 South
Date of Report: 03-13-2008

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	13430691: 23 SSW, in containment		13430698: 23 SSE, in containment		13430693: 23 S, office 2307		13430689: 23 S, equip corridor	
Comments (see below)	None		None		None		None	
Lab ID-Version‡:	1749982-1		1749983-1		1749984-1		1749985-1	
	raw ct.	spores/m ³	raw ct.	spores/m ³	raw ct.	spores/m ³	raw ct.	spores/m ³
Alternaria							1	13
Arthrinium								
Ascospores*								
Aureobasidium								
Basidiospores*							1	53
Bipolaris/Drechslera group								
Botrytis								
Chaetomium								
Cladosporium	4	213					1	53
Curvularia								
Epicoccum								
Fusarium								
Myrothecium								
Nigrospora								
Other colorless								
Penicillium/Aspergillus types†	10	533	2	107	3	160	9	480
Pithomyces								
Rusts*								
Smuts*, Periconia, Myxomycetes*								
Stachybotrys								
Stemphylium								
Torula								
Ulocladium								
Zygomycetes								
Background debris (1-4+)††	2+		3+		3+		3+	
Hyphal fragments/m ³	< 13		< 13		< 13		< 13	
Pollen/m ³	< 13		< 13		< 13		< 13	
Skin cells (1-4+)	< 1+		1+		1+		1+	
Sample volume (liters)	75		75		75		75	
TOTAL SPORE/m³		746		107		160		599

Comments:

* Most of these spore types are not seen with culturable methods (Andersen sampling), although some may appear as non-sporulating fungi. Most of the basidiospores are "mushroom" spores while the rusts and smuts are plant pathogens.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

†† Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.

The Limit of Detection is the product of a raw count of 1 and 100 divided by the percent read. The analytical sensitivity (counts/m³) is the product of the Limit of Detection and 1000 divided by the sample volume.

‡ A "Version" greater than 1 indicates amended data.

EMLab P&K

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Client: Biomax Environmental

Date of Sampling: 03-10-2008

C/O: Mr. Michael Polkabila

Date of Receipt: 03-12-2008

Re: 031008-01, 23 South; 450 N Street, Sacramento, CA, 23 South

Date of Report: 03-13-2008

MoldRANGE™: Extended Outdoor Comparison**Outdoor Location: 13430672, Ambient front entry**

Fungi Identified	Outdoor data	Typical Outdoor Data by Date†				Typical Outdoor Data by Location‡			
		Month: March				State: CA			
	spores/m3	low	med	high	freq %	low	med	high	freq %
Generally able to grow indoors*									
Alternaria	-	7	27	210	43	7	27	230	60
Bipolaris/Drechslera group	-	7	13	120	12	7	13	120	14
Chaetomium	-	7	13	120	8	7	13	110	19
Cladosporium	373	27	320	4,300	91	53	640	6,500	98
Curvularia	-	7	13	210	7	7	13	210	7
Nigrospora	-	7	13	110	7	7	13	170	8
Penicillium/Aspergillus types	853	27	160	1,600	82	40	210	2,500	88
Stachybotrys	-	7	13	310	3	7	13	330	5
Torula	-	7	13	170	8	7	13	150	13
Seldom found growing indoors**									
Ascospores	227	13	130	2,000	74	13	110	1,800	73
Basidiospores	627	13	320	5,700	90	13	270	6,900	95
Rusts	-	7	13	320	17	7	13	270	29
Smuts, Periconia, Myxomycetes	-	7	27	310	54	8	40	470	71
TOTAL SPORES/M3	2,080								

† The Typical Outdoor Data by Date represents the typical outdoor spore levels across North America for the month indicated. The last column represents the frequency of occurrence. The low, medium, and high values represent the 2.5, 50, and 97.5 percentile values of the spore type when it is detected. For example, if the frequency of occurrence is 63% and the low value is 53, it would mean that the given spore type is detected 63% of the time and, when detected, 2.5% of the time it is present in levels above the detection limit and below 53 spores/m³. These values are updated periodically, and if enough data is not available to make a statistically meaningful assessment, it is indicated with a dash.

‡ The Typical Outdoor Data by Location represents the typical outdoor spore levels for the region indicated for the entire year. As with the Typical Outdoor Data by Date, the four columns represent the frequency of occurrence and the typical low, medium, and high concentration values for the spore type indicated. These values are updated periodically, and if enough data is not available to make a statistically meaningful assessment, it is indicated with a dash.

*The spores in this category are generally capable of growing on wet building materials in addition to growing outdoors. Building related growth is dependent upon the fungal type, moisture level, type of material, and other factors. *Cladosporium* is one of the predominant spore types worldwide and is frequently present in high numbers. *Penicillium/Aspergillus* species colonize both outdoor and indoor wet surfaces rapidly and are very easily dispersed. Other genera are usually present in lesser numbers.

**These fungi are generally not found growing on wet building materials. For example, the rusts and smuts are obligate plant pathogens. However, in each group there are notable exceptions. For example, agents of wood decay are members of the basidiomycetes and high counts of a single morphological type of basidiospore on an inside sample should be considered significant.

Interpretation of the data contained in this report is left to the client or the persons who conducted the field work. This report is provided for informational and comparative purposes only and should not be relied upon for any other purpose. "Typical outdoor data" are based on the results of the analysis of samples delivered to and analyzed by EMLab P&K and assumptions regarding the origins of those samples. Sampling techniques, contaminants infecting samples, unrepresentative samples and other similar or dissimilar factors may affect these results. In addition, EMLab P&K may not have received and tested a representative number of samples for every region or time period. EMLab P&K hereby disclaims any liability for any and all direct, indirect, punitive, incidental, special or consequential damages arising out of the use or interpretation of the data contained in, or any actions taken or omitted in reliance upon, this report.