
BioMax Environmental

Environmental Consulting and Industrial Hygiene Services

May 26th, 2008

Mr. Doug Button
Deputy Director
Real Estate Services Division
707 Third Street - 8th Floor
West Sacramento, CA 95605

Post Mitigation Clearance Assessment
Department of General Services Board of Equalization Building
450 N Street, Sacramento, California
24th Floor Men's Restroom and 2427 + 2428 Areas
450 N. Street
Sacramento, California

BioMax Environmental, LLC (BioMax) is pleased to provide The Department of General Services (DGS) with this letter summary report detailing BioMax's findings and recommendations pertaining to our post mitigation microbial inspection and sampling assessment services provided within the 24th Floor Men's Restroom and 2427 + 2428 interior areas within the Board of Equalization (BOE) building (subject building). Such locations were previously identified as containing elevated microbial contamination within building materials as noted in accordance with BioMax's assessment reports entitled Mitigative and Clean Up Procedures for Interior Electrical/Data Rooms, Janitorial Rooms, Supply Rooms, Copy Rooms, Storage Rooms, and Rest Room Areas and Mitigation Procedures for 24th Floor Rooms 2427 and 2428 dated May 7th, 2008 and April 29th, 2008, respectively.

BioMax understands that these post mitigation microbial inspection and clearance sampling assessment services were contracted with BioMax, at your request, in an effort to review and verify the successful completion of microbial mitigative efforts performed by your restoration contractor, JLS Environmental, Inc., within the previously identified mold damaged and moisture impacted areas noted within the subject building. Hence, these post mitigation assessment services are intended to assess the current site conditions wherein mitigative activities were performed to address prior moisture related damages and impacts. Following the completion of prescribed mitigative activities performed by your selected contractor, JLS, Mr. Michael A. Polkaba, CIH, REA of BioMax performed a post mitigation site inspection and sampling assessment within the affected areas of the subject building areas as noted below. BioMax's findings and conclusions pertaining to our post mitigation sampling assessment are summarized herein.

These post mitigation microbial clearance assessment services, thereby, are intended to provide a professional evaluation supported by technical sampling information verifying physical conditions wherein the successful completion of microbial removal and decontamination within the affected areas has been achieved.

SITE OBSERVATIONS

Site inspection and post mitigation assessment sampling activities were performed within rooms 2427 and 2428 on Monday, May 12th, 2008 and Monday and within the Men's Restroom on May 19th, 2008. Site access into contained and non-contained areas was facilitated on each of the noted days by Mr. Rick Boggs of JLS. On the noted days, Mr. Michael A. Polkabila, CIH, REA of BioMax performed a visual site inspection of each of the containment system barriers associated with the two 24th floor containment areas and collected a series of airborne samples within and surrounding the areas of concern the subject structures as noted below.

On-site inspection and clearance sampling assessment activities were performed by Mr. Michael A. Polkabila, CIH, REA, of BioMax in accordance with currently recognized microbial assessment and sampling guideline procedures. Mr. Polkabila has been certified in the Comprehensive Practice of Industrial Hygiene by the American Board of Industrial Hygiene and holds the right to the designation "Certified Industrial Hygienist" (CIH) under certification number CP 7104. Mr. Polkabila is also certified by the California Environmental Protection Agency (Cal/EPA) as a Class I Registered Environmental Assessor (REA) under Cal/EPA certification number 05011. A summary of significant notations and observations gathered during BioMax's site inspections and clearance assessments within each of the noted areas within the subject building are compiled as follows:

Rooms 2427 and 2428:

1. At the time of our site inspection and clearance sampling assessment of Rooms 2427 and 2428 performed on May 12th, 2008 ambient outdoor conditions both prior to and following our interior assessment consisted of sunny and mild conditions with an outdoor temperatures range between 71 and 82 degrees F and relative humidity range between 24 and 26 %. Predominant winds were noted at approximately 0-5 knots from the northwesterly direction at the time of our assessment. Interior environmental conditions within the sampled 24th Floor Room 2427 and 2428 areas consisted of a temperature range between 85 and 88 degrees F with relative humidity range of 26 to 29 percent.
2. Interior containments whereby microbial mitigative activities were previously performed by the mitigation contractor (JLS) included wall and ceiling plastic barriers erected within the impacted interior conference room and hallway areas in accordance with BioMax's protocols. Based on our inspection and review of observations within and surrounding each of the containment areas, BioMax concluded that such barrier systems provided evidence of appropriate control barriers and site protections at the time of our post mitigation assessment.

A detailed site map indicating the delineation of established (as-constructed) containment systems utilized during this procedure may be referenced from JLS site records, as necessary.

3. Based on our post mitigation inspection within and surrounding the containment areas noted above, BioMax documented the absence of visible interior indications of elevated residual moisture and/or microbial indicators (such as staining, delamination, etc.) within the remaining exposed interior walls, wall framing, and wall cavities following the performance of material removal and mitigative measures.
4. Utilization of a TraMex hand-held inductive moisture meter indicated normal moisture content within all remaining walls and building materials inspected within each of the sampled containment areas at the time of our assessment.
5. At the time of our post mitigation inspection, containment system encompassing each of the interior affected areas were observed and verified under appropriate posting and negative pressure differential. Worker and equipment entry and exit chambers comprised of a series of zippered plastic access doorways were also observed attached to the noted containment barriers consistent with BioMax's written mitigation recommendation protocols.
6. As prescribed, all identified affected interior wallboard material and carpeting materials had been removed from each of the affected areas exposing interior wall cavity framing (metal) as well as underlayment wallboard siding and flooring materials. Upon post mitigation inspection, all remaining exposed building materials associated with the interior wall and flooring systems exhibited no significant staining and/or elevated mold growth following the completion of prescribed physical mold removal and chemical decontamination procedures performed by the selected mitigation contractor on the surfaces of such exposed building materials.
7. In conjunction with our visual inspection, BioMax collected series airborne samples within and outside each of the containment areas noted in Table 1 below for subsequent comparative analysis. Such samples collected within the interior containment area were performed in an effort to identify and quantify the presence of any potential significant fugitive airborne mold spores present within (and surrounding) the containment systems following the mitigative effort.
8. BioMax also collected a series of digital images during this post mitigative inspection and sampling assessment activities to document the conditions and significant site observations gathered at this time. Such images are provided as an attachment to this summary report for further reference, as necessary.

Men's Restroom Area:

9. At the time of our site inspection and clearance sampling assessment of the Men's Restroom area performed on May 19th, 2008 ambient outdoor conditions both prior to and following our interior assessment consisted of sunny and mild conditions with an outdoor temperatures

range between 77 and 81 degrees F and relative humidity range between 34 and 42 %. Predominant winds were noted at approximately 0-5 knots from the northwesterly direction at the time of our assessment. Interior environmental conditions within the sampled 24th Floor Men's Restroom areas consisted of an interior temperature of 89 degrees F with relative humidity of 26.

10. Interior containments systems whereby microbial mitigative activities were previously performed by the mitigation contractor (JLS) included doorway and ceiling negative pressure plastic barriers erected within the impacted interior restroom area in accordance with BioMax's recommended procedures. Based on our inspection and review of observations within and surrounding the containment area, BioMax concluded that such barrier systems provided evidence of appropriate control barriers and site protections during the performance of mitigative activities and at the time of our post mitigation assessment. A detailed site map indicating the delineation of established (as-constructed) containment systems utilized during this procedure may be referenced from JLS site records, as necessary.
11. Based on our post mitigation inspection within and surrounding the 24th Floor Men's Restroom containment area noted above, BioMax documented the absence of visible interior indications of elevated residual moisture and/or microbial indicators (such as staining, delamination, etc.) within the remaining exposed interior walls, wall framing, wall cavities and flooring materials following the performance of material removal and mitigative measures.
12. Utilization of a TraMex hand-held inductive moisture meter indicated normal moisture content within all remaining walls and building materials inspected within the sampled containment area at the time of our assessment.
13. At the time of our post mitigation inspection, containment system isolating the interior affected areas were observed and verified under appropriate posting and negative pressure differential. Worker and equipment entry and exit chambers comprised of a series of zippered plastic access doorways were also observed attached to the noted containment barrier consistent with BioMax's written mitigation recommendation protocols.
14. As prescribed by BioMax, all identified affected interior sink cabinetry and affected wallboard building materials had been removed from each of the visibly affected areas exposing interior wall cavity framing (metal) as well as underlayment wallboard siding materials. Upon post mitigation inspection, all remaining exposed building materials associated with the interior wall systems exhibited no significant staining and/or elevated mold growth following the completion of prescribed physical mold removal and chemical decontamination procedures performed by the selected mitigation contractor on the surfaces of such exposed building materials.
15. In conjunction with our visual inspection, BioMax also collected series airborne samples within and outside each of the containment areas noted in Table 2 below for subsequent comparative analysis. Such samples collected within the interior containment area were

performed in an effort to identify and quantify the presence of any potential significant fugitive airborne mold spores present within (and surrounding) the containment systems following the mitigative effort.

16. BioMax also collected a series of digital images during this post mitigative inspection and sampling assessment activities to document the conditions and significant site observations gathered at this time. Such images are provided as an attachment to this summary report for further reference, as necessary.

SAMPLING PROCEDURES

On-site inspection and sampling assessment activities were conducted by Mr. Michael A. Polkabl, CIH, REA, of BioMax Environmental on the dates noted. All sampling equipment, supplies, calibration materials, and collection media were provided by BioMax as part of the performance of this scope of work. Sample collection procedures and methods were performed using aseptic sampling methods following techniques prescribed by the contracted analytical laboratory.

Spore Trap Airborne Microbial and Particulate Sampling:

The collection of airborne Spore Trap microbial samples was achieved using Zefon Air-O-Cell sampling cassette collection devices placed in each of the areas identified in Tables 1 and 2. Airborne Spore Trap samples were collected within and outside each of the containment areas at a height of approximately four feet above ground level using a tripod mounted Quick Take 15 air sampling pump manufactured by SKC. Samples were collected at a calibrated flow rate of 15 liters per minute for a total of five minutes per sample. Resultant total sample volumes, therefore, corresponded to 75 liters collected for each sample. Field calibration of the SKC air sampling pump was conducted and recorded prior to sampling activities using a field rotometer calibrated with a Bios Drycal primary standard flow meter. All spore trap air sampling and analytical procedures were performed in accordance with prescribed manufacturer guidelines as well as applicable professional certified industrial hygiene indoor air quality microbial investigation procedures and certified industrial hygiene practices.

Additional building exterior samples were also similarly collected and analyzed as part of the collection of interior samples in an effort to identify and quantify normal background microbial taxa (types), rank order, and corresponding airborne spore levels present at the time of this assessment. It should be noted that all Heating Ventilation and Air Conditioning (HVAC) systems on the 24th floor had been deactivated in October of 2007 and have remained as such throughout the prescribed mitigative effort noted herein. Hence, the collection of airborne samples performed during this clearance assessment were representative of building conditions present within each of the subject areas sampled at the time and conditions of this assessment. Sampling collection activities performed during this post mitigation clearance assessment study included the collection of identifiable airborne microbial contaminants within the representative areas noted in Tables 1 and 2 below:

Table 1. Airborne Spore Trap Sampling Locations on May 12th - Rooms 2427 and 2428:

Air Sample Number	Spore Trap Air Sampling Location
13811763	24 th floor Interior Area (Outside Containment)
13811904	Room 2427 + 2428 Containment Area
13810533	24 th Floor Hallway Outside Containment (toward South)
13811890	Northeast Building Corner of 24 th Floor Interior
13811702	Ambient Sample 23 rd Floor North Balcony
13810630	Ambient Sample 4 th Floor Garage Roof

Table 2. Airborne Spore Trap Sampling Locations on May 19th - Men's Restroom:

Air Sample Number	Spore Trap Air Sampling Location
13811883	Ambient 3 rd level Garage Area
13811939	Ambient Sample 23 rd Floor West Balcony
13811752	24 th Floor Men's Restroom
13810314	Northwest Building Hallway of 24 th Floor Interior
13811894	Northwest Cubicle Area of 24 th Floor Interior
13810312	Northern Leg Center Area of 24 th Floor Interior
13811954	Northeast Corner of 24 th Floor Interior
13810359	Eastern Hallway Near 2445
13811917	Southeast Corner Near 2410
13811944	Library East Side of 24 th Floor Interior
13810616	Office 2416 of 24 th Floor Interior
13810340	Library West Side of 24 th Floor Interior
13811882	West Hallway Near 2423

Air Sample Number	Spore Trap Air Sampling Location
13811738	Ambient Sample 4 th Floor Garage Roof

Preparation and shipping of the collected samples were accomplished in accordance with standard industrial hygiene chain of custody (COC) documentation procedures and quality assurance/quality control practices at the conclusion of sampling activities performed on each day noted. Once collected, labeled, and recorded, all samples were double sealed within airtight plastic Ziploc shipping containers and transported via Federal Express Priority Mail to Environmental Microbial Laboratories (EMLabs) in San Bruno, California. EMLabs holds current applicable analytical accreditation and specializes in microbial analytical procedures. Sampling and chain of custody records are provided as an attachment to this letter report for further reference.

ANALYTICAL FINDINGS AND CONCLUSIONS

Airborne Spore Trap Findings:

Laboratory analytical methods for the identification and enumeration of microbial (mold) taxa and particulate contaminants were conducted in accordance with prescribed analytical procedures and quality control/assurance measures. Original laboratory results including the enumeration of recognizable microbial spore and particulate types are also attached to this letter report for further detail. Analytical comments provided by the microbial laboratory regarding relative levels are noted as a semi-quantitative assessment based on historical and regional data. A summary of airborne Spore Trap microbial (mold) and particulate findings within and surrounding each of the subject containment areas are presented in Tables 3 and 4 below:

Table 3. Summary of Airborne Microbial and Particulate Findings on 5/12/08

Location Desc.	Total Mold Spores (Cts/m ³)	Background Debris (scale of 1-4)	Skin Cell Fragments (scale of 1-4)
24 th floor Interior Area (Outside Containment)	ND	1+	<1+
Room 2427 + 2428 Containment Area	ND	1+	1+
24 th Floor Hallway Outside Containment (toward South)	66	1+	<1+

Location Desc.	Total Mold Spores (CFU/m ³)	Background Debris (Scale of 1-4)	Stain Cell Fragments (Scale of 1-4)
Northeast Building Corner of 24 th Floor Interior	ND	1+	<1+
Ambient Sample 23 rd Floor North Balcony	771	3+	<1+
Ambient Sample 4 th Floor Garage Roof	1,106	3+	<1+

Table 4. Summary of Airborne Microbial and Particulate Findings on 5/19/08

Location Desc.	Total Mold Spores (CFU/m ³)	Background Debris (Scale of 1-4)	Stain Cell Fragments (Scale of 1-4)
Ambient 3 rd level Garage Area	1,617	3+	<1+
Ambient Sample 23 rd Floor West Balcony	1,840	3+	<1+
24 th Floor Men's Restroom	345	2+	1+
Northwest Building Hallway of 24 th Floor Interior	ND	1+	1+
Northwest Cubicle Area of 24 th Floor Interior	66	2+	1+
Northern Leg Center Area of 24 th Floor Interior	107	2+	1+
Northeast Corner of 24 th Floor Interior	66	2+	1+
Eastern Hallway Near 2445	66	2+	1+
Southeast Corner Near 2410	ND	1+	1+

Location Desc	Total Mold Spores (CFU/m ³)	Background Debris (scale of 1-4)	Spore Cell Fragments (scale of 1-4)
Library East Side of 24 th Floor Interior	53	2+	1+
Office 2416 of 24 th Floor Interior	ND	2+	1+
Library West Side of 24 th Floor Interior	ND	2+	1+
West Hallway Near 2423	ND	2+	1+
Ambient Sample 4 th Floor Garage Roof	1,227	3+	<1+

The analytical findings presented in Summary Tables 3 and 4 clearly indicate the presence of significantly lower concentrations of microbial (mold) spores measured within each of the interior containment area samples when compared to the levels currently measured within the corresponding ambient outside environment. Analytical findings also indicate similar fungal taxa distribution (mold types) and rank order (predominant taxa) of molds identified within the mitigated areas as well as the adjacent non containment areas sampled and evaluated. Such airborne samples collected and assessed within the surrounding non-impacted areas also provide reasonable evidence indicating that current containment barriers and mitigative control procedures utilized during these 24th Floor efforts have successfully contained and controlled mold spore and particulate debris within the identified containment areas. Also particularly worthy of note, were the absence of elevated levels of hydrophilic (moisture loving) airborne mold taxa such as Penicillium, Aspergillus, and Stachybotrys present within the containment barriers following the mitigative efforts as comparatively summarized in this assessment report.

Although there are currently no regulatory standards or limits pertaining to allowable airborne fungal concentrations (for any mold taxa) present in indoor environments, there is a general consensus among indoor air quality experts that microbial contamination found within "typical healthy" living and/or working spaces are generally similar in kind and present at levels which are below those found in the corresponding native outside environment. BioMax believes that the absence of elevated residual moisture, absence of significant visible residual mold and/or staining, and relatively fewer total airborne mold levels with typical taxa and rank order distribution following mitigative clean-up activities are consistent with these generally acceptable conditions. BioMax, therefore, believes that these findings provide reasonable evidence indicating that current microbial clean-up measures have successfully mitigated and contained

mold contamination within previously affected containment areas and previously affected materials to acceptable levels.

Based on these findings, BioMax believes that the current site conditions and verification sampling performed within the mitigated areas meets the mitigative clearance criteria established for these activities as presented in BioMax's Post Mitigation Clearance Assessment Protocols dated February 15th, 2008 as reviewed and approved by BOE and their environmental consultant, Hygientech, Inc. (HTI). Therefore, BioMax believes that achievement of such criteria supports BioMax's professional determination and recommendation that the previously impacted areas may be considered acceptable for normal reconstruction following standard industry building maintenance practices. Areas evaluated through similar airborne sampling throughout the surrounding 24th Floor areas also indicated current microbial airborne levels which are consistent with normal "unremarkable" background levels as no current significant data or current evidence has been identified to the contrary.

Airborne Particulate Findings:

Analytical findings pertaining to the airborne particulates consisting primarily of debris, pollen, and skin cell fragments identified within the collected air samples within and surrounding the previously impacted areas also provide reasonable evidence indicating that current particulate clean-up and mitigative control measures have successfully controlled and contained particulate debris within the identified containment areas to normal background ranges.

Although, there are similarly no currently applicable regulatory standards pertaining to allowable particulate levels with which to compare, it is BioMax's professional opinion that interior particulate levels should continue to be minimized wherever possible. Therefore, additional (and ongoing) recommendations for optional particulate control measures have been provided at the end of this report for client consideration.

RECOMMENDATIONS

Based on the findings and conclusions presented in this report, BioMax believes that the current airborne microbial levels sampled and analyzed from within the identified 24th Floor Men's Restroom and Rooms 2427 and 2428 provides no significant evidence of elevated residual microbial contamination or airborne migration following the completion of prescribed microbial mitigative measures. BioMax's 24th Floor-wide airborne sampling assessment similarly performed in areas outside the noted containment barriers also indicated current airborne levels which are consistent with normal "unremarkable" microbial spore levels. Hence, based on our direct site observations, measurements, and review of these findings at this time, BioMax believes that the previously affected areas (and surrounding 24th Floor-wide areas) may be considered acceptable for general reconstruction following prudent reconstruction practices with the implementation of the noted additional measures discussed below. Therefore, based on these findings, BioMax recommends consideration of the following post-mitigation measures and actions:

1. BioMax believes that current airborne microbial (mold) levels and mold types were identified at levels which are believed to fall within generally acceptable ranges and parameters at present. Hence, BioMax recommends that no further airborne microbial sampling activities are warranted within the previously mitigated areas nor within the surrounding 24th Floor wide areas at this time. Certainly, due to the knowledge that microbial contamination, by nature, may change over time due to additional moisture intrusion, favorable growth conditions, and changing environments, these recommendations are subject to revision in the event that such conditions and/or environments arise.
2. During the performance of interior reconstruction activities, BioMax recommends that a qualified and experienced building inspector/contractor be utilized to verify the current functional integrity of all exposed and applicable remaining plumbing, flashing, sealing, and drainage systems in accordance with current building codes and construction practices. Any identified deficiencies should be appropriately documented, corrected, and then functionally verified (tested) prior to subsequent reconstruction and commercial use. Certainly, the establishment/installation of any additional engineering controls (as identified through additional professional engineering consultation) should also be performed and implemented in accordance with applicable standards, building codes, and ordinances, as necessary.
3. Due to the anticipated forthcoming renovation and reuse of the men's restroom area, as an additional precautionary measure, BioMax recommends that all remaining interior wall cavity wallboard materials be reconstructed utilizing an appropriate grade of sheetrock materials where moisture barrier and microbial resistant properties are desired. As a client option, any interior sheetrock surfaces may also receive the application of a spray-on sealant with microbial growth inhibitors prior to wall cavity construction. The application of such sealant (if desired) would provide an additional benefit in quickly identifying material surfaces and areas which were treated as part of this mitigative effort for future forensic assessment. Sealant application (if desired) should certainly be performed by your selected contractor in accordance with all product manufacturer's use specifications and application guidelines.
4. BioMax recommends that all reconstruction of interior structural building materials should only be undertaken utilizing high quality, visibly clean (hand selected) construction grade building materials obtained from reputable commercial sources and which are believed and visually free from elevated microbial contamination and/or elevated moisture content. Building materials, which are notably moist and/or visibly stained, should not be used during the reconstruction undertaken within the subject residence. BioMax recommends that all current plastic barriers (as established during this mitigative effort) may remain during such reconstruction so as to minimize the potential transmission of associated construction dust and debris within the subject structure.
5. As previously noted in is report, detectable levels of airborne particulates consisting primarily of skin cell fragments, pollen, and general debris particles were identified within the sampled interior areas surrounding the containment systems. Hence, and as an additional

precautionary measure due to the presence of such materials, BioMax recommends that DGS considers the performance of supplemental post construction vacuuming and cleaning following the completion of interior painting, re-carpeting, and renovation activities.

- 6. BioMax believes that any potential transmission and accumulation of the identified indoor airborne particulates may also be significantly reduced (if desired) on an immediate and ongoing basis through the implementation and use of routine HEPA filtered vacuuming and damp-wipe O&M cleaning methods employed by DGS maintenance personnel. BioMax's experience has indicated that these relatively simple measures and methods have been shown to significantly reduce the accumulation of settled particulate debris on an immediate and ongoing basis if so desired.
- 7. Reasonable additional assessment and investigative measures may also be required upon the identification of new or previously undiscovered materials and/or information related to moisture/microbial impacts within the subject building structures, as necessary. Any occurrence and/or re-occurrence of moisture intrusion following routine O&M and/or general reconstruction within the subject building should also be reviewed and addressed through professional consultation, as necessary. BioMax is certainly prepared to provide such additional consultation pertaining to these and any follow-up investigative measures as necessary and upon request.

BioMax believes that the conclusions and recommendations outlined above are consistent with standard industry microbial mitigative and assessment practices as well as prudent industrial hygiene hazard control methods. Please do not hesitate to contact our offices directly at (510) 724-3100 if you have any additional questions, comments, or require further assistance regarding this important matter.

Sincerely,

Michael A. Polkabila, CIH, REA
Vice President, Principal



LIMITATIONS

Please note that the professional opinions presented in this review are intended for the sole use of the California State Department of General Services (DGS) and their designated beneficiaries. No other party should rely on the information contained herein without the prior written consent of BioMax Environmental and DGS. The professional opinions provided herein are based on BioMax's review and understanding of current site information and observed site conditions present within the areas inspected at the time these services were performed. Professional recommendations provided as part of this limited scope of work are intended for client consideration only and are not intended as a professional or regulatory mandate. Implementation of any of the above measures or recommendations does not, in any way, warrant the day-to-day health and/or safety of building occupants, residents, site workers, nor regulatory or building code compliance status during normal and changing environmental conditions. As microbial contamination, by nature, may change over time due to additional moisture intrusion, favorable growth conditions, and changing environments, the findings of this report are subject to change in the event that such conditions and/or environments arise. Also, the professional opinions expressed here are subject to revision in the event that new or previously undiscovered information is obtained or uncovered.

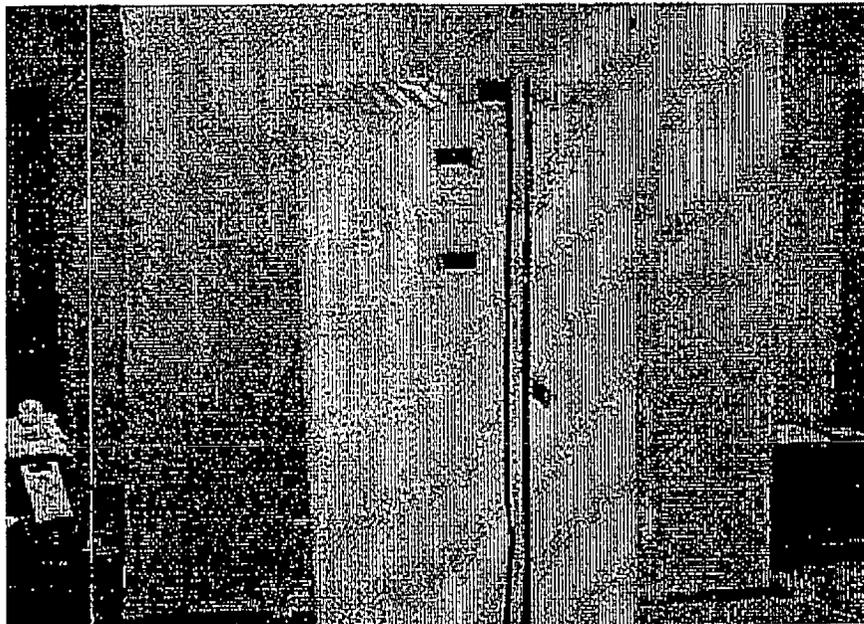
The information contained in this and any other applicable communication is for consideration purposes only. It is not intended, nor should it be construed as providing legal advice or warranting any level of safety or regulatory compliance. The sole purpose of such information is to assist with the anticipation, identification, evaluation and control of elevated and/or unnecessary health of physical hazards. Any action taken based on this information, including but not limited to opinions, suggestions and recommendations, whether implied or expressed, is the sole responsibility of the individual taking the action. The management of acceptable health and safety is criteria dependent and situation specific in nature, therefore requiring extensive knowledge and prudent value assessments so as to be properly determined and maintained.

These services were performed by BioMax in accordance with generally accepted professional industrial hygiene principals, practices, and standards of care. Under the existing Industrial Hygiene Definition and Registration Act, all reports, opinions or official documents prepared by a Certified Industrial Hygienist (CIH) constitutes an expression of professional opinion regarding those facts or findings which are subject of a certification and does not constitute a warranty or guarantee, either expressed or implied.

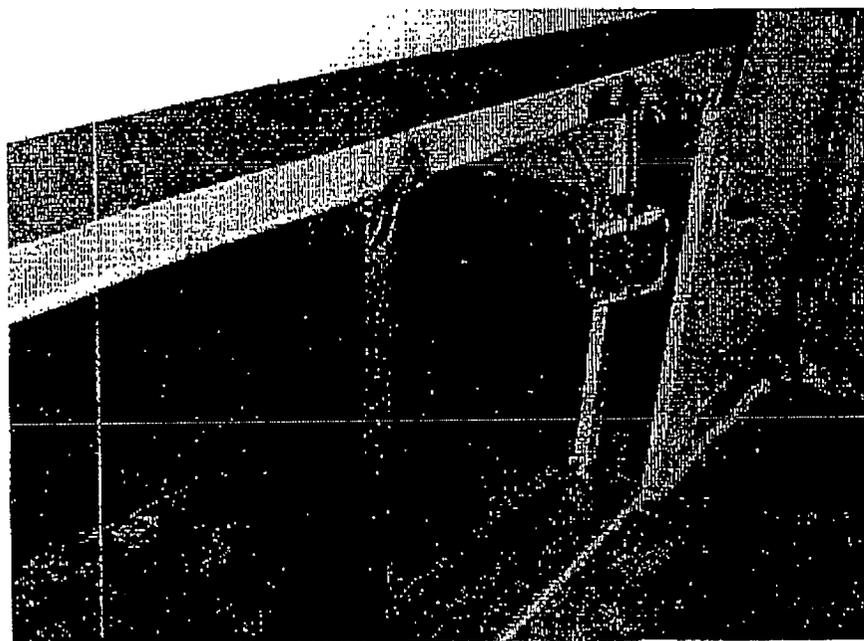
Attachment A: Digital Images
May 19th, 2008
BOE Building 2427 and 2428 Clearance
Sacramento, CA

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[Click here for color photos](#)



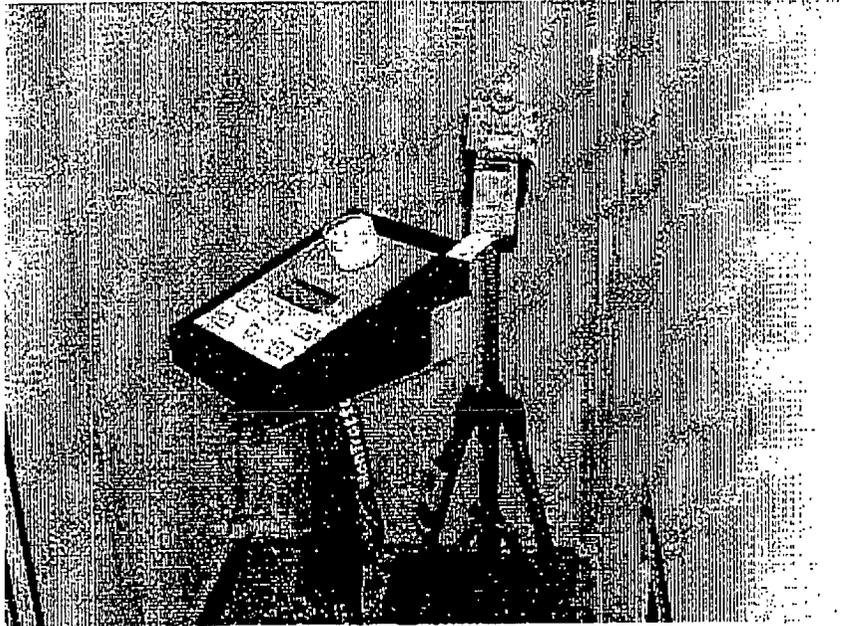
1) Image of containment leading to area 2427 within BOE Building (Subject Building) located at 450 N Street, Sacramento, California at time of assessment.



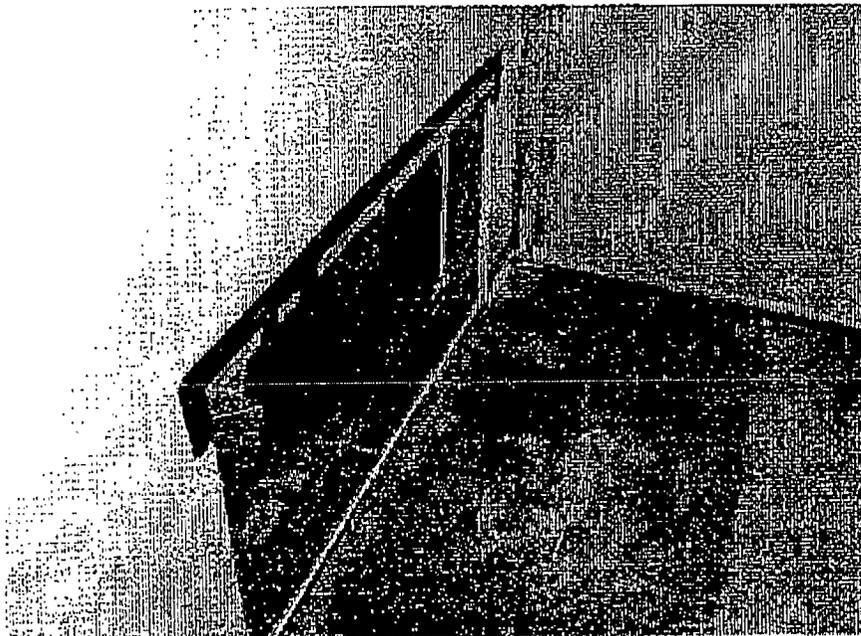
2) Image of wallboard removal within 2427 at time of clearance inspection and sampling assessment.

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Sacramento, CA

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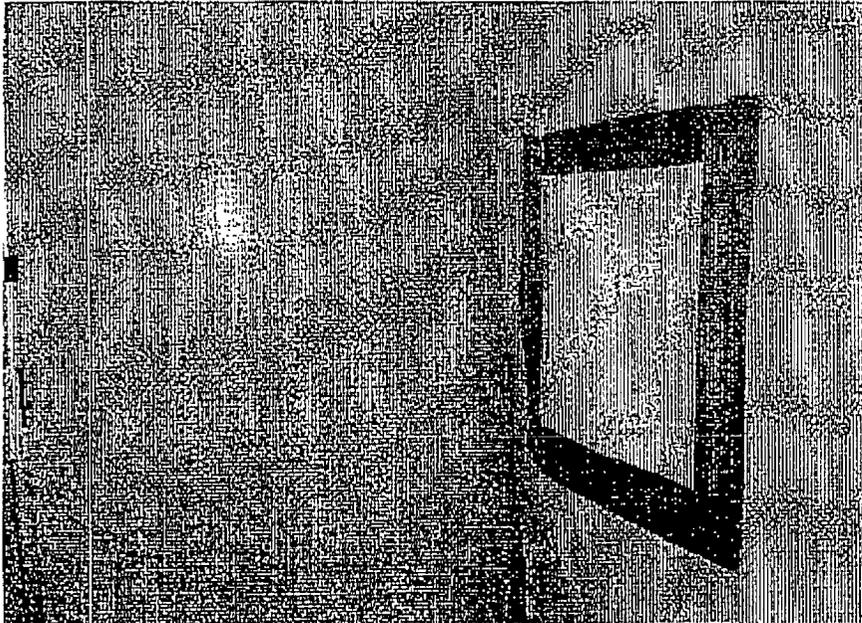
3) Image of air sampling equipment staged by BioMax and HTI within containment area 2427 at time of assessment.



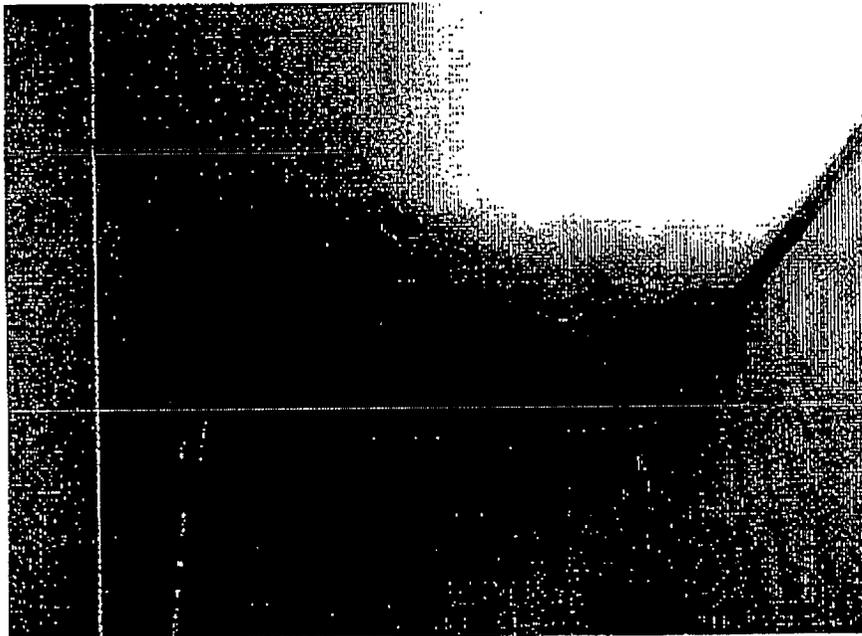
4) Image of physical wallboard removal as viewed from 2427.

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Sacramento, CA

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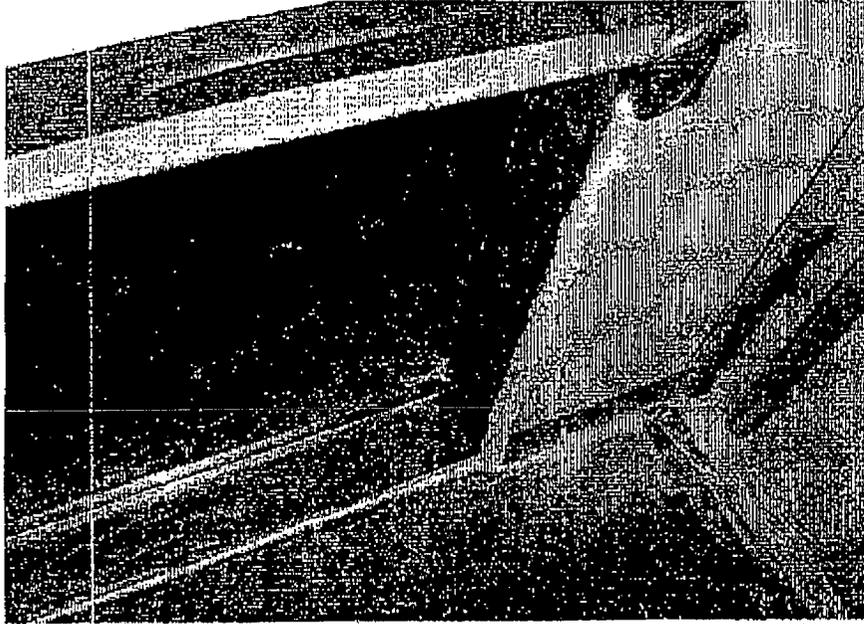
- 5) Image within containment system of filtered intake air element and location within 2427 plastic barriers at time of assessment.



- 6) Additional image within 2427 containment of ceiling barrier system and support structures as prescribed by BioMax.

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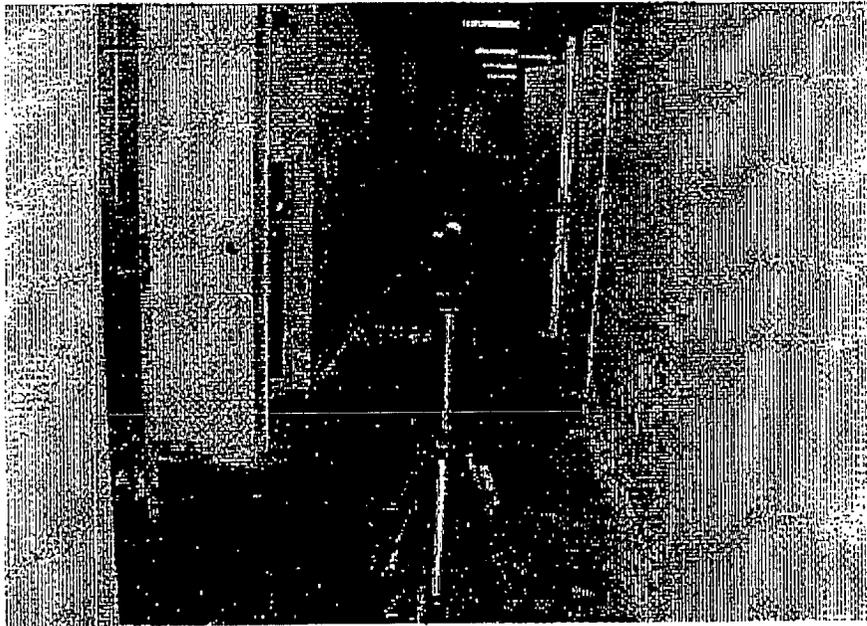
7) Close-up image of wall framing and sheetrock edge material within 2427 containment system.



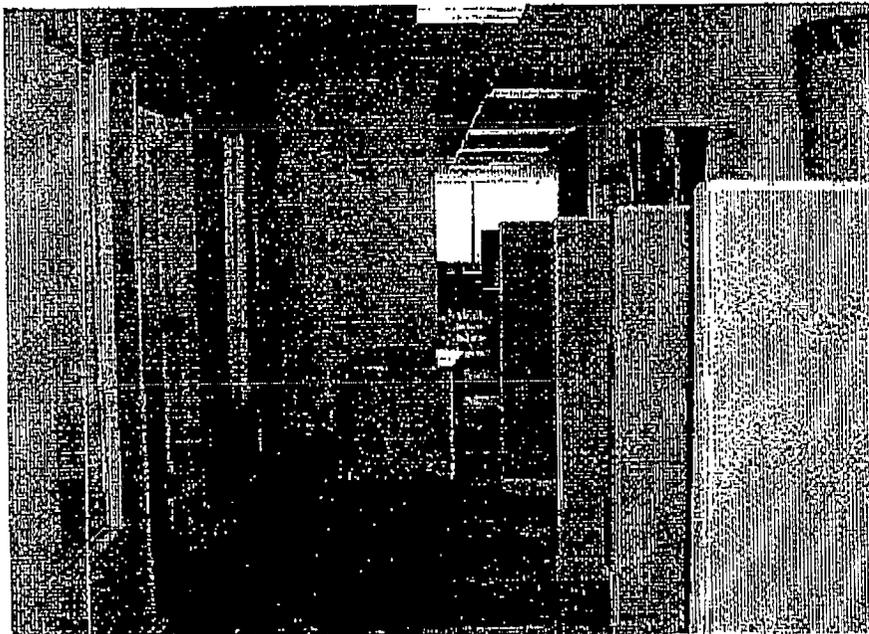
8) Image of left wall side within 2427 containment space located along west hallway adjacent to containment area.

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Sacramento, CA

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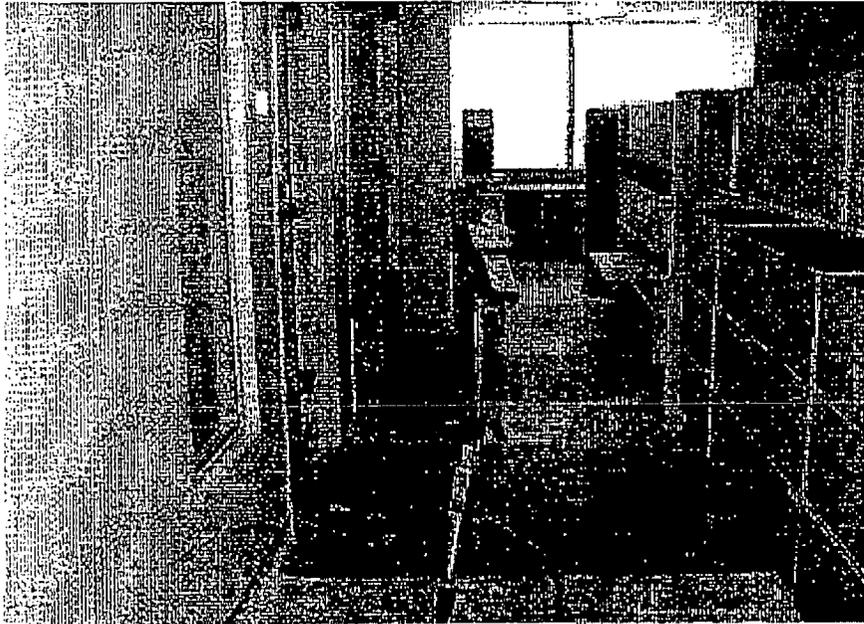
9) Image of air sampling equipment location within hallway outside containment area at time of assessment.



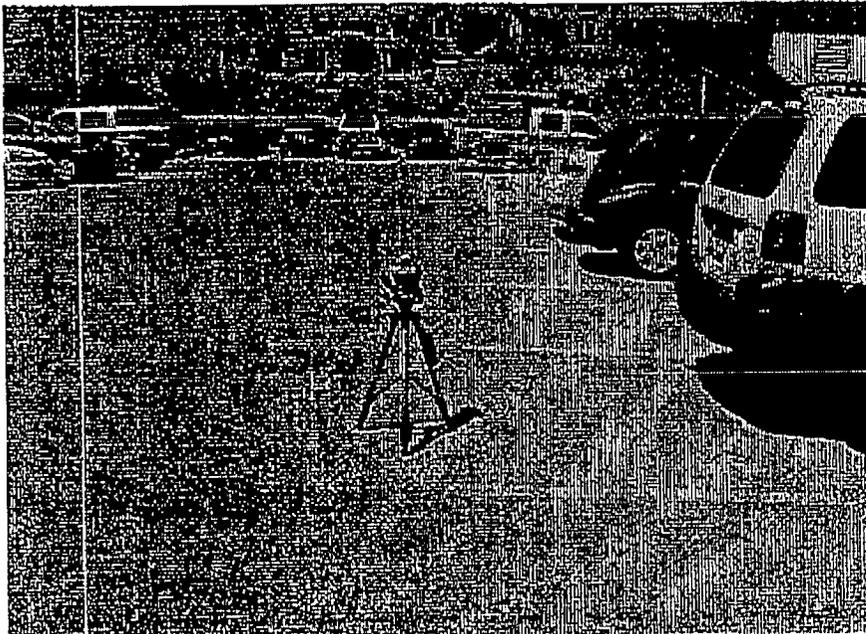
10) Image in hallway looking toward northeastern corner of building where air sampling location was performed for comparative purposes.

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Sacramento, CA

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11) Image of air sampling equipment / location within hallway outside containment area at time of assessment.



12) Image of ambient air sampling location on 4th floor garage rooftop as performed for ambient background comparative purposes.

**EMLab P&K**

Report for:

Mr. Michael Polkabila
Biomax Environmental
775 San Pablo Ave.
Pinole, CA 94564

Regarding: Project: 051908-02
EML ID: 424021

Approved by:

Lab Manager
Dr. Kamashwaran Ramanathan

Dates of Analysis:
Spore trap analysis: 05-21-2008

Project SOPs: Spore trap analysis (1100000)

This coversheet is included with your report in order to comply with ALHA and ISO accreditation requirements.

For clarity, we report the number of significant digits as calculated; but, due to the nature of this type of biological data, the number of significant digits that is used for interpretation should generally be one or two. All samples were received in acceptable condition unless noted in the Report Comments portion in the body of the report. Due to the nature of the analyses performed, field blank corrections of results is not a standard practice. The results relate only to the items tested.

EMLab P&K ("the Company") shall have no liability to the client or the client's customer with respect to decisions or recommendations made, actions taken or courses of conduct implemented by either the client or the client's customer as a result of or based upon the Test Results. In no event shall the Company be liable to the client with respect to the Test Results except for the Company's own willful misconduct or gross negligence nor shall the Company be liable for incidental or consequential damages or lost profits or revenues to the fullest extent such liability may be disclaimed by law, even if the Company has been advised of the possibility of such damages, lost profits or lost revenues. In no event shall the Company's liability with respect to the Test Results exceed the amount paid to the Company by the client therefor.

Document Number: 200091 - Revision Number: 5

EMLab P&K

1150 Bayhill Drive, Suite 100, San Bruno, CA 94066
(650) 829-5800 Fax (650) 829-5852 www.emlab.com

Client: Biomax Environmental
C/O: Mr. Michael Polkabila
Re: 051908-02

Date of Sampling: 05-19-2008
Date of Receipt: 05-20-2008
Date of Report: 05-21-2008

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	13811883: Ambient, 3rd level garage		13811939: Ambient, 23rd west		13811752: Men's restroom		13810314: NW hallway	
Comments (see below)	None		None		None		None	
Lab ID-Version†:	1863915-1		1863916-1		1863917-1		1863918-1	
	raw ct.	spores/m3	raw ct.	spores/m3	raw ct.	spores/m3	raw ct.	spores/m3
<i>Alternaria</i>			2	27				
<i>Arthrinium</i>								
Ascospores*	2	107	1	13	1	53		
<i>Aureobasidium</i>								
Basidiospores*	2	107	2	107				
<i>Beltrania</i>	1	13						
<i>Bipolaris/Drechslera</i> group								
<i>Botrytis</i>								
<i>Chaetomium</i>								
<i>Cladosporium</i>	23	1,230	27	1,440	1	53		
<i>Curvularia</i>								
<i>Epicoccum</i>								
<i>Fusarium</i>								
<i>Myrothecium</i>								
<i>Nigrospora</i>								
Other brown					1	13		
Other colorless								
<i>Penicillium/Aspergillus</i> types‡			1	53	4	213		
<i>Pithomyces</i>								
Rusts*					1	13		
Smuts*, <i>Periconia</i> , <i>Myxomycetes</i> *	6	80	14	187				
<i>Stachybotrys</i>								
<i>Stemphylium</i>								
<i>Torula</i>	6	80	1	13				
<i>Ulocladium</i>								
Zygomycetes								
Background debris (1-4+)††	3+		3+		2+		1+	
Hyphal fragments/m3	40		< 13		< 13		< 13	
Pollen/m3	40		80		13		< 13	
Skin cells (1-4+)	< 1+		< 1+		1+		1+	
Sample volume (liters)	75		75		75		75	
TOTAL SPORE/m3		1,617		1,840		345		< 13

Comments:

* Most of these spore types are not seen with culturable methods (Andersen sampling), although some may appear as non-sporulating fungi. Most of the basidiospores are "mushroom" spores while the rusts and smuts are plant pathogens.

‡ The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

†† Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for sample volumes when evaluating dust levels.

The Limit of Detection is the product of a raw count of 1 and 100 divided by the percent read. The analytical sensitivity (counts/m3) is the product of the Limit of Detection and 1000 divided by the sample volume.

† A "Version" greater than 1 indicates amended data.

EMLab P&K

1150 Bayhill Drive, Suite 100, San Bruno, CA 94066
(650) 829-5800 Fax (650) 829-5852 www.emlab.com

Client: Biomax Environmental
C/O: Mr. Michael Polkabila
Re: 051908-02

Date of Sampling: 05-19-2008
Date of Receipt: 05-20-2008
Date of Report: 05-21-2008

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	13811894: NW cubicle area		13810312: North leg center area		13811954: Northeast corner		13810359: Eastern hallway near 2445	
Comments (see below)	None		None		None		None	
Lab ID-Version†:	1863919-1		1863920-1		1863921-1		1863922-1	
	raw ct.	spores/m ³	raw ct.	spores/m ³	raw ct.	spores/m ³	raw ct.	spores/m ³
Alternaria								
Arthrinium								
Ascospores*								
Aureobasidium								
Basidiospores*								
Beltrania								
Bipolaris/Drechslera group								
Botrytis								
Chaetomium								
Cladosporium			2	107	1	53	1	53
Curvularia								
Epicoccum								
Fusarium								
Myrothecium								
Nigrospora								
Other brown								
Other colorless								
Penicillium/Aspergillus types‡	1	53						
Pithomyces								
Rusts*					1	13		
Smuts*, Periconia, Myxomycetes*	1	13					1	13
Stachybotrys								
Stemphylium								
Torula								
Ulocladium								
Zygomycetes								
Background debris (1-4+)††	2+		2+		2+		2+	
Hyphal fragments/m ³	< 13		< 13		< 13		< 13	
Pollen/m ³	< 13		13		13		27	
Skin cells (1-4+)	1+		1+		1+		1+	
Sample volume (liters)	75		75		75		75	
TOTAL SPORE/m³		66		107		66		66

Comments:

* Most of these spore types are not seen with culturable methods (Andersen sampling), although some may appear as non-sporulating fungi. Most of the basidiospores are "mushroom" spores while the rusts and smuts are plant pathogens.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paeecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

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EMLab ID: 424021, Page 2 of 4

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Client: Biomax Environmental
C/O: Mr. Michael Polkabila
Re: 051908-02

Date of Sampling: 05-19-2008
Date of Receipt: 05-20-2008
Date of Report: 05-21-2008

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	13811917: SE corner hallway near 2410	13811944: Library E	13810616: 2416 office, south side center	13810340: Library west				
Comments (see below)	None	None	None	None				
Lab ID-Version†:	1863923-1	1863924-1	1863925-1	1863926-1				
	raw ct.	spores/m3	raw ct.	spores/m3	raw ct.	spores/m3	raw ct.	spores/m3
Alternaria								
Arthrinium								
Ascospores*								
Aureobasidium								
Basidiospores*								
Beltrania								
Bipolaris/Drechslera group								
Botrytis								
Chaetomium								
Cladosporium			1	53				
Curvularia								
Epicoccum								
Fusarium								
Myrothecium								
Nigrospora								
Other brown								
Other colorless								
Penicillium/Aspergillus types†								
Pithomyces								
Rusts*								
Smuts*, Periconia, Myxomycetes*								
Stachybotrys								
Stemphylium								
Torula								
Ulocladium								
Zygomycetes								
Background debris (1-4+)††	1+		2+		2+		2+	
Hypheal fragments/m3	< 13		< 13		< 13		< 13	
Pollen/m3	< 13		13		< 13		13	
Skin cells (1-4+)	1+		1+		1+		1+	
Sample volume (liters)	75		75		75		75	
TOTAL SPORE/m3		< 13		53		< 13		< 13

Comments:

* Most of these spore types are not seen with culturable methods (Andersen sampling), although some may appear as non-sporulating fungi. Most of the basidiospores are "mushroom" spores while the rusts and smuts are plant pathogens.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paccilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

†† Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels. The Limit of Detection is the product of a raw count of 1 and 100 divided by the percent read. The analytical sensitivity (counts/m3) is the product of the Limit of Detection and 1000 divided by the sample volume.

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Re: 051908-02

Date of Sampling: 05-19-2008
Date of Receipt: 05-20-2008
Date of Report: 05-21-2008

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	13811882: Hallway west near 2423		13811738: Ambient, level 1 main entry	
Comments (see below)	None		None	
Lab ID-Version†:	1863927-1		1863928-1	
	raw ct.	spores/m3	raw ct.	spores/m3
Alternaria				
Arthrinium				
Ascospores*				
Aureobasidium				
Basidiospores*				
Beltrania				
Bipolaris/Drechslera group				
Botrytis				
Chaetomium			1	13
Cladosporium			15	800
Curvularia				
Epicoccum				
Fusarium				
Myrothecium				
Nigrospora				
Other brown				
Other colorless				
Penicillium/Aspergillus types†			3	160
Pithomyces				
Rusts*				
Smuts*, Periconia, Myxomycetes*			11	147
Stachybotrys				
Stemphylium				
Tomia			8	107
Ulocladium				
Zygomycetes				
Background debris (1-4+)‡	2+		3+	
Hyphal fragments/m3	< 13		40	
Pollen/m3	< 13		107	
Skin cells (1-4+)	1+		< 1+	
Sample volume (liters)	75		75	
TOTAL SPORE/m3		< 13		1,227

Comments:

* Most of these spore types are not seen with culturable methods (Andersen sampling), although some may appear as non-sporulating fungi. Most of the basidiospores are "mushroom" spores while the rusts and smuts are plant pathogens.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paccilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

‡‡ Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.

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Re: 051908-02

Date of Sampling: 05-19-2008
Date of Receipt: 05-20-2008
Date of Report: 05-21-2008

MoldRANGE™: Extended Outdoor Comparison**Outdoor Location: 13811883, Ambient, 3rd level garage**

Fungi Identified	Outdoor data	Typical Outdoor Data by Date†				Typical Outdoor Data by Location‡			
		Month: May				State: CA			
	spores/m3	low	med	high	freq %	low	med	high	freq %
Generally able to grow indoors*									
Alternaria	-	7	33	320	62	7	27	230	60
Bipolaris/Drechslera group	-	7	13	120	16	7	13	120	14
Chaetomium	-	7	13	100	15	7	13	110	19
Cladosporium	1,230	53	530	6,900	96	53	640	6,400	98
Curvularia	-	7	13	360	8	7	13	210	7
Nigrospora	-	7	13	120	7	7	13	170	8
Penicillium/Aspergillus types	-	27	160	1,700	80	40	210	2,500	88
Stachybotrys	-	7	13	230	4	7	13	300	5
Torula	80	7	13	170	15	7	13	150	13
Seldom found growing indoors**									
Ascospores	107	13	160	5,300	81	13	110	1,800	72
Basidiospores	107	13	270	6,900	93	13	240	6,700	94
Beltrania	13	7	13	53	1	7	13	53	2
Rusts	-	7	25	330	29	7	13	270	29
Smuts, Periconia, Myxomycetes	80	7	53	1,000	76	8	40	470	71
TOTAL SPORES/M3	1,617								

† The Typical Outdoor Data by Date represents the typical outdoor spore levels across North America for the month indicated. The last column represents the frequency of occurrence. The low, medium, and high values represent the 2.5, 50, and 97.5 percentile values of the spore type when it is detected. For example, if the frequency of occurrence is 63% and the low value is 53, it would mean that the given spore type is detected 63% of the time and, when detected, 2.5% of the time it is present in levels above the detection limit and below 53 spores/m³. These values are updated periodically, and if enough data is not available to make a statistically meaningful assessment, it is indicated with a dash.

‡ The Typical Outdoor Data by Location represents the typical outdoor spore levels for the region indicated for the entire year. As with the Typical Outdoor Data by Date, the four columns represent the frequency of occurrence and the typical low, medium, and high concentration values for the spore type indicated. These values are updated periodically, and if enough data is not available to make a statistically meaningful assessment, it is indicated with a dash.

*The spores in this category are generally capable of growing on wet building materials in addition to growing outdoors. Building related growth is dependent upon the fungal type, moisture level, type of material, and other factors. *Cladosporium* is one of the predominant spore types worldwide and is frequently present in high numbers. *Penicillium/Aspergillus* species colonize both outdoor and indoor wet surfaces rapidly and are very easily dispersed. Other genera are usually present in lesser numbers.

**These fungi are generally not found growing on wet building materials. For example, the rusts and smuts are obligate plant pathogens. However, in each group there are notable exceptions. For example, agents of wood decay are members of the basidiomycetes and high counts of a single morphological type of basidiospore on an inside sample should be considered significant.

Interpretation of the data contained in this report is left to the client or the persons who conducted the field work. This report is provided for informational and comparative purposes only and should not be relied upon for any other purpose. "Typical outdoor data" are based on the results of the analysis of samples delivered to and analyzed by EMLab P&K and assumptions regarding the origins of those samples. Sampling techniques, contaminants infecting samples, unrepresentative samples and other similar or dissimilar factors may affect these results. In addition, EMLab P&K may not have received and tested a representative number of samples for every region or time period. EMLab P&K hereby disclaims any liability for any and all direct, indirect, punitive, incidental, special or consequential damages arising out of the use or interpretation of the data contained in, or any actions taken or omitted in reliance upon, this report.

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Date of Sampling: 05-19-2008
Date of Receipt: 05-20-2008
Date of Report: 05-21-2008

MoldRANGE™: Extended Outdoor Comparison

Outdoor Location: 13811939, Ambient, 23rd west

Fungi Identified	Outdoor data	Typical Outdoor Data by Date†				Typical Outdoor Data by Location‡			
		Month: May				State: CA			
	spores/m3	low	med	high	freq %	low	med	high	freq %
Generally able to grow indoors*									
Alternaria	27	7	33	320	62	7	27	230	60
Bipolaris/Drechslera group	-	7	13	120	16	7	13	120	14
Chaetomium	-	7	13	100	15	7	13	110	19
Cladosporium	1,440	53	530	6,900	96	53	640	6,400	98
Curvularia	-	7	13	360	8	7	13	210	7
Nigrospora	-	7	13	120	7	7	13	170	8
Penicillium/Aspergillus types	53	27	160	1,700	80	40	210	2,500	88
Stachybotrys	-	7	13	230	4	7	13	300	5
Torula	13	7	13	170	15	7	13	150	13
Seldom found growing indoors**									
Ascospores	13	13	160	5,300	81	13	110	1,800	72
Basidiospores	107	13	270	6,900	93	13	240	6,700	94
Beltrania	-	7	13	53	1	7	13	53	2
Rusts	-	7	25	330	29	7	13	270	29
Smuts, Periconia, Myxomycetes	187	7	53	1,000	76	8	40	470	71
TOTAL SPORES/M3	1,840								

† The Typical Outdoor Data by Date represents the typical outdoor spore levels across North America for the month indicated. The last column represents the frequency of occurrence. The low, medium, and high values represent the 2.5, 50, and 97.5 percentile values of the spore type when it is detected. For example, if the frequency of occurrence is 63% and the low value is 53, it would mean that the given spore type is detected 63% of the time and, when detected, 2.5% of the time it is present in levels above the detection limit and below 53 spores/m3. These values are updated periodically, and if enough data is not available to make a statistically meaningful assessment, it is indicated with a dash.

‡ The Typical Outdoor Data by Location represents the typical outdoor spore levels for the region indicated for the entire year. As with the Typical Outdoor Data by Date, the four columns represent the frequency of occurrence and the typical low, medium, and high concentration values for the spore type indicated. These values are updated periodically, and if enough data is not available to make a statistically meaningful assessment, it is indicated with a dash.

*The spores in this category are generally capable of growing on wet building materials in addition to growing outdoors. Building related growth is dependent upon the fungal type, moisture level, type of material, and other factors. *Cladosporium* is one of the predominant spore types worldwide and is frequently present in high numbers. *Penicillium/Aspergillus* species colonize both outdoor and indoor wet surfaces rapidly and are very easily dispersed. Other genera are usually present in lesser numbers.

**These fungi are generally not found growing on wet building materials. For example, the rusts and smuts are obligate plant pathogens. However, in each group there are notable exceptions. For example, agents of wood decay are members of the basidiomycetes and high counts of a single morphological type of basidiospore on an inside sample should be considered significant.

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Re: 051908-02

Date of Sampling: 05-19-2008
Date of Receipt: 05-20-2008
Date of Report: 05-21-2008

MoldRANGEM™: Extended Outdoor Comparison**Outdoor Location: 13811738, Ambient, level 1 main entry**

Fungi Identified	Outdoor data	Typical Outdoor Data by Date†				Typical Outdoor Data by Location‡			
		Month: May				State: CA			
	spores/m3	low	med	high	freq %	low	med	high	freq %
Generally able to grow indoors*									
Alternaria		7	33	320	62	7	27	230	60
Bipolaris/Drechslera group		7	13	120	16	7	13	120	14
Chaetomium	13	7	13	100	15	7	13	110	19
Cladosporium	800	53	530	6,900	96	53	640	6,400	98
Curvularia		7	13	360	8	7	13	210	7
Nigrospora		7	13	120	7	7	13	170	8
Penicillium/Aspergillus types	160	27	160	1,700	80	40	210	2,500	88
Stachybotrys		7	13	230	4	7	13	300	5
Torula	107	7	13	170	15	7	13	150	13
Seldom found growing indoors**									
Ascospores		13	160	5,300	81	13	110	1,800	72
Basidiospores		13	270	6,900	93	13	240	6,700	94
Beltrania		7	13	53	1	7	13	53	2
Rusts		7	25	330	29	7	13	270	29
Smuts, Periconia, Myxomycetes	147	7	53	1,000	76	8	40	470	71
TOTAL SPORES/M3	1,227								

† The Typical Outdoor Data by Date represents the typical outdoor spore levels across North America for the month indicated. The last column represents the frequency of occurrence. The low, medium, and high values represent the 2.5, 50, and 97.5 percentile values of the spore type when it is detected. For example, if the frequency of occurrence is 63% and the low value is 53, it would mean that the given spore type is detected 63% of the time and, when detected, 2.5% of the time it is present in levels above the detection limit and below 53 spores/m³. These values are updated periodically, and if enough data is not available to make a statistically meaningful assessment, it is indicated with a dash.

‡ The Typical Outdoor Data by Location represents the typical outdoor spore levels for the region indicated for the entire year. As with the Typical Outdoor Data by Date, the four columns represent the frequency of occurrence and the typical low, medium, and high concentration values for the spore type indicated. These values are updated periodically, and if enough data is not available to make a statistically meaningful assessment, it is indicated with a dash.

*The spores in this category are generally capable of growing on wet building materials in addition to growing outdoors. Building related growth is dependent upon the fungal type, moisture level, type of material, and other factors. *Cladosporium* is one of the predominant spore types worldwide and is frequently present in high numbers. *Penicillium/Aspergillus* species colonize both outdoor and indoor wet surfaces rapidly and are very easily dispersed. Other genera are usually present in lesser numbers.

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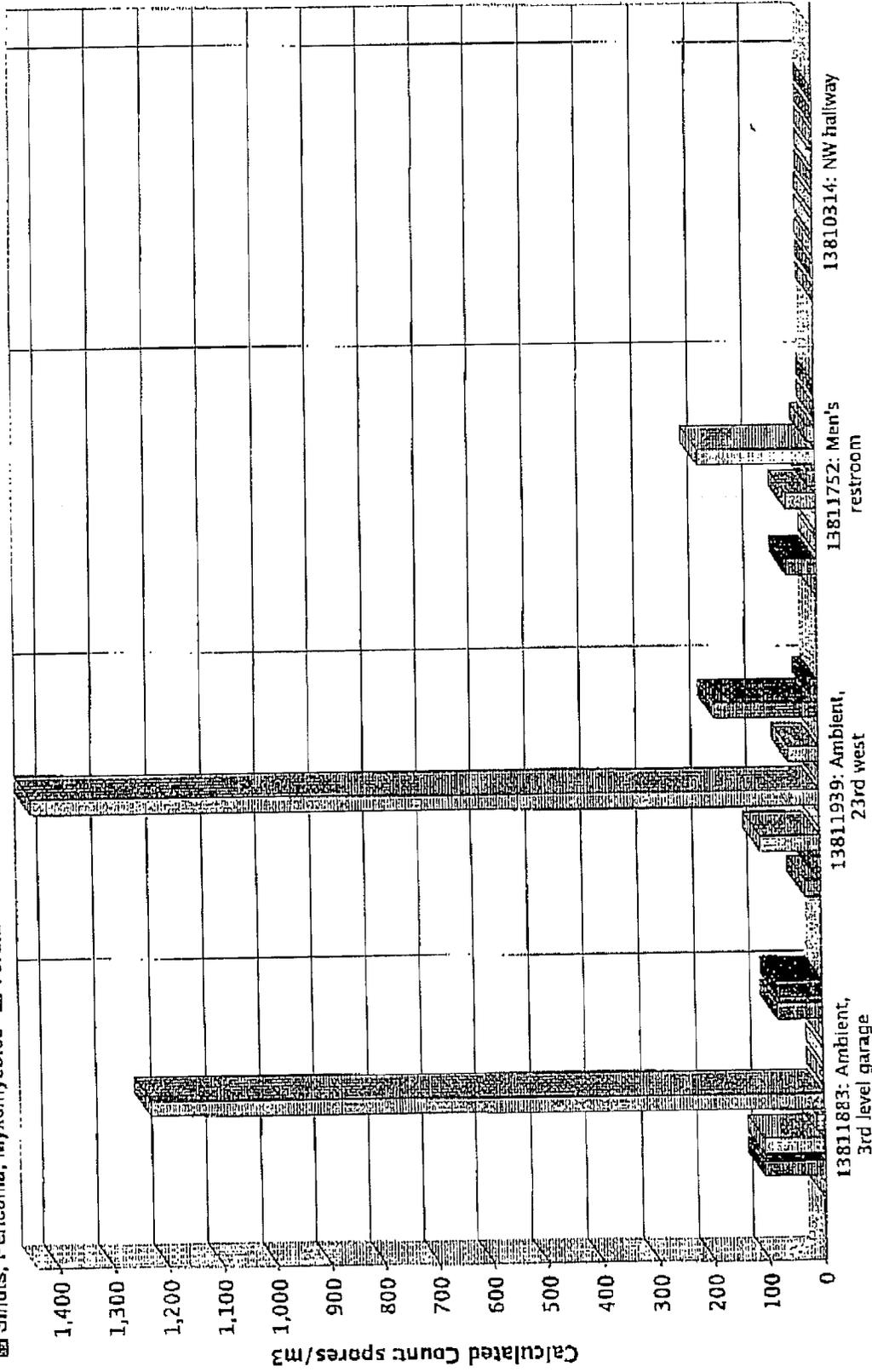
Interpretation of the data contained in this report is left to the client or the persons who conducted the field work. This report is provided for informational and comparative purposes only and should not be relied upon for any other purpose. "Typical outdoor data" are based on the results of the analysis of samples delivered to and analyzed by EMLab P&K and assumptions regarding the origins of those samples. Sampling techniques, contaminants infecting samples, unrepresentative samples and other similar or dissimilar factors may affect these results. In addition, EMLab P&K may not have received and tested a representative number of samples for every region or time period. EMLab P&K hereby disclaims any liability for any and all direct, indirect, punitive, incidental, special or consequential damages arising out of the use or interpretation of the data contained in, or any actions taken or omitted in reliance upon, this report.

EMLab P&K
1150 Bayhill Drive, Suite 100, San Bruno, CA 94066
(650) 829-5800 Fax (650) 829-5852 www.emlab.com

05-21-2008: 051908-02

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

- Alternaria Ascospores Basidiospores Beltrania Cladosporium Other brown Penicillium/Aspergillus types Rusts
- Smuts, Periconia, Myxomycetes Torula



Comments:

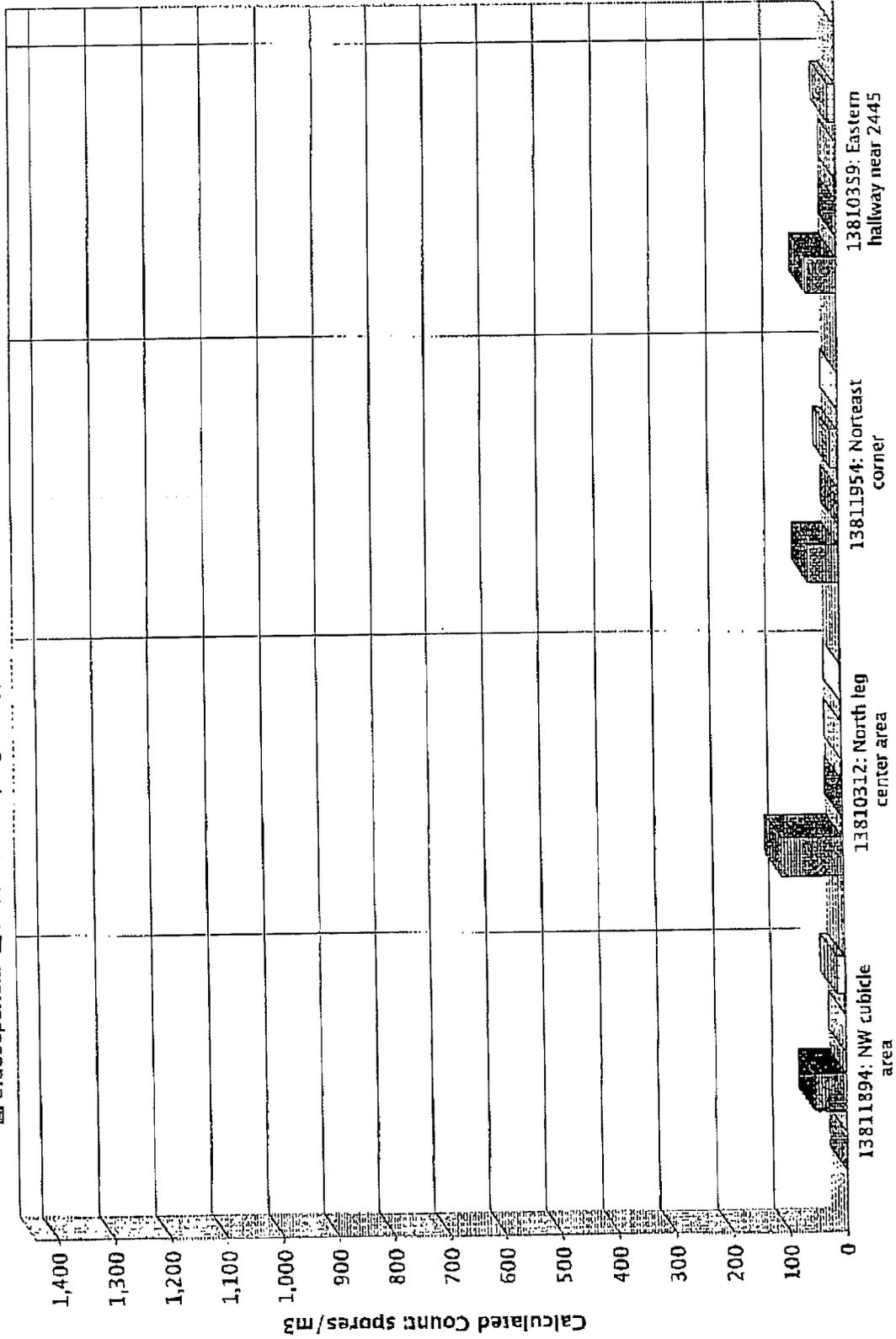
Note: Graphical output may understate the importance of certain "marker" genera.

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(650) 829-5800 Fax (650) 829-5852 www.emlab.com

05-21-2008: 051908-02

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Cladosporium Penicillium/Aspergillus types Rusts Smuts, Periconia, Myxomycetes



Comments:

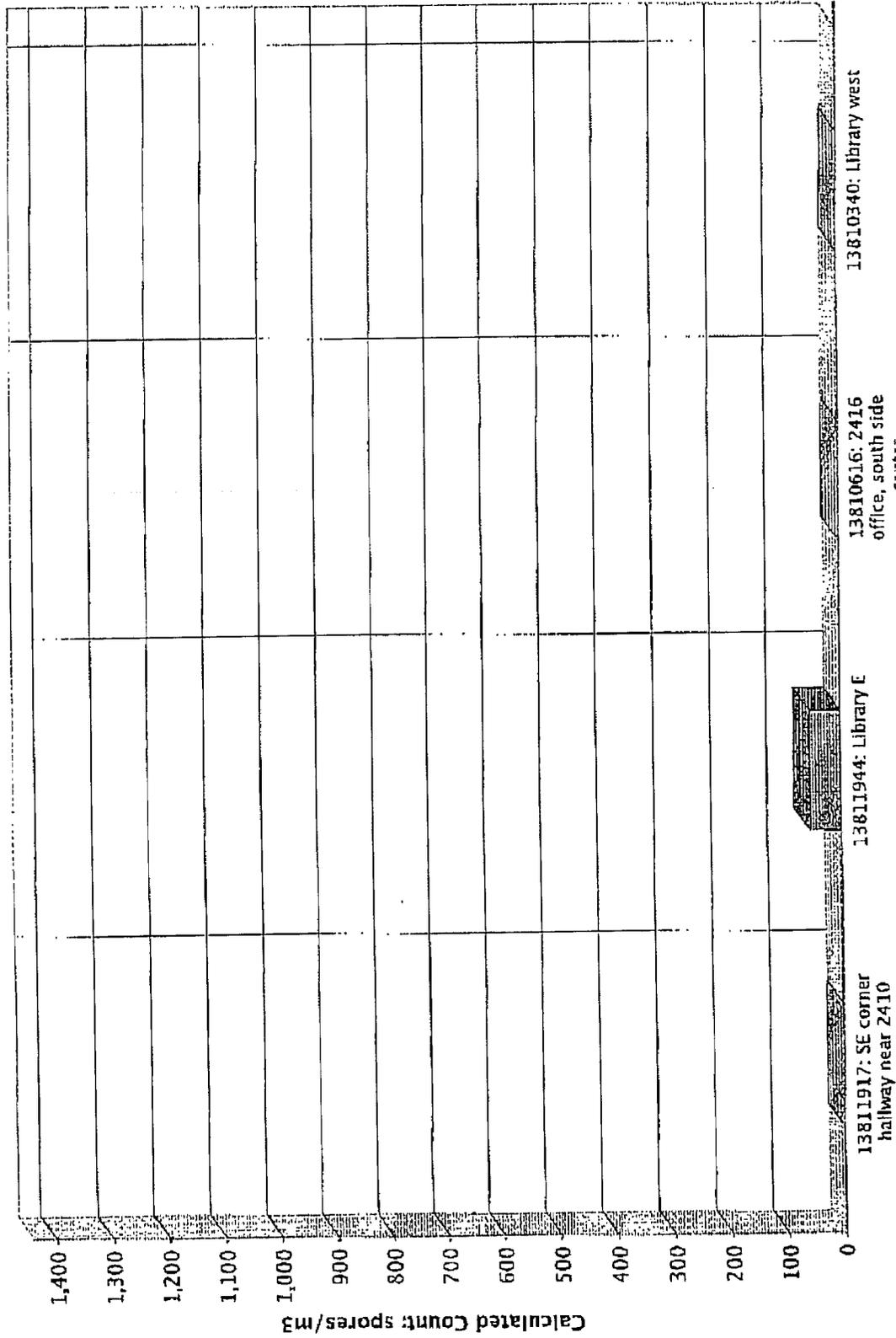
Note: Graphical output may understate the importance of certain "marker" genera.

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05-21-2008: 051908-02

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

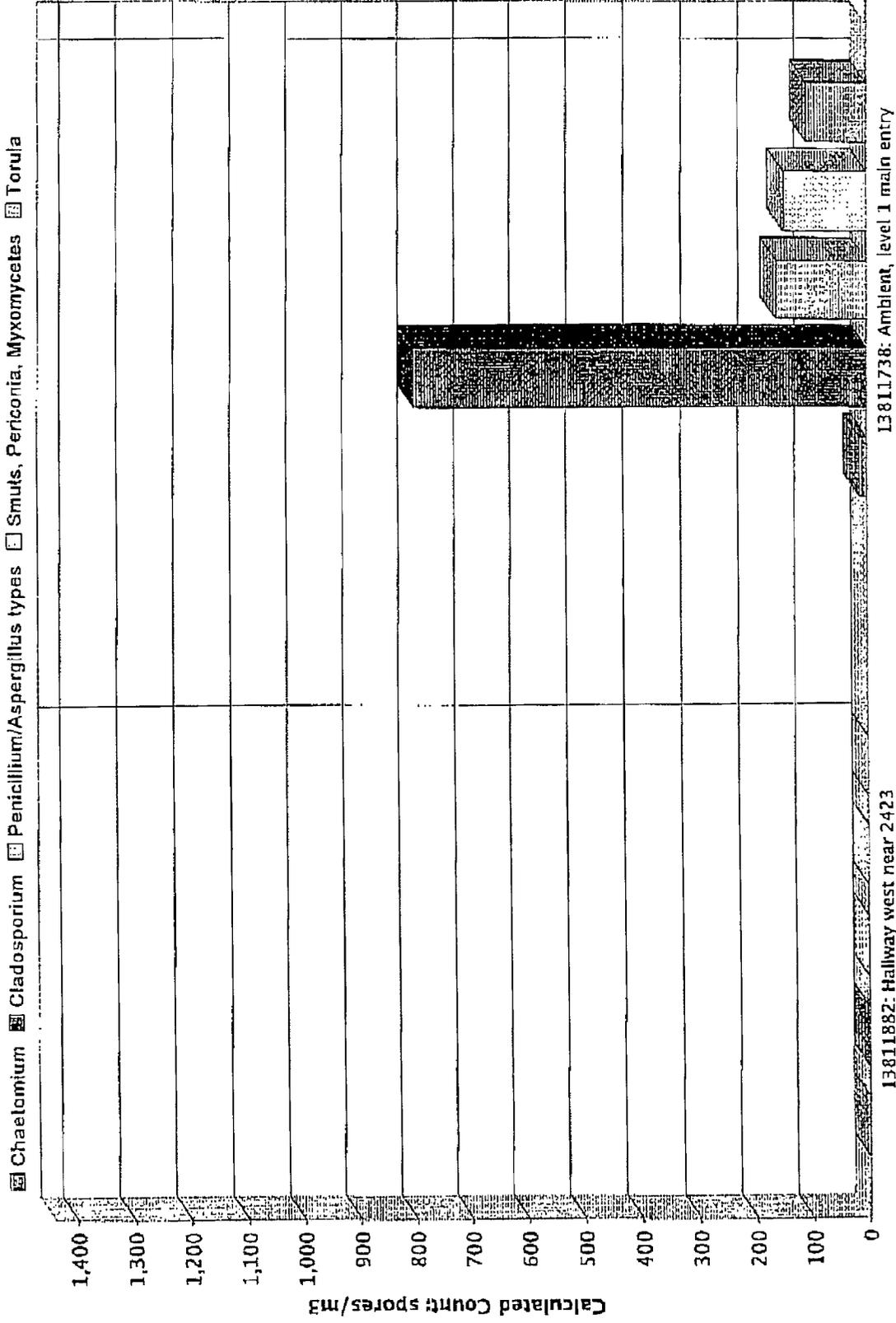
■ Cladosporium



Comments:

Note: Graphical output may understate the importance of certain "marker" genera.

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY



Comments:

Note: Graphical output may understate the importance of certain "marker" genera.

13811738: Ambient, level 1 main entry

13811882: Hallway west near 2423

MICROBIAL SPORE TRAP AIR SAMPLING RECORD



000424021

Page 1 of 2

BioMax Environmental
775 San Pablo Ave.
Pinole, CA 94564

www.biomaxenvironmental.com

Phone: (510) 724-3100

Fax: (510) 724-3145

biomaxenv@aol.com

Location: <u>BOE BIA 450 N Street (24th Floor)</u>	Client: <u>Dept Gen. Svc.</u>
Date: <u>5/19/08</u>	Project #: <u>051908-02</u>
Collected by: <u>M. A. Polkalis</u>	Laboratory: <u>EmLabs San Bruno CA</u>
Signature: <u>[Signature]</u>	Req. Turn Around: <u>24 HR</u>
	Analysis: <u>Fungal Particulate ID with Quantification</u>

Sample Number	Time	Location	Temp/Hum
A 13811883	1205	Ambient 3rd level Garage	77°/42%
B 13811737	1305	Ambient 23rd West	84°/32%
C 13811752	1320	Mari's Rest Room	89°/26%
D 13810314	1330	NW Hallway	91°/25%
E 13811817	1340	NW Cubicle Area	92°/26%
F 13810312	1348	North Log Center Area	93°/25%
G 13811959	1355	Northeast corner	97°/27%
H 13810359	1404	Easton Hallway near 2445	97°/24%
I 13811917	1410	SE Corner Hallway near 2410	95°/24%
J 13811944	1415	Library E	94°/25%
Total Sample Time (min): <u>5</u>	Flow Rate (l/min): <u>75 l/min</u>	Total Sample Volume (liters): <u>75L</u>	Ambient Conditions: <u>Clear + mild</u>
			Comments: <u>:</u>

Please sign this form below acknowledging sample receipt and return executed form with laboratory reports. Fax, send, e-mail results to BioMax Environmental at (510) 724-3145 biomaxenv@aol.com
Other Instructions: _____

Relinquished by: <u>[Signature]</u>	Received By: <u>Am Morrissey</u>
Method of Transportation: <u>Fed Ex</u>	Time/Date Received: <u>5-20-08 9am</u>
Time/Date Sent: <u>4:30 5/19/08</u>	

**EMLab P&K**

Report for:

Mr. Michael Polkaba
Biomax Environmental
775 San Pablo Ave.
Pinole, CA 94564

Regarding: Project: 051208-02; DGS, 450 N Street, Rms 2427 and 2428, 24th Floor
EML ID: 421540

Approved by:

Lab Manager
Magzoub Ismail

Dates of Analysis:
Spore trap analysis: 05-14-2008

Project SOPs: Spore trap analysis (I100000)

This coversheet is included with your report in order to comply with AIHA and ISO accreditation requirements.

For clarity, we report the number of significant digits as calculated; but, due to the nature of this type of biological data, the number of significant digits that is used for interpretation should generally be one or two. All samples were received in acceptable condition unless noted in the Report Comments portion in the body of the report. Due to the nature of the analyses performed, field blank corrections of results is not a standard practice. The results relate only to the items tested.

EMLab P&K ("the Company") shall have no liability to the client or the client's customer with respect to decisions or recommendations made, actions taken or courses of conduct implemented by either the client or the client's customer as a result of or based upon the Test Results. In no event shall the Company be liable to the client with respect to the Test Results except for the Company's own willful misconduct or gross negligence nor shall the Company be liable for incidental or consequential damages or lost profits or revenues to the fullest extent such liability may be disclaimed by law, even if the Company has been advised of the possibility of such damages, lost profits or lost revenues. In no event shall the Company's liability with respect to the Test Results exceed the amount paid to the Company by the client therefor.

Document Number: 200091 - Revision Number: 5

EMLab P&K

1010 N Central Avenue, Glendale, CA 91202
 (858) 569-5800 Fax (858) 569-5806 www.emlab.com

Client: Biomax Environmental
 C/O: Mr. Michael Polkabra
 Re: 051208-02; DGS, 450 N Street, Rms 2427 and
 2428, 24th Floor

Date of Sampling: 05-12-2008
 Date of Receipt: 05-13-2008
 Date of Report: 05-14-2008

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	13811763: 24th floor, area outside cont		13811904: 2427/2428 containment		13810533: Hallway outside cont South 24		13811890: NE corner area of 24th	
Comments (see below)	None		None		None		None	
Lab ID-Version†:	1853551-1		1853552-1		1853553-1		1853554-1	
	raw ct.	spores/m3	raw ct.	spores/m3	raw ct.	spores/m3	raw ct.	spores/m3
Alternaria								
Arthrinium								
Ascospores*								
Aureobasidium								
Basidiospores*								
Bipolaris/Drechslera group								
Botrytis								
Chaetomium								
Cladosporium					1	53		
Curvularia								
Epicoccum								
Fusarium								
Myrothecium								
Nigrospora								
Oidium								
Other brown								
Penicillium/Aspergillus types†								
Pithomyces								
Rusts*								
Smuts*, Periconia, Myxomycetes*					1	13		
Stachybotrys								
Stemphylium								
Torula								
Ulocladium								
Zygomycetes								
Background debris (1-4+)††	1+		1+		1+		1+	
Hyphal fragments/m3	< 13		13		< 13		< 13	
Pollen/m3	< 13		< 13		< 13		< 13	
Skin cells (1-4+)	< 1+		1+		< 1+		< 1+	
Sample volume (liters)	75		75		75		75	
TOTAL SPORE/m3		< 13		< 13		66		< 13

Comments:

* Most of these spore types are not seen with culturable methods (Andersen sampling), although some may appear as non-sporulating fungi. Most of the basidiospores are "mushroom" spores while the rusts and smuts are plant pathogens.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

†† Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.

The Limit of Detection is the product of a raw count of 1 and 100 divided by the percent read. The analytical sensitivity (counts/m3) is the product of the Limit of Detection and 1000 divided by the sample volume.

‡ A "Version" greater than 1 indicates amended data.

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Client: Biomax Environmental
C/O: Mr. Michael Polkabla
Re: 051208-02; DGS, 450 N Street, Rms 2427 and
2428, 24th Floor

Date of Sampling: 05-12-2008
Date of Receipt: 05-13-2008
Date of Report: 05-14-2008

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	13811702: 23rd floor N balcony		13810630: Ambient 4th floor garage Rf	
Comments (see below)	None		None	
Lab ID-Version†:	1853555-1		1853556-1	
	raw ct.	spores/m3	raw ct.	spores/m3
Alternaria	2	27	1	13
Arthrinium				
Ascospores*	1	13	1	13
Aureobasidium				
Basidiospores*	1	13	1	53
Bipolaris/Drechslera group				
Botrytis				
Chaetomium	1	13	2	27
Cladosporium	6	320	6	320
Curvularia	1	13		
Epicoccum				
Fusarium				
Myrothecium				
Nigrospora				
Oidium	4	53	7	93
Other brown	1	13		
Other colorless				
Penicillium/Aspergillus types†			5	267
Pithomyces				
Rusts*				
Smuts*, Periconia, Myxomycetes*	22	293	24	320
Stachybotrys	1	13		
Stemphylium				
Torula				
Ulocladium				
Zygomycetes				
Background debris (1-4+)††	3+		3+	
Hyphal fragments/m3	27		120	
Pollen/m3	240		693	
Skin cells (1-4+)	< 1+		< 1+	
Sample volume (liters)	75		75	
TOTAL SPORE/m3		771		1,106

Comments:

* Most of these spore types are not seen with culturable methods (Andersen sampling), although some may appear as non-sporulating fungi. Most of the basidiospores are "mushroom" spores while the rusts and smuts are plant pathogens.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

†† Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting viability for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.

The Limit of Detection is the product of a raw count of 1 and 100 divided by the percent read. The analytical sensitivity (counts/m3) is the product of the Limit of Detection and 1000 divided by the sample volume.

‡ A "Version" greater than 1 indicates amended data.

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Client: Biomax Environmental
C/O: Mr. Michael Polkabila
Re: 051208-02; DGS, 450 N Street, Rms 2427 and
2428, 24th Floor

Date of Sampling: 05-12-2008
Date of Receipt: 05-13-2008
Date of Report: 05-14-2008

MoldRANGE™: Extended Outdoor Comparison**Outdoor Location: 13811702, 23rd floor N balcony**

Fungi Identified	Outdoor data	Typical Outdoor Data by Date†				Typical Outdoor Data by Location‡			
		Month: May				State: CA			
	spores/m3	low	med	high	freq %	low	med	high	freq %
Generally able to grow indoors*									
Alternaria	27	7	33	320	62	7	27	230	60
Bipolaris/Drechslera group	-	7	13	120	16	7	13	120	14
Chaetomium	13	7	13	100	15	7	13	110	19
Cladosporium	320	53	530	6,900	96	53	640	6,400	98
Curvularia	13	7	13	360	8	7	13	210	7
Nigrospora	-	7	13	120	7	7	13	170	8
Other brown	13	7	13	80	36	7	13	80	37
Penicillium/Aspergillus types	-	27	160	1,700	80	40	210	2,500	88
Stachybotrys	13	7	13	230	4	7	13	300	5
Torula	-	7	13	170	15	7	13	150	13
Seldom found growing indoors**									
Ascospores	13	13	160	5,300	81	13	110	1,800	72
Basidiospores	13	13	270	6,900	93	13	240	6,700	94
Oidium	53	7	22	250	27	7	13	190	20
Rusts	-	7	25	330	29	7	13	270	29
Smuts, Periconia, Myxomycetes	293	7	53	1,000	76	8	40	470	71
TOTAL SPORES/M3	771								

† The Typical Outdoor Data by Date represents the typical outdoor spore levels across North America for the month indicated. The last column represents the frequency of occurrence. The low, medium, and high values represent the 2.5, 50, and 97.5 percentile values of the spore type when it is detected. For example, if the frequency of occurrence is 63% and the low value is 53, it would mean that the given spore type is detected 63% of the time and, when detected, 2.5% of the time it is present in levels above the detection limit and below 53 spores/m³. These values are updated periodically, and if enough data is not available to make a statistically meaningful assessment, it is indicated with a dash.

‡ The Typical Outdoor Data by Location represents the typical outdoor spore levels for the region indicated for the entire year. As with the Typical Outdoor Data by Date, the four columns represent the frequency of occurrence and the typical low, medium, and high concentration values for the spore type indicated. These values are updated periodically, and if enough data is not available to make a statistically meaningful assessment, it is indicated with a dash.

*The spores in this category are generally capable of growing on wet building materials in addition to growing outdoors. Building related growth is dependent upon the fungal type, moisture level, type of material, and other factors. *Cladosporium* is one of the predominant spore types worldwide and is frequently present in high numbers. *Penicillium/Aspergillus* species colonize both outdoor and indoor wet surfaces rapidly and are very easily dispersed. Other genera are usually present in lesser numbers.

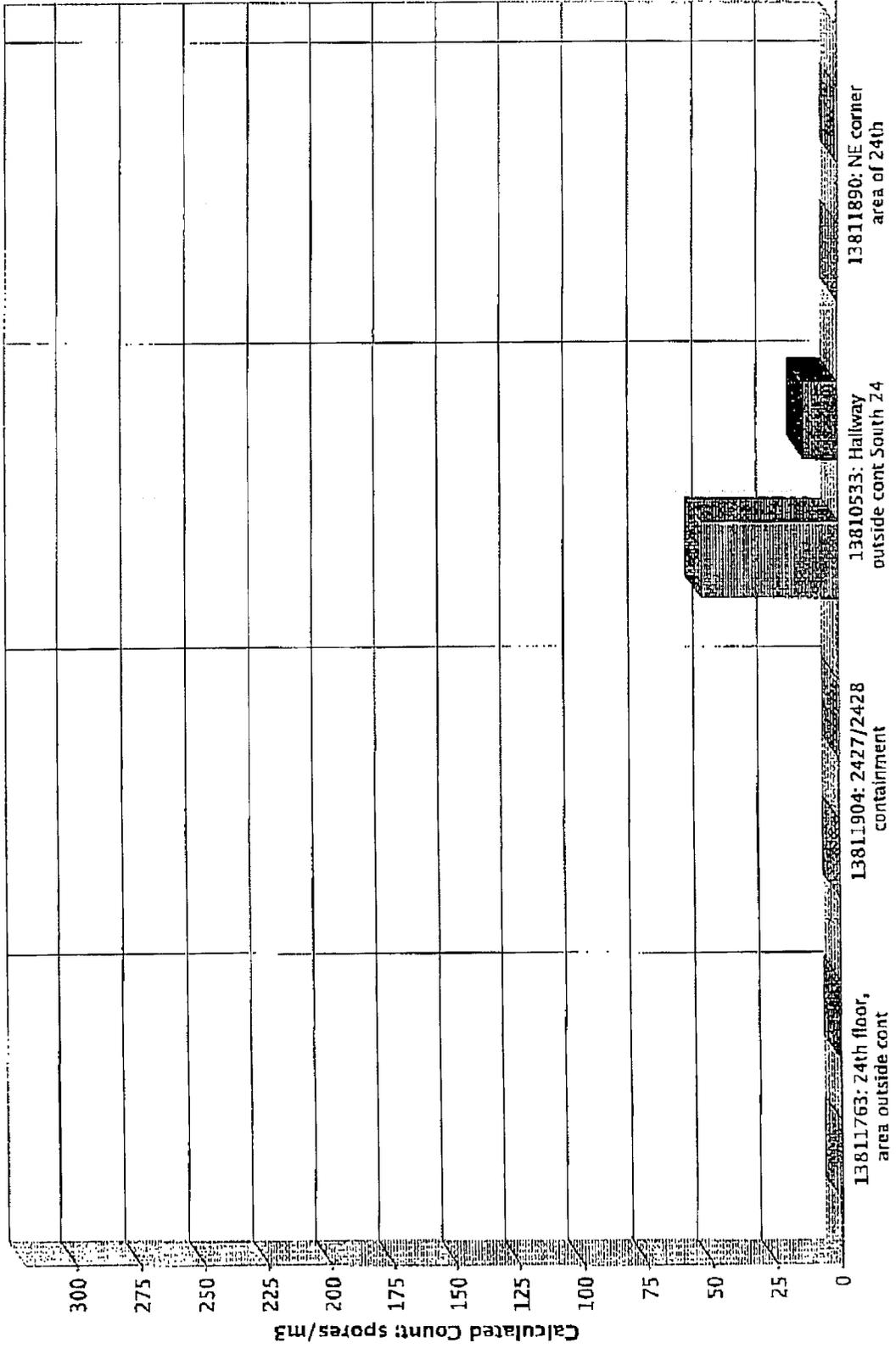
**These fungi are generally not found growing on wet building materials. For example, the rusts and smuts are obligate plant pathogens. However, in each group there are notable exceptions. For example, agents of wood decay are members of the basidiomycetes and high counts of a single morphological type of basidiospore on an inside sample should be considered significant.

Interpretation of the data contained in this report is left to the client or the persons who conducted the field work. This report is provided for informational and comparative purposes only and should not be relied upon for any other purpose. "Typical outdoor data" are based on the results of the analysis of samples delivered to and analyzed by EMLab P&K and assumptions regarding the origins of those samples. Sampling techniques, contaminants infecting samples, unrepresentative samples and other similar or dissimilar factors may affect these results. In addition, EMLab P&K may not have received and tested a representative number of samples for every region or time period. EMLab P&K hereby disclaims any liability for any and all direct, indirect, punitive, incidental, special or consequential damages arising out of the use or interpretation of the data contained in, or any actions taken or omitted in reliance upon, this report.

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SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Cladosporium
 Smuts, Periconia, Myxomycetes

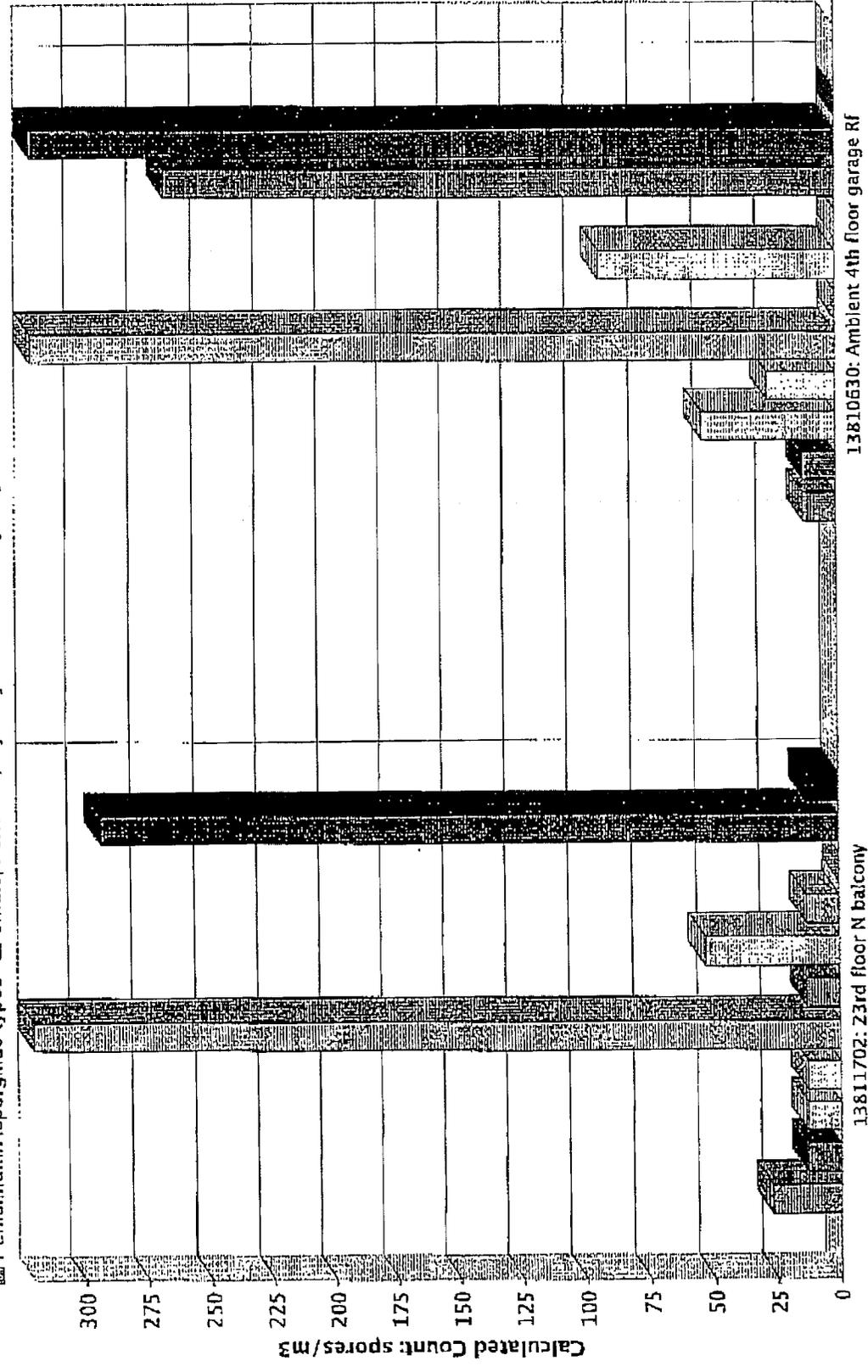


Comments:

Note: Graphical output may understate the importance of certain "marker" genera.

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

- Alternaria
- Ascospores
- Basidiospores
- Chaetomium
- Cladosporium
- Curvularia
- Oidium
- Other brown
- Penicillium/Aspergillus types
- Smuts, Periconia, Myxomycetes
- Stachybotrys



13811702: 23rd floor N balcony

13810630: Ambient 4th floor garage Rf

13811702: 23rd floor N balcony

Comments:
 Note: Graphical output may understate the importance of certain "marker" genera.

MICROBIAL SPORE TRAP AIR SAMPLING RECORD

Page 1 of 1

BioMax Environmental
775 San Pablo Ave.
Pinole, CA 94564

www.biomaxenvironmental.com

Phone: (510) 724-3100
Fax: (510) 724-3145
biomaxenv@aol.com

Location: 450 N Street Rms 2427 + 2428 (24th Floor)	Client: DGS
Date: 5/12/08	Project #: 05/208-02
Collected by: M. A. Polka Signature: <i>M. A. Polka</i>	Laboratory: Req. Turn Around: 24HR Analysis: <u>Fungal</u> <u>Particulate ID</u> <u>with Quantification.</u>

Sample Number	Time	Location/Notes	Temp/Hum
13811763	1330	24th Floor (Area outside cont.)	82° / 26%
13811904	1338	2427 / 2428 Containment	85° / 26%
13810533	1345	Hallway outside cont South 24	86° / 27%
13811890	1352	NE Corner Area of 24th	88° / 29%
13811702	1415	23rd Floor N. Balcony	71° / 25%
13810630	1505	Ambient 4th Floor Garage Rt.	76° / 29%
Total Sample Time (min):	Flow Rate (l/min):	Total Sample Volume (liters):	Ambient Conditions:
- 5	15	75	m. bl windy 10-20 mph
			Comments:

Please sign this form below acknowledging sample receipt and return executed form with laboratory reports. Fax, send, e-mail results to BioMax Environmental at (510) 724-3145 biomaxenv@aol.com
Other Instructions: _____

Relinquished by: <i>M. A. Polka</i>	Received By: <i>Wagner</i>
Method of Transportation: <i>FedEx</i>	Time/Date Received: <i>5/13/08 9:00</i>
Time/Date Sent: <i>4:30 5/12/08</i>	

