
BioMax Environmental

Environmental Consulting and Industrial Hygiene Services

May 26th, 2008

Mr. Doug Button
Deputy Director
Real Estate Services Division
707 Third Street - 8th Floor
West Sacramento, CA 95605

**Microbial Assessment and
Mitigation Procedures for 20th Floor Mitigation – Break Room Area
Department of General Services Board of Equalization Building
450 N. Street
Sacramento, California**

Dear Mr. Button,

BioMax Environmental, LLC (BioMax) is pleased to provide the Department of General Services (DGS) with this letter summary report detailing BioMax's findings and recommendations pertaining to our inspection and microbial sampling assessment services provided within the moisture and mold impacted areas associated with the 20th Level Break Room of your 450 N Street Building (subject building) located in Sacramento, California. BioMax understands that these microbial inspection and sampling assessment services were contracted with BioMax in an effort to evaluate the recently discovered visible moisture damage and potential microbial growth identified within the noted Break Room (2014) located on the 20th floor of the subject building. According to DGS personnel, such areas were identified during routine daily activities and operations by the building tenant (BOE). Following such discovery, BioMax understands that the break room was immediately vacated (and precluded from use) at BOE's option pending further investigative activity.

Hence, these microbial inspection and assessment services are intended to obtain analytical sampling data and physical inspection information pertaining to the current environmental conditions present within the affected interior area and impacted materials identified. Site access was provided on Tuesday, May 13th, 2008 by DGS representatives. On this day, Mr. Michael A. Polkaba, CIH, REA of BioMax performed a site inspection and sampling assessment within and adjacent to the areas of concern identified by DGS representatives. Based on current information provided and our visual observations gathered at this time, BioMax collected a series of airborne and surface microbial samples within and surrounding the areas of concern and representative affected materials so as to evaluate and assess the current environmental microbial conditions within and surrounding the impacted areas at this time.

SITE OBSERVATIONS

On-site inspection and sampling assessment activities were performed by Mr. Michael A. Polkaba, CIH, REA, of BioMax in accordance with currently recognized microbial assessment and sampling guideline procedures. Mr. Polkaba has been certified in the Comprehensive Practice of Industrial Hygiene by the American Board of Industrial Hygiene and holds the right to the designation "Certified Industrial Hygienist" (CIH) under certification number CP 7104. Mr. Polkaba is also certified by the California Environmental Protection Agency (Cal/EPA) as a Class I Registered Environmental Assessor (REA) under Cal/EPA certification number 05011. A summary of significant notations and observations gathered during BioMax's site inspection and assessment of the subject areas are compiled as follows:

1. At the time of our preliminary site inspection performed on May 13th, 2008 interior environmental conditions within the subject area consisted of a temperature of 73 degrees F with relative humidity of 27 %. Ambient outdoor conditions both prior to and following our interior assessment consisted of mild sunny conditions with predominant winds noted at approximately 5-10 knots from the northwesterly direction. Outdoor temperatures ranged between 76 to 77 degrees and relative humidity range of 35 to 38 %, respectively.
2. Site observations noted within the subject areas are as follows:

Break Room (# 2014) – At the time of our assessment, the sink cabinet located along the northern wall-side of the break room area indicated evidence of historical moisture damages as evidenced by floor cabinet base and floor tile cracking and delamination. Upon localized removal (for inspection) of cabinet laminant, visual indications of “spotty” visible mold growth was observed present beneath the white venire material and within the particle board underlayment. Based on such findings, BioMax collected a series of bulk material and surface samples from the impacted materials. Following sampling, BioMax re-taped the access cabinet door pending review of the findings gathered as part of this assessment. For comparative purposes, supplemental surface samples were also collected from non-impacted material surfaces within the break room as well as from a representative surface located outside the break room within the BOE occupied work area.

Cubicle #078 (Nikkole Thorn) - At the time of our assessment, BioMax inspected the cubicle adjacent to the noted break room occupied by Ms. Nikkole Thorn. No visual evidence of mold-like staining was observed present on the shared real wall of Ms. Thorn's cubicle area at the time inspected. Utilization of a hand-held moisture meter also indicated normal moisture content present within the cubicle divider material at the nearest location to the impacted area at the time of our assessment. An air sample was also collected from this area at this time as noted below.

MAP Scanning Room (# 2013) – At the time of our assessment, BioMax inspected the MAP Scanning Room with a shared wall located to the west and adjacent to the noted break room.

No visual evidence of mold-like staining was observed present on the shared wall. Utilization of a hand-held moisture meter also indicated normal moisture content present within the wallboard material at the nearest approximate location to the impacted area at the time of our assessment. A single air sample was similarly collected from this adjacent area for comparative purposes.

3. As noted above, BioMax collected a series of airborne, bulk and surface samples from representative impacted building materials surfaces and areas located within and surrounding the visibly affected interior materials noted above. Utilization of hand-held moisture detection equipment indicated elevated localized moisture content within the affected sink cabinet wall materials surveyed within the break room at the time of our assessment.
4. In an effort to further evaluate the potential for exposure to airborne spores and/or spore deposition resultant from airborne transmission from the noted areas, BioMax also collected a series of SporeTrap and surface samples within the adjacent immediate work area as noted within Table 1 below. Such supplemental samples were collected from accessible surfaces during normal daily building ventilation conditions and routine employee operations and activities so as to model representative conditions within the working environment.
5. A series of digital images were also collected during BioMax's inspection and sampling assessment activities. Images are attached to this summary report for further reference, as necessary. A detailed site map sketch indicating the extent of visibly affected areas noted at the time of this assessment and relative surface sampling locations may also be provided for further reference upon request.

SAMPLING PROCEDURES

On-site inspection and sampling assessment activities were conducted by Mr. Michael A. Polkabl, CIH, REA, of BioMax Environmental on February 5th, 2008. All sampling equipment, supplies, calibration materials, and collection media were provided by BioMax as part of the performance of this scope of work. Sample collection procedures and methods were performed using aseptic sampling methods following techniques prescribed by the contracted analytical laboratory.

SporeTrap Airborne Sampling:

The collection of airborne Spore Trap microbial samples was achieved using Zefon Air-O-Cell sampling cassette collection devices placed in each of the areas identified in Table 1. A total of seven (7) airborne Spore Trap samples were collected within and outside the areas of concern at a height of approximately four feet above ground level using a tripod mounted Quick Take 15 air sampling pump manufactured by SKC International. Samples were collected at a calibrated flow rate of 15 liters per minute for a total of five minutes per sample. Resultant total sample volumes, therefore, corresponded to 75 liters collected for each sample. Field calibration of the

SKC air sampling pump was conducted and recorded prior to and following sampling activities using a Bios Drycal primary standard flow meter and field rotometer. All spore trap air sampling and analytical procedures were performed in accordance with prescribed manufacturer guidelines as well as applicable professional certified industrial hygiene indoor air quality microbial investigation procedures and certified industrial hygiene practices.

Additional exterior samples were also similarly collected and analyzed during the collection of interior samples in an effort to evaluate and quantify typical background microbial taxa (types), rank order, and corresponding airborne spore levels present during the time of this assessment. Efforts were made in the collection of airborne samples to capture such samples during representative building occupancy conditions and activities so as to closely approximate normal air handling system ventilation conditions within each of the subject areas located within and surrounding the evaluated areas of concern. Sampling collection activities performed during this assessment included the collection of identifiable airborne microbial contaminants within the representative areas noted in Table 1 below:

Table 1. Airborne Spore Trap Sampling Locations:

Air Sample Number	Spore Trap Air Sampling Location
13811704	20 th Floor Break Room (#2014)
13811714	Hallway Area Southwest of Break Room
13810632	Cubicle Area 078 (Nikkole Thorn)
13811941	Break Room 2014 (w/Cabinet Doors "Open")
13811760	Room 1013 MAP Scanning Room
13811685	Ambient Mezzanine Floor SW Balcony
13810584	Ambient outside location (Main Entry Level)

At the conclusion of sampling activities, preparation and shipping of the collected airborne samples were accomplished in accordance with standard industrial hygiene chain of custody (COC) documentation procedures and quality assurance/quality control practices. Once collected, labeled, and recorded, all samples were double sealed within airtight plastic Ziploc shipping containers and transported via Federal Express Priority Mail to Environmental Microbial Laboratories (EMLabs) in San Bruno, California. EMLabs holds current applicable analytical accreditation and specializes in microbial analytical procedures. Sampling and chain of custody records are provided as an attachment to this letter report for further reference.

Bulk Material and BioTape Surface Sampling:

During our site inspection and sampling assessment activities, representative bulk material and surface material samples were collected from interior areas and materials of concern noted within in Table 2 below. All surface samples were collected using "same-lot" BioTape collection media prepared and supplied by SKC International in accordance with manufacturers sampling guidelines as well as applicable professional certified industrial hygiene microbial sampling practices. Bulk material samples were similarly collected utilizing aseptic sample collection technique in accordance with standard microbial sampling practices. Disposable gloves utilized during sample collection and changed between each sample.

Written sampling procedural guidance material prepared by the analytical laboratory and/or sample media manufacturer may also be provided upon request. A summary of bulk material and surface material sampling locations are provided in Table 2. Specific sample locations may also be referenced within the digital image attachment and referenced site map diagram, as necessary.

Table 2. Bulk Material and BioTape Surface Sample Locations:

Sample Number	Material Sampling Location
S01	Break Room cabinet base surface right side
S02	Break Room cabinet base surface left side
S03	Break Room metal cabinet top surface
S04	Break Room wall mounted cabinet top surface
S05	Cubicle divider top surface within adjacent occupied work area
B01	Break Room cabinet base venire and underlayment at center
B02	Break Room cabinet base venire and underlayment at rear

Following sample collection, bulk material and surface samples were subsequently labeled and placed within individual plastic Ziploc storage bags for transportation via Federal Express Priority Mail to the analytical laboratory noted below. Preparation and shipping of the collected samples were accomplished in accordance with standard industrial hygiene chain of custody (COC) documentation procedures and quality assurance/quality control QA/QC practices. Once collected, labeled, and recorded, the samples were double sealed within airtight plastic Ziploc bag containers and similarly transported via Federal Express Priority Mail to Environmental Microbial Laboratories (EMLabs) of San Bruno, California. Sampling and chain of custody records are provided as an attachment to this letter report for further reference.

ANALYTICAL FINDINGS AND CONCLUSIONS

Airborne Spore Trap Sample Findings:

Laboratory analytical methods for the identification and enumeration of microbial (mold) taxa contaminants were conducted in accordance with prescribed analytical procedures and quality control/assurance measures. Original laboratory results including the enumeration of recognizable microbial spore types are also attached to this letter report for further detail. Analytical comments provided by the microbial laboratory regarding relative background debris and particulate levels are noted as a semi-quantitative assessment based on analyst interpretation and historical regional data. A summary of airborne Spore Trap microbial (mold) and particulate findings pertaining to each of the subject areas are presented in Table 3 below:

Table 3. Summary of Airborne Microbial Findings

Location Desc.	Total Mold Spores (CIS/m ³)	Background Debris (Scale of 1-4)	Skin Cell Fragments (Scale of 1-4)
20 th Floor Break Room (#2014)	66	2+	1+
Hallway Area Southwest of Break Room	93	1+	1+
Cubicle Area 078 (Nikkole Thorn)	13	1+	1+
Break Room 2014 (w/Cabinet Doors "Open")	119	1+	1+
Room 1013 MAP Scanning Room	13	1+	1+
Ambient Mezzanine Floor SW Balcony	146	1+	1+
Ambient outside location (Main Entry Level)	466	1+	<1+

The analytical findings presented in Table 3 indicate the presence of significantly lower concentrations of microbial (mold) spores measured within each of the interior samples collected within and surrounding the subject areas when compared to the levels currently measured from the samples collected from the corresponding ambient outside environment. Analytical findings also indicate similar fungal taxa distribution (mold types) and rank order (predominant taxa) of molds identified within the mitigated areas as well as the worker occupied area samples.

Analytical findings also indicated the absence of significantly elevated airborne levels of fungal taxa uniquely present within these noted adjacent areas currently occupied and accessed by BOE staff. Particularly worthy of note, was the absence of elevated levels of hydrophilic (moisture loving) mold taxa such as *Penicillium*, *Aspergillus*, and *Stachybotrys* within any of the air samples collected from within or adjacent to the noted break room areas. Hence, all analytical airborne findings gathered within the adjacent occupied area performed as part of this assessment currently fall within generally acceptable ranges and parameters. Hence, BioMax believes that the current airborne sample data evaluated within the adjacent occupied areas (areas outside the Break Room) provide no significant evidence of airborne transmission, distribution, and/or elevated mold contamination at the present time. Therefore, BioMax believes that no supplemental area specific cleaning measures are warranted within areas outside of the affected break room impacted areas at this time.

Although there are currently no regulatory standards or limits pertaining to allowable airborne fungal concentrations (for any mold taxa) present in indoor environments, there is a general consensus among indoor air quality experts that microbial contamination found within "typical healthy" working and living spaces are generally similar in kind and present at levels which are below those found in the corresponding native outside environment. BioMax believes that the absence of elevated moisture, absence of significant visible residual mold, and relatively fewer total airborne mold levels with typical taxa and rank order distribution within are consistent with these generally acceptable conditions. BioMax, therefore, believes that these findings provide reasonable evidence indicating that current airborne microbial conditions present within the break room and adjacent working areas are consistent with normal representative levels.

Therefore, BioMax believes that verification of such current airborne microbial conditions under the conditions sampled may be considered acceptable for normal BOE use of the break room area with precluded access into the impacted cabinet compartment material at this time. Such limited access into the affected cabinet area should be maintained (through barrier tape) until appropriate mitigative measures have been performed and successfully completed thorough verified post mitigation "clearance" sampling as recommended below.

Airborne Particulate Debris Findings:

Analytical findings pertaining to the relatively low levels of airborne particulates debris identified within the collected air samples within and surrounding the previously impacted areas also provide reasonable evidence indicating that current environmental conditions present within the break room and surrounding areas are typical and unremarkable at this time.

Bulk Material and Surface Sample Findings:

Laboratory analytical methods for the identification and enumeration of microbial taxa were conducted in accordance with prescribed analytical procedures and quality control/assurance measures. Laboratory analytical methods for the identification and enumeration of microbial fungal contaminants within the collected surface material samples were achieved through direct microscopic analysis using bright field microscopy.

Original laboratory results including the identification of recognizable microbial taxa are provided as an attachment to this letter report for further reference. Sampling and chain of custody records are provided as an attachment to this report for further reference. A summary of analytical findings pertaining to the collected bulk material and surface samples are presented in Table 4 below:

Table 4. Summary of Bulk Material and Surface Findings:

Sample Number	Sample Material and Location	Mold Genera Identified Present
S01	Break Room cabinet base surface right side	Cladosporium and Rust type spores detected. 2 Counts.
S02	Break Room cabinet base surface left side	Chaetomium spores and Smuts detected. 12 Counts
S03	Break Room metal cabinet top surface	No Mold Spores Detected
S04	Break Room wall mounted cabinet top surface	Smuts, Periconia, Myxomycetes-type spores detected. 1 Count
S05	Cubicle divider top surface within adjacent occupied work area	No Mold Spores Detected
B01	Break Room cabinet base venire and underlayment at center	Eight (8) raw count of Penicillium/Aspergillus type spores identified on surface.
B02	Break Room cabinet base venire and underlayment at rear	Forty one Eight (41) raw count of Penicillium/Aspergillus type spores identified on surface.

Noted relative levels should be used for comparative purposes only and are not intended to establish "safe" or "acceptable" indoor levels/conditions.

Analytical findings as presented in Table 4 above clearly indicated the presence of unique microbial fragments (spores) present in each of the majority of the materials sampled where staining was noted. The identified hydrophilic (moisture loving) mold taxa, such as Penicillium/Aspergillus and Chaetomium, identified within the visibly "stained" bulk materials sampled, represent what BioMax believes to be likely indicative of prior historical mold growth and likely not resultant directly from any singular recent water release incident.

Although there are currently no regulatory standards or limits pertaining to allowable surface fungal concentrations (for any mold taxa) present on interior working environment surfaces,

there is a general consensus among indoor air quality and microbial experts that significant visible microbial contamination found within occupied space building materials should be treated, removed, and/or otherwise minimized wherever practicable. Hence, BioMax believes that the findings detailed in this report warrant the implementation of the recommended precautions, continued area controls, and the performance of mitigative measures pertaining to the areas of identified visible microbial contamination.

Analytical findings pertaining to the surface sampling data gathered within the adjacent occupied area performed as part of this assessment currently fall within generally acceptable ranges and parameters. Hence, BioMax believes that the current surface sample data evaluated within the adjacent occupied areas (areas outside the Break Room) provide no significant evidence of airborne transmission, distribution, or mold spore deposition onto adjacent area surfaces at present. Therefore, BioMax believes that no area specific particulate removal and/or cleaning measures are warranted within areas outside of the affected break room impacted areas at this time.

RECOMMENDATIONS

Based on our preliminary observations within the subject areas and review of current analytical findings available at this time, BioMax recommends that the following corrective measures and mitigative actions be considered as follows:

1. Due to the confirmed findings of elevated microbial contamination present within the sampled building materials and Break Room areas noted in this report, BioMax recommends that additional deconstructive inspection and appropriate mitigation the affected interior structures, walls, and wall cavities within the subject areas be performed as noted below. The purpose of these activities should be to adequately assess and evaluate the full extent of all moisture intrusion and microbial damages within each of the noted areas under appropriate microbial mitigative protective containment systems.
2. In performing such mitigative measures, BioMax recommends that a qualified and experienced microbial abatement contractor be selected to erect critical containment barriers at the hallway and entrance Break Room 2014 so as to perform prescribed microbial mitigative measures within the affected interior areas and structures noted. The selected contractor must be specifically trained in the field of microbial abatement techniques and methods as well as maintain demonstrated proficiency in the establishment and use of appropriate barriers, personal protective equipment, abatement techniques and methods in the removal and decontamination of microbial affected and impacted materials.
3. Due to the current occupancy and client use within the areas adjacent to the affected break room, as a precautionary measure, BioMax recommends that the tenant continues to be precluded from access into the break room until the areas have been appropriately mitigated and verified as acceptable for reuse. The mitigation contractor should be directed to install a fully enclosed negative pressure environmental containment barriers encompassing the

entirety of the break room area and affected materials during removal, inspection, and treatment. These containment systems shall be designed for the purposes of containing and controlling possible fugitive emissions of airborne fungal spore contaminants during all forthcoming deconstruction, inspection, and mitigative activities within the premises. All critical containment systems shall be constructed of plastic and/or otherwise airtight materials so as to create a negative pressure system within the noted areas of concern. Due to physical constraints, all negative air pressure shall be maintained within the critical areas with the use of a High Efficiency Particulate Aerosol (HEPA) filtered "negative air machine" vented to the outside workspace environment. An adequate supply of filtered intake air shall also be established to allow an adequate supply of "clean" filtered make-up air into the critical containment. Wherever possible, clear translucent plastic observation windows shall be placed on the critical containment barrier within direct sight of the affected areas for the purposes of inspection during the performance of prescribed mitigative measures. BioMax is prepared to provide your selected contractor with additional and ongoing detail pertaining to the establishment maintenance, and specific locations of critical containment barriers, as necessary. Once, containment parameters have been established, the site contractor shall maintain an "as built" record of exact containment locations and materials for further review and reference.

4. As an additional precautionary measure (and at the request of BOE's environmental consultant), HEPA filtered air scrubber units will be operated in the hallway outside the containment area for the duration of mitigative activities.
5. A series of similar plastic and/or otherwise impermeable zippered entry chambers shall also be erected at the entrance of the containment systems for the purpose of establishing worker entrance/exit and clean personal protective equipment donning and decontamination area. HEPA filtered vacuum equipment capable of the effective removal of particulate contaminants from tools and personal protective equipment shall be placed within each of the zippered chambers closest to the working area. During such measures, appropriate signage and warnings must be posted on the exterior of containment entrances to preclude uninformed access from unauthorized personnel. Data logging monitoring equipment employed to record pressure differentials on a 24-hour basis shall be used for the duration of functional barrier use.
6. Upon establishment of critical containment barriers, BioMax recommends that the selected microbial abatement contractor also places and maintains appropriate HEPA filtered air-scrubbing and/or dehumidification units within the affected areas, as necessary. All Heating Ventilation and Air Conditioning (HVAC) supply vents and ceiling or wall mounted recessed lighting/ fan penetrations within the containment systems shall be deactivated and covered within similar plastic barrier systems. All appropriate wall and ceiling penetrations present within the containment systems shall also be sealed and/or otherwise rendered airtight and inoperable so as to minimize unfiltered particulate intrusion into and out of the established containment systems. It is specifically recommended that the ceiling tile level materials be critically sealed from the working areas within each of the noted containment rooms so as to preclude fugitive emissions from exiting the noted containments. Any smoke detectors

- and/or fire suppression systems shall NOT be covered nor rendered inoperable within the subject building unless authorized to do so under the direction and supervision of personnel.
7. Workers engaged in mold remediation/mitigation activities must be adequately trained and equipped with properly selected personal protective equipment (PPE) including, at minimum, hooded Tyvek coveralls, air purifying full face respirators with N100 minimum HEPA filter rating or similar PAPR systems, nitrile or latex gloves, chemical resistant boots or boot covers, with taped joints. Site control zones shall be established with exclusion, contaminant reduction (decontamination), and support zones in accordance with published Environmental Protection Agency (EPA) and California Department of Occupational Safety and Health (Cal/OSHA) guidelines. BioMax would be happy in providing the selected contractor with further site-specific detail regarding PPE regimen and appropriate site control zones, as necessary.
 8. BioMax recommends that all interior items and/or furnishings located within the break room remain in place for cleaning by the mitigation contractor while in containment. All such materials and furnishings currently present within the Break Room area shall remain in place for appropriate disposal and/or decontamination at the option of the tenant and DGS. All hard surface furnishings within the break room deemed shall receive a thorough inspection, cleaning, mildicide wet-wiping, and HEPA vacuuming as part of these recommended procedures prior to subsequent clearance testing and reuse.
 9. BioMax specifically recommends that all damaged sink cabinet materials and impacted sheetrock and wallboard underlayment materials be inspected and removed. As verified through visual inspection, any stained and/or moisture/mold affected interior sheetrock and building materials should be removed, wherever feasible, to the extent of visible staining, at a minimum. Floor tile materials shall also be removed and disposed under containment controls for appropriate inspection of subflooring. Removal of moisture impacted and mold damaged materials may also employ the use of appropriate item-specific containment methods and systems (such as sealed plastic glove-bag containment systems, or equivalent) applicable to the materials being removed at the direction of the Project CIH. BioMax currently anticipates that all visually affected floor mounted cabinets, sheetrock underlayment, and floor covering materials present within the break room shall be removed for disposal, and physical inspection of wall cavities and underlayment, as necessary. Any underlayment materials exhibiting visible signs of moisture staining shall also be removed or decontaminated, as necessary.
 10. Other potentially affected areas and building materials encountered during these deconstructive and investigative stages, such as adjacent walls and building material framing, underlayment, etc., must be thoroughly inspected during these deconstructive stages to identify the extent of any additional microbial related materials and water damage indicators. In general, all microbial impacted materials shall be removed to the extent of visible staining and at least 2 feet beyond such identified perimeters, wherever possible.

11. All remaining moisture/mold affected porous and non-porous building materials deemed infeasible for removal and/or disposal (due to structural integrity concerns) shall be inspected and receive a series of decontamination treatment measures designed to minimize and control the presence of microbial related substances. Decontamination methods employed shall, at a minimum, include treatment of all identified surfaces with a series of thorough chlorine based mildicide (minimum 10 parts water to 1 part chlorine soln.) applications followed by a series of thorough HEPA filtered vacuuming procedures using power sanding and/or brush agitation. The duration and frequency of mildicide and HEPA sanding/brushing applications employed may vary depending on local material contamination but shall be sufficient in removing and decontaminating all visible surface staining to levels deemed by BioMax to be consistent with representative background levels. Reasonable additional mitigative measures and controls may be required, as necessary, upon discovery of additional contaminated materials as well as BioMax's site inspection findings and observations performed during this scope of work. BioMax will be available to provide ongoing consultation with the contractor pertaining to these measures and site/material specific decontamination measures upon request.
12. Upon completion of mitigation efforts performed by the selected mitigation contractor, BioMax recommends the performance of a visual inspection conducted by the Project Certified Industrial Hygienist (Project CIH) to verify that all significant mold related staining and moisture indicators have been removed and/or treated and that all prescribed mitigative efforts and measures have been appropriately achieved. Once established, the Project CIH will collect a series of microbial "clearance" air samples to verify that all affected interior areas have been appropriately decontaminated to acceptable background airborne levels and that the affected areas within the subject building are verified as "cleared" for reconstruction, forthcoming reoccupancy, and reuse. Such Post Mitigative "clearance" evaluation criteria have been developed in BioMax's February 15th, 2008 letter report titled Post Mitigation Clearance Assessment Protocols and previously approved by HygieneTech, Inc. (HTI) in their approval letter dated February 22nd, 2008. Additional "punch-list" action items may be provided to the contractor following the performance of this site clearance inspection prior to receipt of analytical results, as deemed necessary.
13. Upon review of analytical sampling results by the Project CIH and achievement of acceptable post mitigative clearance criteria, BioMax recommends that DGS directs the mitigation/reconstruction contractor apply a mildicide-based sealant onto all remaining organic-based building materials and previously treated surfaces. Use of a recognized commercially available sealant product with microbial growth inhibitors in accordance with manufacturer's application and use instructions is believed to be currently acceptable for these purposes. The provision of appropriate access shall be provided to BOE and its consultants for inspection of affected areas and materials prior to final encapsulation and reconstruction upon request.
14. Following the performance of these mitigative measures, the designated site reconstruction contractor is strongly encouraged to verify that repairs to any faulty and/or deficient building penetration, drainage, plumbing and/or building envelop sealing systems have been

appropriately inspected, replaced/repaired, and function tested prior to the reconstruction of the affected interior structures and cavities. Certainly, the repair/replacement and/or establishment of any such additional engineering controls (as recommended through additional professional consultation) must be performed and implemented in accordance with applicable standards, building codes, and ordinances, as necessary.

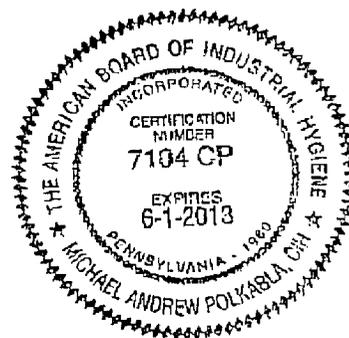
15. Upon completion, reconstruction of interior structural materials should be undertaken utilizing visibly clean (hand selected) construction grade materials in accordance with applicable building codes and requirements. The reconstruction contractor shall be required to only select materials which are obtained from reputable commercial sources and which are believed and visually verified to be free from elevated microbial contamination and/or elevated moisture content. New building materials, which are notably moist and/or visibly stained, shall NOT be used during the reconstruction of the subject structure. BioMax specifically recommends that reconstruction materials selected for use in the break room areas be specifically selected based on their moisture deterrent and anti-microbial properties wherever feasible.

16. Reasonable additional assessment and mitigative measures may also be required upon the identification of new or previously undiscovered materials and/or information related to moisture/microbial impacts, as necessary. Any reoccurrence of moisture intrusion following reconstruction should certainly be reviewed and addressed through further professional consultation, as necessary. BioMax would be happy to provide additional microbial consultative services pertaining to the mitigation of such structures so as to minimize potential adverse impacts to the interior working environment during the performance of any such activities upon request..

Once again, it has been a pleasure working with DGS on these important matters. If you have any additional questions, comments, or require further assistance, please do not hesitate to contact me directly at (510) 724-3100.

Sincerely,

Michael A. Polkabila, CIH, REA
Vice President, Principal



LIMITATIONS

Please note that the professional opinions presented in this review are intended for the sole use of DGS and their designated beneficiaries. No other party should rely on the information contained herein without the prior written consent of BioMax Environmental and DGS. The professional opinions provided herein are based on BioMax's review and understanding of current site information and observed site conditions present within the areas inspected at the time these services were performed. Professional recommendations provided as part of this limited scope of work are intended for client consideration only and are not intended as a professional or regulatory mandate. Implementation of any of the above measures or recommendations does not, in any way, warrant the day-to-day health and/or safety of building occupants, residents, site workers, nor regulatory or building code compliance status during normal and changing environmental conditions. As microbial contamination, by nature, may change over time due to additional moisture intrusion, favorable growth conditions, and changing environments, the findings of this report are subject to change in the event that such conditions and/or environments arise. Also, the professional opinions expressed here are subject to revision in the event that new or previously undiscovered information is obtained or uncovered.

The information contained in this and any other applicable report communication is intended for consideration purposes only. It is not intended, nor should it be construed as providing legal advice or warranting any level of safety or regulatory compliance. The sole purpose of such information is to assist with the identification, evaluation and control of potential contamination or unnecessary physical, chemical, and/or biological hazards. Any action taken based on this information, including but not limited to opinions, suggestions and recommendations, whether implied or expressed, is the sole responsibility of the individual taking the action. Risk management and safety is criteria dependent and situation specific requiring extensive knowledge and value assessments to be properly determined by competent professionals.

These services were performed by BioMax in accordance with generally accepted professional industrial hygiene principals, practices, and standards of care. Under the existing Industrial Hygiene Definition and Registration Act, all reports, opinions or official documents prepared by a Certified Industrial Hygienist (CIH) constitutes an expression of professional opinion regarding those facts or findings which are subject of a certification and does not constitute a warranty or guarantee, either expressed or implied.

**EMLab P&K**

Report for:

Mr. Michael Polkabila
Biomax Environmental
775 San Pablo Ave.
Pinole, CA 94564

Regarding: Project: 051308-01; 450 N. Street, 20th Floor, Breakroom 2014
EML ID: 422036

Approved by:

Lab Manager
Dr. Kamashwaran Ramanathan

Dates of Analysis:
Quantitative spore count direct exam: 05-14-2008
Spore trap analysis: 05-14-2008

Project SOPs: Quantitative spore count direct exam (1100008), Spore trap analysis (1100000)

This coversheet is included with your report in order to comply with AIHA and ISO accreditation requirements.

For clarity, we report the number of significant digits as calculated; but, due to the nature of this type of biological data, the number of significant digits that is used for interpretation should generally be one or two. All samples were received in acceptable condition unless noted in the Report Comments portion in the body of the report. Due to the nature of the analyses performed, field blank corrections of results is not a standard practice. The results relate only to the items tested.

EMLab P&K ("the Company") shall have no liability to the client or the client's customer with respect to decisions or recommendations made, actions taken or courses of conduct implemented by either the client or the client's customer as a result of or based upon the Test Results. In no event shall the Company be liable to the client with respect to the Test Results except for the Company's own willful misconduct or gross negligence nor shall the Company be liable for incidental or consequential damages or lost profits or revenues to the fullest extent such liability may be disclaimed by law, even if the Company has been advised of the possibility of such damages, lost profits or lost revenues. In no event shall the Company's liability with respect to the Test Results exceed the amount paid to the Company by the client therefor.

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EMLab P&K

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(650) 829-5800 Fax (650) 829-5852 www.emlab.com

Client: Biomax Environmental
C/O: Mr. Michael Polkabla
Re: 051308-01; 450 N. Street, 20th Floor, Breakroom
2014

Date of Sampling: 05-13-2008
Date of Receipt: 05-14-2008
Date of Report: 05-14-2008

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	13811704: Room 2014, breakroom		13811714: Area to SW at breakroom		13810632: Cube 078		13811941: Room 2014 with cabinets open	
Comments (see below)	None		None		None		None	
Lab ID-Version†:	1855566-1		1855567-1		1855568-1		1855569-1	
	raw ct	spores/m3	raw ct	spores/m3	raw ct	spores/m3	raw ct	spores/m3
Alternaria								
Arthrinium								
Ascospores*							1	53
Aureobasidium								
Basidiospores*	1	53	1	53			1	53
Bipolaris/Drechslera group								
Botrytis								
Chaetomium								
Cladosporium								
Curvularia								
Epicoccum								
Fusarium								
Myrothecium								
Nigrospora								
Oidium								
Other brown					1	13		
Penicillium/Aspergillus types†								
Pithomyces								
Rusts*			1	13			1	13
Smuts*, Periconia, Myxomycetes*	1	13	2	27				
Stachybotrys								
Stemphylium								
Torula								
Ulocladium								
Zygomycetes								
Background debris (1-4+)††	2+		1+		1+		1+	
Hyphal fragments/m3	13		<13		<13		<13	
Pollen/m3	13		<13		<13		13	
Skin cells (1-4+)	1+		1+		1+		1+	
Sample volume (liters)	75		75		75		75	
TOTAL SPORE/m3		66		93		13		119

Comments:

* Most of these spore types are not seen with culturable methods (Andersen sampling), although some may appear as non-sporulating fungi. Most of the basidiospores are "mushroom" spores while the rusts and smuts are plant pathogens.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Poecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

†† Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for sample volumes when evaluating dust levels.

The Limit of Detection is the product of a raw count of 1 and 100 divided by the percent read. The analytical sensitivity (counts/m3) is the product of the Limit of Detection and 1000 divided by the sample volume.

‡ A "Version" greater than 1 indicates amended data.

EMLab P&K

1150 Bayhill Drive, Suite 100, San Bruno, CA 94066
(650) 829-5800 Fax (650) 829-5852 www.emlab.com

Client: Biomax Environmental
C/O: Mr. Michael Polkaba
Re: 051308-01; 450 N. Street, 20th Floor, Breakroom
2014

Date of Sampling: 05-13-2008
Date of Receipt: 05-14-2008
Date of Report: 05-14-2008

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	13811760: Room 2013 scanning room		13811685: M floor balcony SW		13810584: Front entry area NE	
Comments (see below)	None		None		None	
Lab ID-Version†:	1855570-1		1855571-1		1855572-1	
	raw ct.	spores/m3	raw ct.	spores/m3	raw ct.	spores/m3
Alternaria						
Arthrospora						
Ascospores*					1	53
Aureobasidium						
Basidiospores*			2	107	3	160
Bipolaris/Drechslera group						
Botrytis						
Chaetomium						
Cladosporium					3	160
Curvularia						
Epicoccum						
Fusarium						
Myrothecium						
Nigrospora						
Oidium					1	13
Other brown	1	13	1	13	1	13
Other colorless						
Penicillium/Aspergillus types†						
Pithomyces						
Rusts*					2	27
Smuts*, Periconia, Myxomycetes*			1	13	3	40
Stachybotrys						
Stemphylium			1	13		
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)††	1+		1+		1+	
Hyphal fragments/m3	< 13		< 13		40	
Pollen/m3	< 13		< 13		133	
Skin cells (1-4+)	1+		1+		< 1+	
Sample volume (liters)	75		75		75	
TOTAL SPORE/m3		13		146		466

Comments:

* Most of these spore types are not seen with culturable methods (Andersen sampling), although some may appear as non-sporulating fungi. Most of the basidiospores are "mushroom" spores while the rusts and smuts are plant pathogens.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paeecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

†† Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting viability for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.

The Limit of Detection is the product of a raw count of 1 and 100 divided by the percent read. The analytical sensitivity (counts/m3) is the product of the Limit of Detection and 1000 divided by the sample volume.

‡ A "Version" greater than 1 indicates amended data.

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Client: Biomax Environmental
C/O: Mr. Michael Polkabila
Re: 051308-01; 450 N. Street, 20th Floor, Breakroom
2014

Date of Sampling: 05-13-2008

Date of Receipt: 05-14-2008

Date of Report: 05-14-2008

MoldRANGE™: Extended Outdoor Comparison**Outdoor Location: 13810584, Front entry area NE**

Fungi Identified	Outdoor data	Typical Outdoor Data by Date†				Typical Outdoor Data by Location‡			
		Month: May				State: CA			
	spores/m3	low	med	high	freq %	low	med	high	freq %
Generally able to grow indoors*									
Alternaria	-	7	33	320	62	7	27	230	60
Bipolaris/Drechslera group	-	7	13	120	16	7	13	120	14
Chaetomium	-	7	13	100	15	7	13	110	19
Cladosporium	160	53	530	6,900	96	53	640	6,400	98
Curvularia	-	7	13	360	8	7	13	210	7
Nigrospora	-	7	13	120	7	7	13	170	8
Other brown	13	7	13	80	36	7	13	80	37
Penicillium/Aspergillus types	-	27	160	1,700	80	40	210	2,500	88
Stachybotrys	-	7	13	230	4	7	13	300	5
Torula	-	7	13	170	15	7	13	150	13
Seldom found growing indoors**									
Ascospores	53	13	160	5,300	81	13	110	1,800	72
Basidiospores	160	13	270	6,900	93	13	240	6,700	94
Oidium	13	7	22	250	27	7	13	190	20
Rusts	27	7	25	330	29	7	13	270	29
Smuts, Periconia, Myxomycetes	40	7	53	1,000	76	8	40	470	71
TOTAL SPORES/M3	466								

† The Typical Outdoor Data by Date represents the typical outdoor spore levels across North America for the month indicated. The last column represents the frequency of occurrence. The low, medium, and high values represent the 2.5, 50, and 97.5 percentile values of the spore type when it is detected. For example, if the frequency of occurrence is 63% and the low value is 53, it would mean that the given spore type is detected 63% of the time and, when detected, 2.5% of the time it is present in levels above the detection limit and below 53 spores/m³. These values are updated periodically, and if enough data is not available to make a statistically meaningful assessment, it is indicated with a dash.

‡ The Typical Outdoor Data by Location represents the typical outdoor spore levels for the region indicated for the entire year. As with the Typical Outdoor Data by Date, the four columns represent the frequency of occurrence and the typical low, medium, and high concentration values for the spore type indicated. These values are updated periodically, and if enough data is not available to make a statistically meaningful assessment, it is indicated with a dash.

*The spores in this category are generally capable of growing on wet building materials in addition to growing outdoors. Building related growth is dependent upon the fungal type, moisture level, type of material, and other factors. *Cladosporium* is one of the predominant spore types worldwide and is frequently present in high numbers. *Penicillium/Aspergillus* species colonize both outdoor and indoor wet surfaces rapidly and are very easily dispersed. Other genera are usually present in lesser numbers.

**These fungi are generally not found growing on wet building materials. For example, the rusts and smuts are obligate plant pathogens. However, in each group there are notable exceptions. For example, agents of wood decay are members of the basidiomycetes and high counts of a single morphological type of basidiospore on an inside sample should be considered significant.

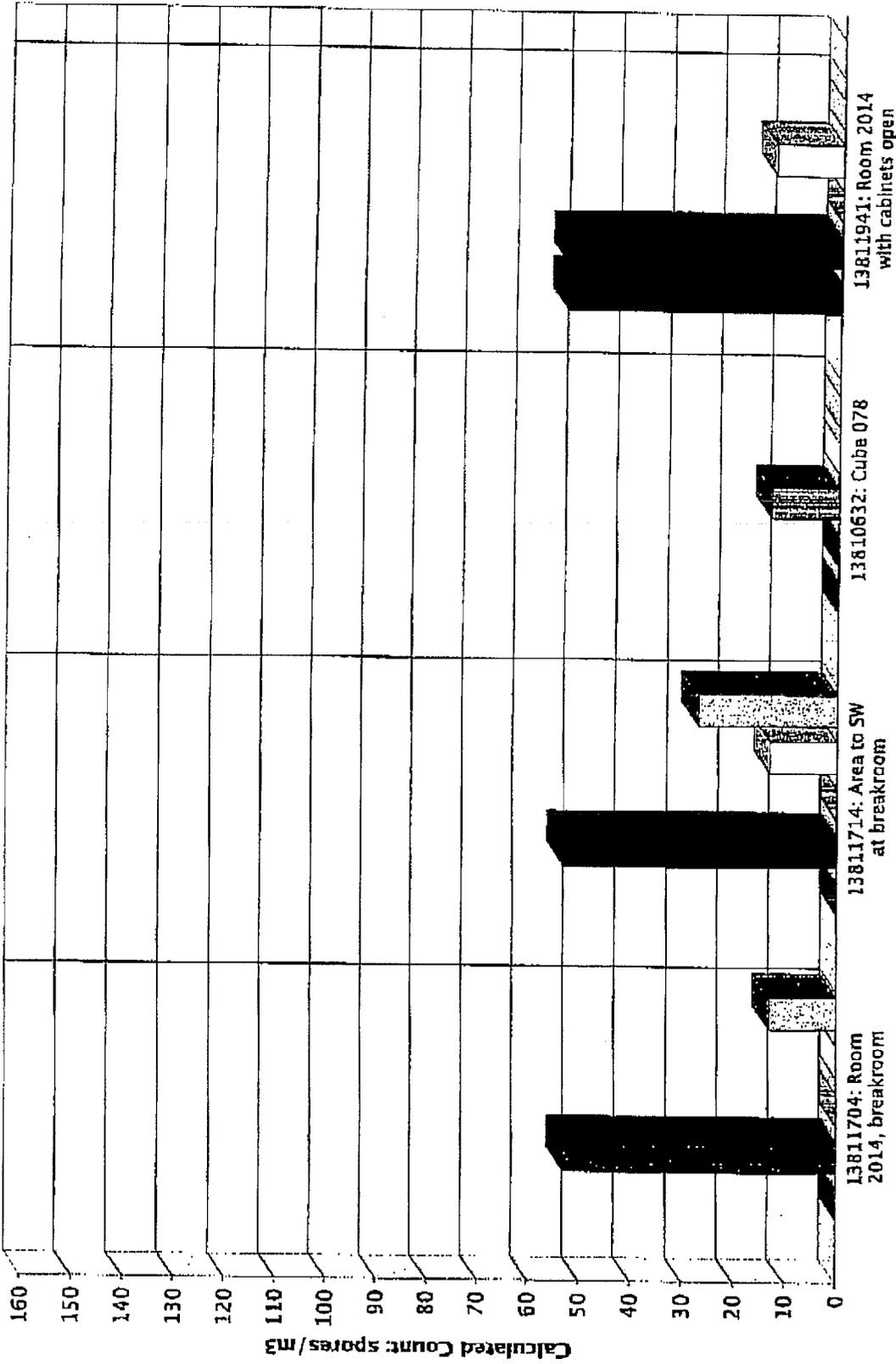
Interpretation of the data contained in this report is left to the client or the persons who conducted the field work. This report is provided for informational and comparative purposes only and should not be relied upon for any other purpose. "Typical outdoor data" are based on the results of the analysis of samples delivered to and analyzed by EMLab P&K and assumptions regarding the origins of those samples. Sampling techniques, contaminants infecting samples, unrepresentative samples and other similar or dissimilar factors may affect these results. In addition, EMLab P&K may not have received and tested a representative number of samples for every region or time period. EMLab P&K hereby disclaims any liability for any and all direct, indirect, punitive, incidental, special or consequential damages arising out of the use or interpretation of the data contained in, or any actions taken or omitted in reliance upon, this report.

05-14-2008: 051308-01

EMLab P&K
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(650) 829-3800 Fax (650) 829-5852 www.emlab.com

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

■ Ascospores ■ Basidiospores ▨ Other brown □ Rusts □ Smuts, Periconia, Myxomycetes

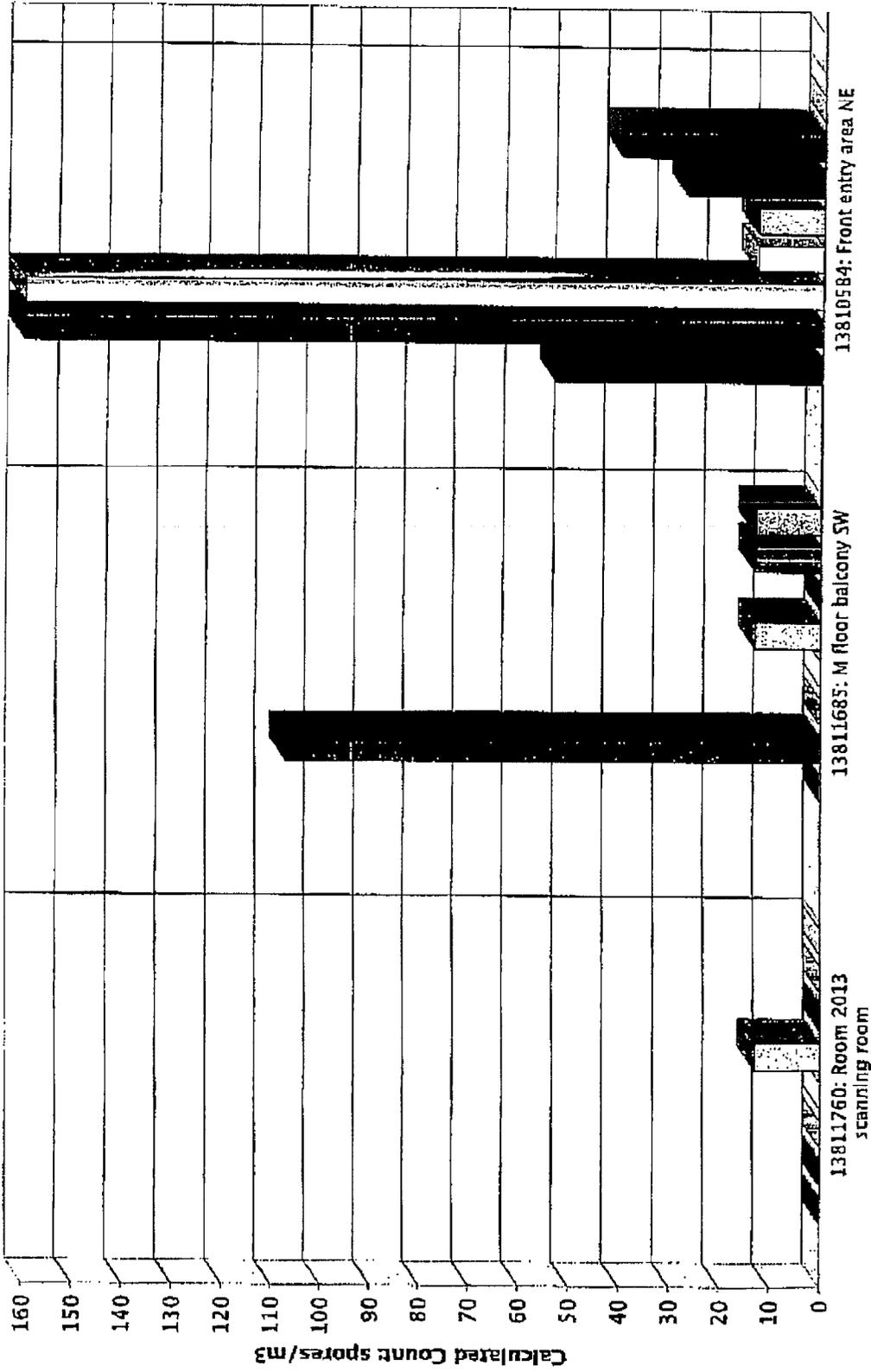


Comments:

Note: Graphical output may understate the importance of certain "marker" genera.

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

- Ascospores ■ Basidiospores ■ Cladosporium □ Oidium □ Other brown ■ Rusts ■ Smuts, Periconia, Myxomycetes
- Stemphylium



Comments:

Note: Graphical output may underestimate the importance of certain "marker" genera.

Client: Biomax Environmental
 C/O: Mr. Michael Polkabila
 Re: 051308-01; 450 N. Street, 20th Floor, Breakroom
 2014

EMLab P&K
 1150 Bayhill Drive, Suite 100, San Bruno, CA 94066
 (650) 829-5800 Fax (650) 829-5852 www.emlab.com

Date of Sampling: 05-13-2008
 Date of Receipt: 05-14-2008
 Date of Report: 05-14-2008

QUANTITATIVE SPORE COUNT REPORT

Location:	B01: Venir cabinet base surface fragment		B02: Cracked venir at rear of cabinet		S01: Cracked right cabinet surface, R		S02: Cracked right cabinet surface, L	
Comments (see below)	None		None		None		None	
Sample type	Bulk sample		Bulk sample		Tape sample		Tape sample	
Lab ID-Version†:	1855559-1		1855560-1		1855561-1		1855562-1	
	raw ct.	spores/unit	raw ct.	spores/unit	raw ct.	spores/unit	raw ct.	spores/unit
Alternaria								
Arthrinium								
Ascospores*								
Aureobasidium								
Basidiospores*								
Bipolaris/Drechslera group								
Botrytis								
Chaetomium							11	13
Cladosporium					1	0.12		
Curvularia								
Epicoccum								
Fusarium								
Myrothecium								
Nigrospora								
Other colorless								
Penicillium/Aspergillus types†	8	0.93	41	4.8				
Phthomyces								
Rusts*					1	0.12		
Smuts*, Periconia, Myxomycetes*							1	0.12
Stachybotrys								
Stemphylium								
Torula								
Ulocladium								
Zygomycetes								
Background debris (1-4+)††	N/A		N/A		2+		2+	
Sample size	100		100		100		100	
Unit	1 mm ²		1 mm ²		1 mm ²		1 mm ²	
TOTAL SPORES/UNIT		0.93		4.8		0.24		1.42

Comments:

* Most of these spore types are not seen with culturable methods (Anderson sampling), although some may appear as nonsporulating colonies. Most of the basidiospores are 'mushroom' spores while the rusts and smuts are plant pathogens.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Faecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

†† Background debris is an indication of the amount of non-biological particulate matter present on the slide (dust in the air) and is graded from 1+ to 4+ with 4+ indicating the largest amounts. This background material is also an indication of visibility for the analyst and resultant difficulty reading the slide. For example, high background debris may obscure the small spores such as the *Penicillium/Aspergillus* group. Counts from areas with 4+ background debris should be regarded as minimal counts and may actually be higher than reported.

‡ A "Version" greater than 1 indicates amended data.

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Client: Biomax Environmental
C/O: Mr. Michael Polkabla
Re: 051308-01; 450 N. Street, 20th Floor, Breakroom
2014

Date of Sampling: 05-13-2008
Date of Receipt: 05-14-2008
Date of Report: 05-14-2008

QUANTITATIVE SPORE COUNT REPORT

Location:	S03: Metal cabinet		S04: Cabinet top surface		S05: Adjacent cubicle divider top	
Comments (see below)	None		None		None	
Sample type	Tape sample		Tape sample		Tape sample	
Lab ID-Version†:	1855563-1		1855564-1		1855565-1	
	raw ct.	spores/unit	raw ct.	spores/unit	raw ct.	spores/unit
Alternaria						
Arthrimum						
Ascospores*						
Aureobasidium						
Basidiospores*						
Bipolaris/Drechslera group						
Botrytis						
Chaetomium						
Cladosporium						
Curvularia						
Epicoccum						
Fusarium						
Myrothecium						
Nigrospora						
Other colorless						
Penicillium/Aspergillus types†						
Pithomyces						
Rusts*						
Smuts*, Periconia, Myxomycetes*			1	0.12		
Stachybotrys						
Stemphylium						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)††	1+		3+		< 1+	
Sample size	100		100		100	
Unit	1 mm ²		1 mm ²		1 mm ²	
TOTAL SPORES/UNIT		< 0.01		0.12		< 0.01

Comments:

* Most of these spore types are not seen with culturable methods (Andersen sampling), although some may appear as nonsporulating colonies. Most of the basidiospores are 'mushroom' spores while the rusts and smuts are plant pathogens.

† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium*, *Paeclomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.

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‡ A "Version" greater than 1 indicates amended data.

BULK / SURFACE SAMPLING RECORD BIOMAX ENVIRONMENTAL, LLC

775 San Pablo Avenue
Pinole, CA 94564

Phone (510) 724-3100 Fax (510) 724-31435 biomaxenv@aol.com

000422036



Project Name and Location: 2014 Break Room BOE Building
Client: DGS 450 N Street
Sacramento, CA

Analysis Requested: Fungal ID

Analytical Laboratory: EM Labs Date of Sampling: 5/13/08

Required Turn Around Time: Same Day Sampled By: [Signature]

Sample ID Number	Sample Type (Bulk/Surface)	Area/Volume Sampled	Location/Description
S01	Surf	1x1"	Cracked cabinet surface (R)
S02	Surf	1x1"	Cracked Right cabinet surface (L)
S03	Surf	1x1"	Metal Cabinet
S04	Surf	1x1"	Cabinet Top Surface
S05	Surf	1x1"	Adjacent Cubicle Divider Top
B01	Bulk	1x2.5"	venir cabinet base surface fragment
B02	Bulk	1x2.5"	cracked venir @ Rear of cabinet

Instructions and Comments: Same Day Rush

Please sign this form below acknowledging sample receipt and return executed form with laboratory reports. Fax, send and e-mail results to BioMax Environmental at (510) 724-3145 biomaxenv@aol.com

Relinquished by: <u>[Signature]</u>	Received By: <u>Anna Marinsky</u>
Method of Transportation: <u>Fed Ex</u>	Time/Date Received: <u>5-14-08 9:30</u>
Time/Date Sent: <u>4:30 5/12/08</u>	

MICROBIAL SPORE TRAP AIR SAMPLING RECORD



000422036

Page 1 of 1

BioMax Environmental
775 San Pablo Ave.
Pinole, CA 94564

www.biomaxenvironmental.com

Phone: (510) 724-3100

Fax: (510) 724-3145

biomaxenv@aol.com

Location: 450 N Street 20th Floor Break Room (2014)	Client: DGS
Date: 5/13/08	Project #: OS1308-01
Collected by: M. A. Polkoba, CIH	Laboratory: EM Labs
Signature: <i>M. A. Polkoba</i>	Req. Turn Around: Same Day
	Analysis: <u>Fungal</u> <u>Particulate(D)</u> with <u>Quantification.</u>

Sample Number	Time	Location	Temp/Hum
A 13811704	1052	Rm 2014 : Break Room	73 / 29%
B 13811714	11:00	Area to SW of Break Rm	73 / 27%
C 13810652	1110	Cube OTR	73 / 28%
D 13811941	1115	Rm 2014 w/ Cabinets open	75 / 29%
E 13811760	1125	Rm 2013 Scanning Rm	72 / 28%
F 13811685	1140	M Floor Balcony SW	76 / 35%
G 13810584	1230	Front Entry Area NE	77 / 38%
Total Sample Time (min): 5 min	Flow Rate (l/min): 15 l/min	Total Sample Volume (liters): 75 L	Ambient Conditions: Clear / Mild
Comments:			

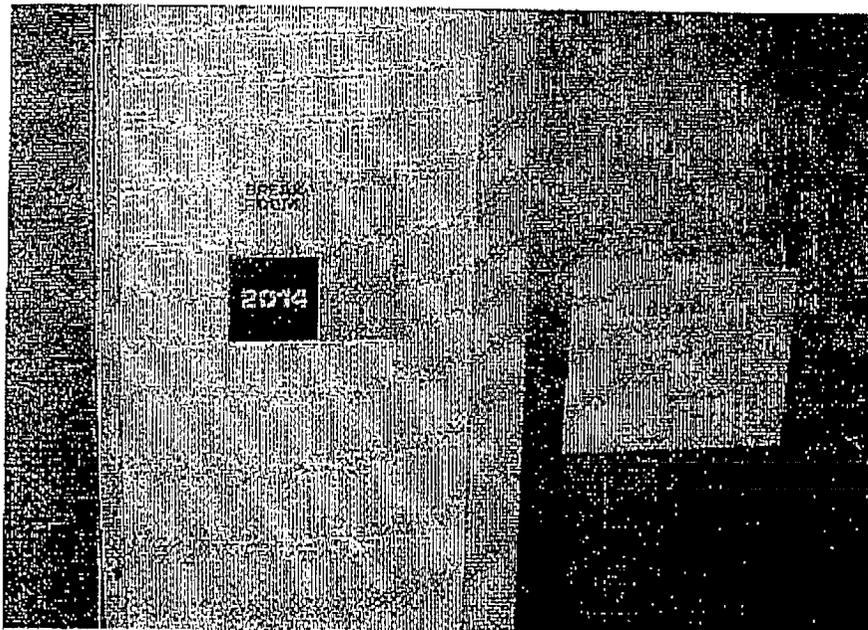
Please sign this form below acknowledging sample receipt and return executed form with laboratory reports. Fax, send, e-mail results to BioMax Environmental at (510) 724-3145 biomaxenv@aol.com
Other Instructions: Same Day Rush

Relinquished by: <i>M. A. Polkoba</i>	Received By: <i>Ann Morrissey</i>
Method of Transportation: Fed Ex Priority	Time/Date Received: 5-14-08 9:30
Time/Date Sent: 5/13/08 @ 4:45 pm	

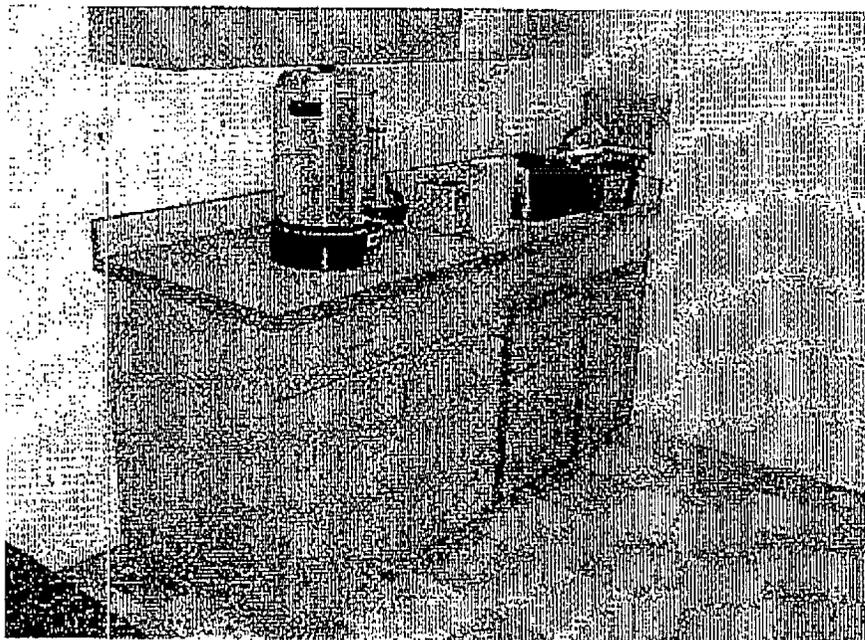
Attachment A: Digital Images
 May 13th, 2008
 BOE Building 20th Floor Break Room
 Sacramento, CA

Page 1 of 7

[Click here for color photos](#)



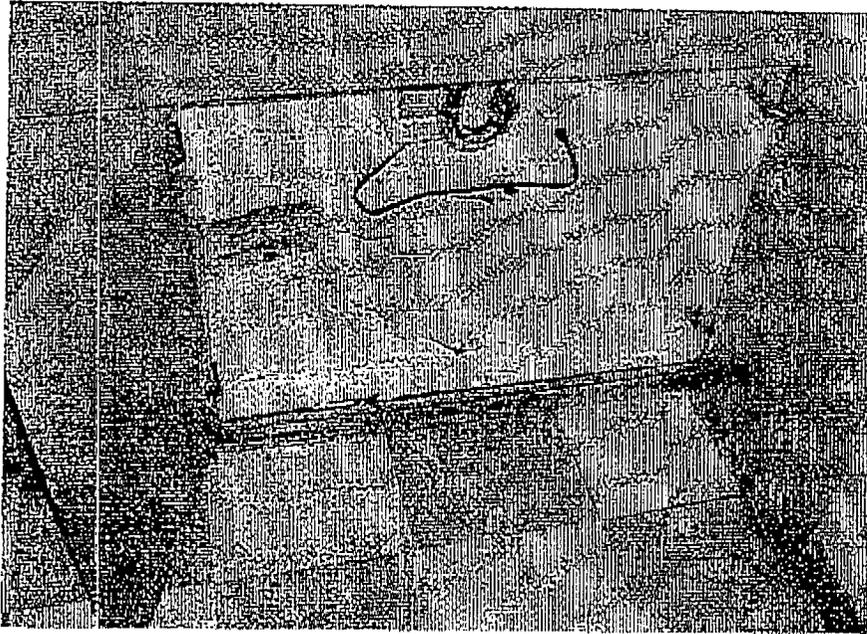
1) Image of entry to 20th Floor Break Room (2014) with BOE generated posting present on doorway at the time of BioMax's assessment.



2) Image of Break Room cabinet at time of preliminary inspection. Blue tape present on impacted cabinet material as applied by BOE staff prior to BioMax's inspection.

May 13th, 2008
BOE Building 20th Floor Break Room
Sacramento, CA

Page 2 of 7



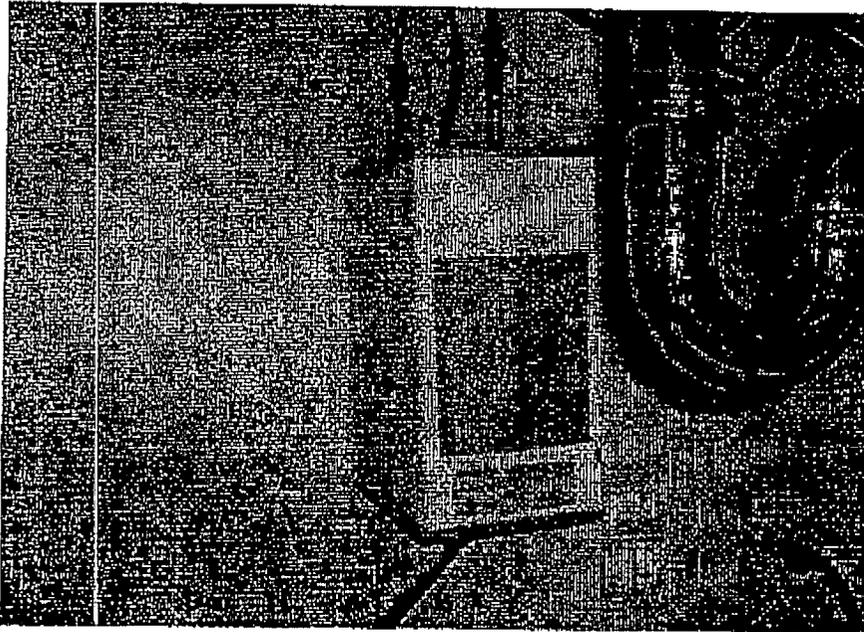
- 3) Image within cabinetry of Break Room upon removal of tape barrier. Note cracking and staining present on laminant cabinet base materials.



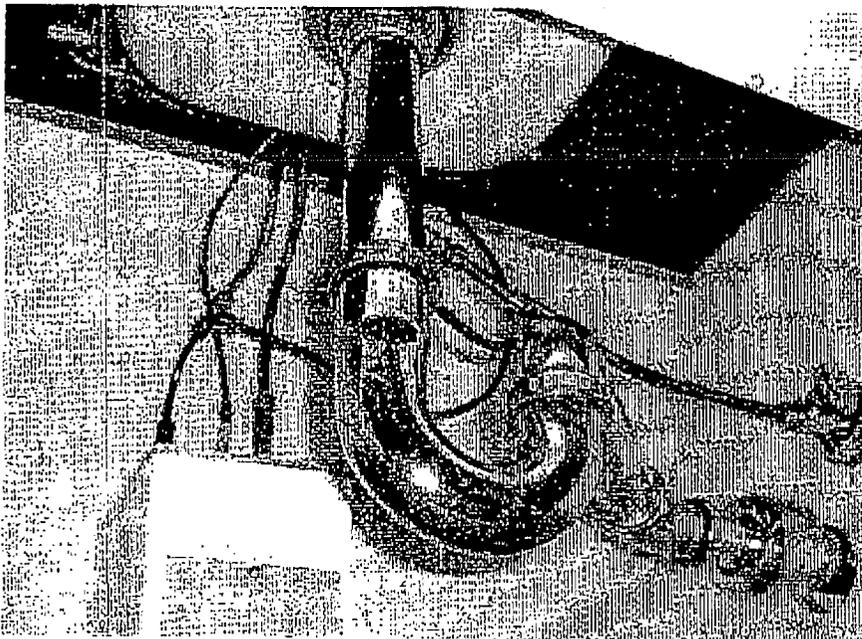
- 4) Close-up image of cracking and delamination within cabinet base materials. Note adjacent vinyl baseboard and tile flooring material delamination at time of assessment.

May 13th, 2008
BOE Building 20th Floor Break Room
Sacramento, CA

Page 3 of 7



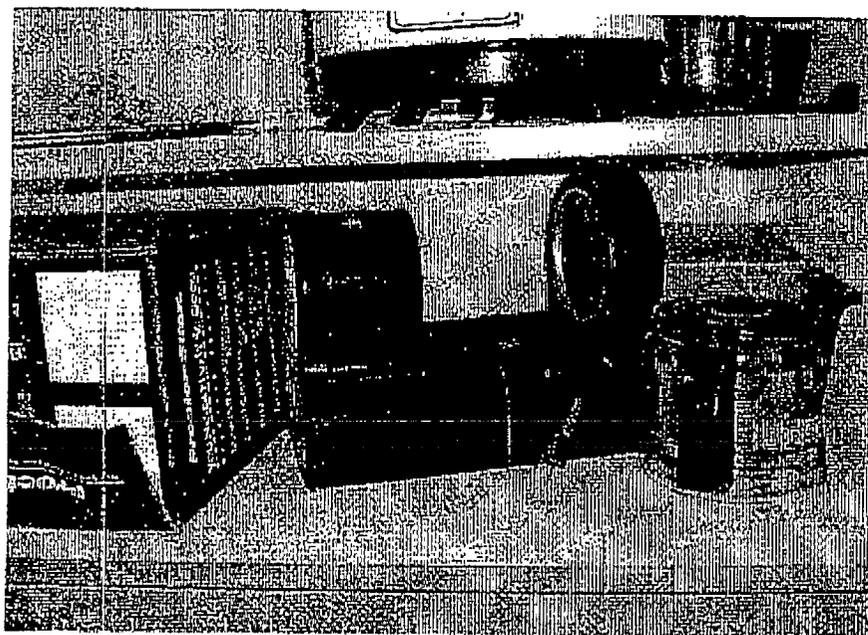
- 5) Image within sink cabinet of "Insta-hot" water treatment unit with staining along the rear vertical wall emanating from behind the noted unit.



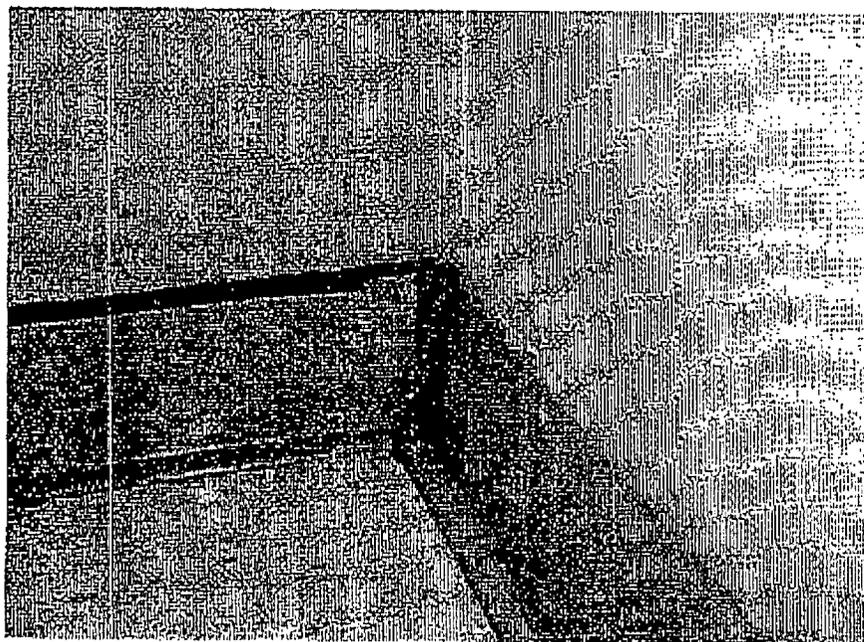
- 6) Image of sink insert base and plumbing within cabinet base. Evidence of historical leaking at base of "S" trap evident at time of assessment.

May 13th, 2008
BOE Building 20th Floor Break Room
Sacramento, CA

Page 4 of 7



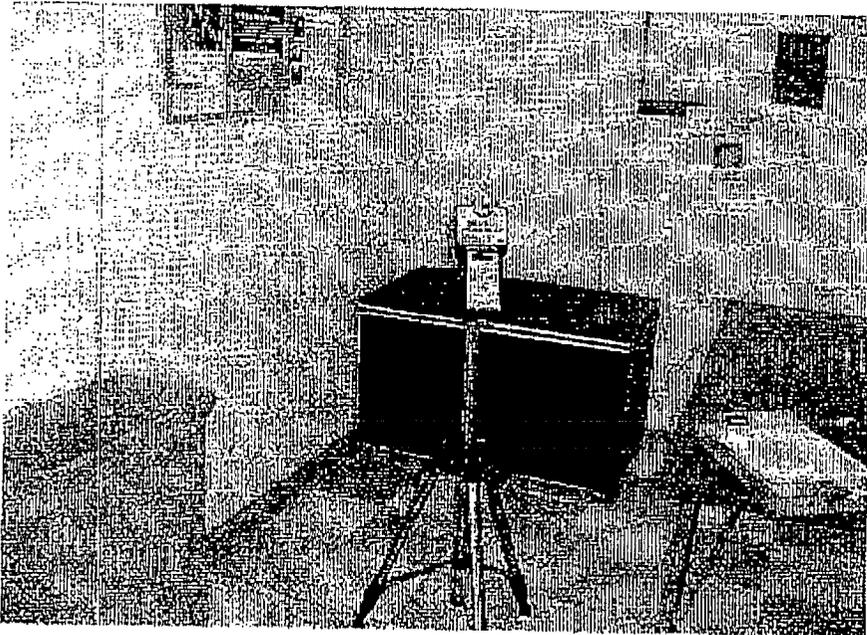
7) Close-up image of Personal appliances storage within adjacent cabinet compartment at time of assessment.



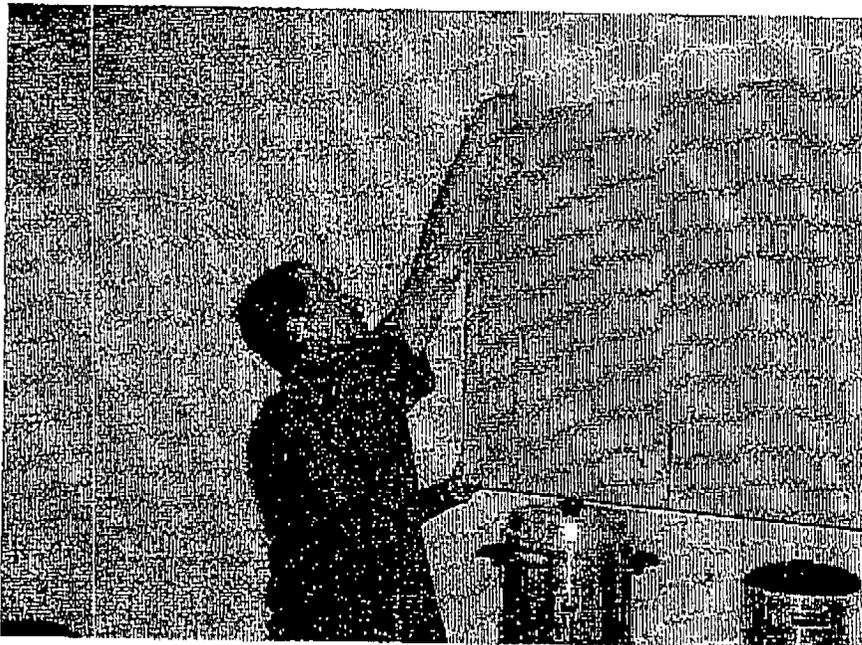
8) Image of right side of cabinet at floor level. Evidence of historical water staining present on cabinet door, vinyl baseboard, and floor tile materials.

May 13th, 2008
BOE Building 20th Floor Break Room
Sacramento, CA

Page 5 of 7



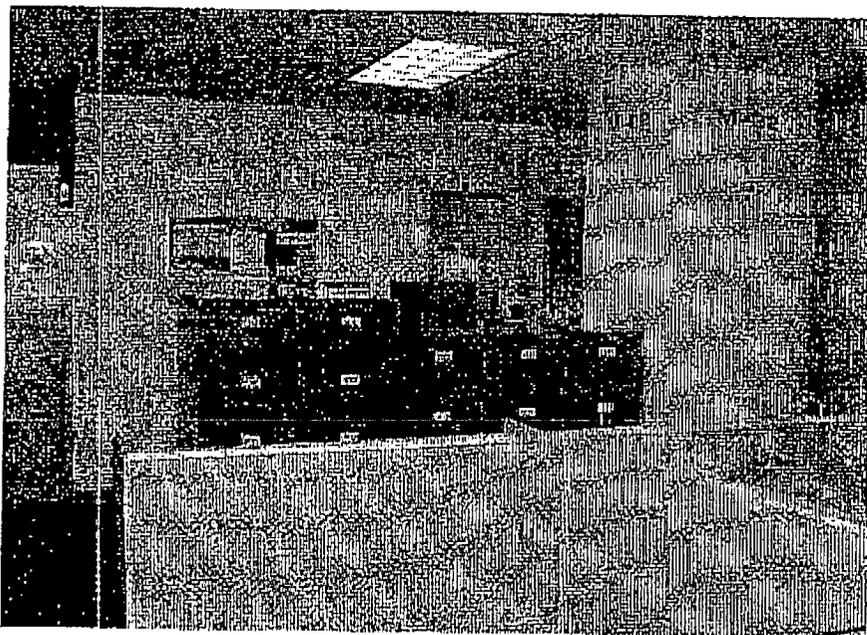
- 9) Image of air handling equipment utilized within break room and reference locations of existing furnishings where no evidence of moisture damages were noted at this time.



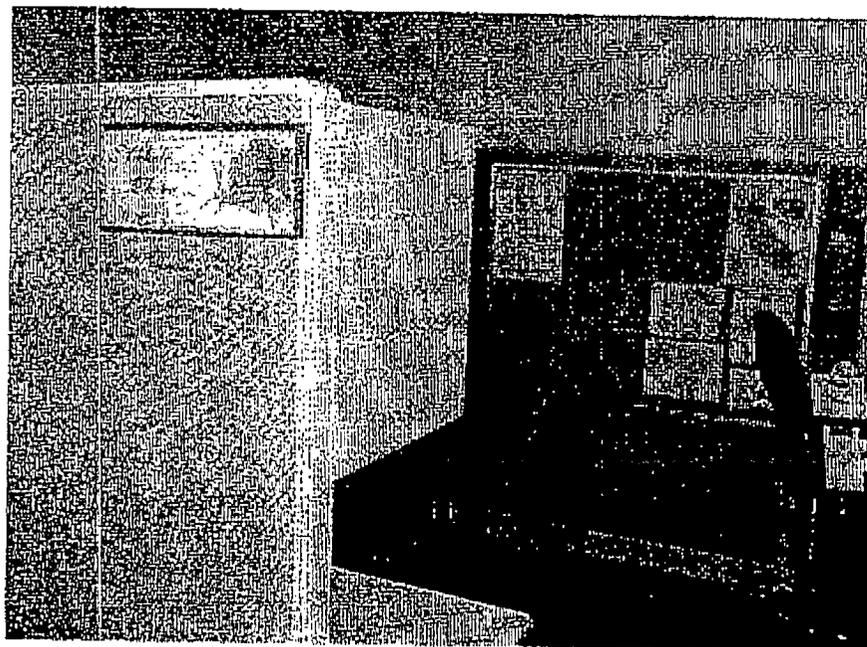
- 10) Image of Mr. Polkabla of BioMax collecting surface sample from representative material within break room not directly affected by moisture contact/release.

May 13th, 2008
BOE Building 20th Floor Break Room
Sacramento, CA

Page 6 of 7



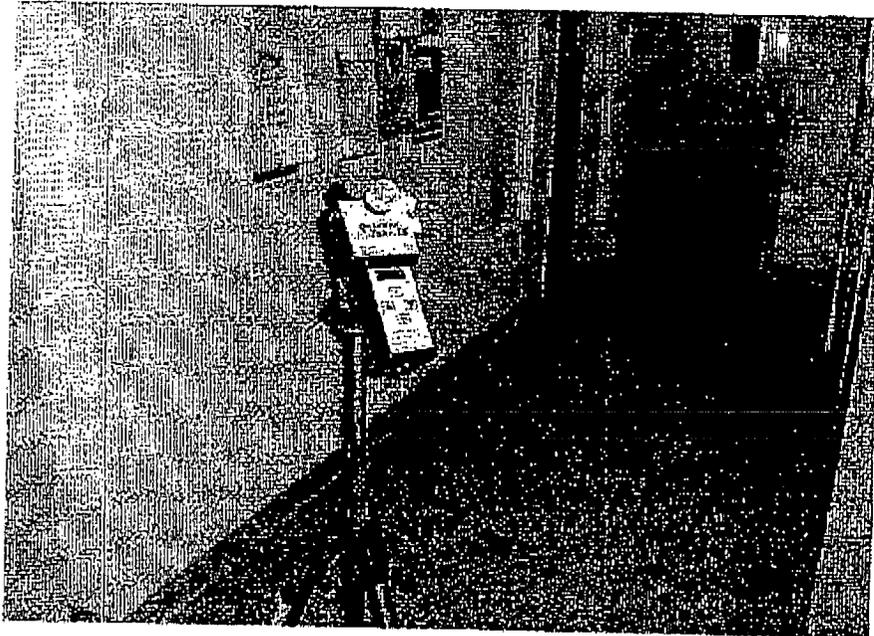
11) Image of occupied cubicle area adjacent to break room. Cubicle divider top frame in foreground served as a representative area for surface sampling and comparative analysis.



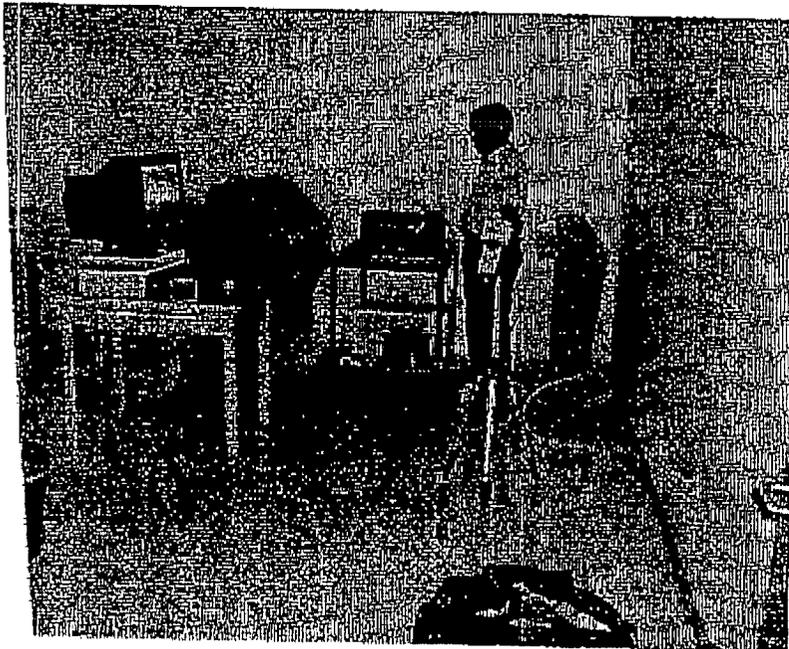
12) Image of air sampling activity performed within Nikkole Thom's cubicle area adjacent to (with shared rear wall) the break room.

May 13th, 2008
BOE Building 20th Floor Break Room
Sacramento, CA

Page 7 of 7



13) Image of air sampling activity performed within hallway of occupied cubicle area adjacent to break room.



14) Image of air sampling activity performed within MAP Scanning room (#2013) adjacent to (with shared rear wall) the noted break room.

